### **REVIEW**



# Behavioral effects of insect-resistant genetically modified crops on phytophagous and beneficial arthropods: a review

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**Abstract** Behavior is a main component of the survival and performance of arthropods. We have witnessed widespread adoption of insect-resistant genetically modified (IRGM) crops nowadays; however, no literature has reviewed the behavioral effects of IRGM crops on phytophagous and beneficial arthropods. In this review, we assessed the current information related to the effects of IRGM crops on arthropod behavior, mainly including locomotion (mobility, escape behavior and dispersal behavior), foraging (orientation, host plant selection/preference and feeding), mating, oviposition and other behaviors (associative learning). Almost all the studies have been conducted on Bt crops. The behavioral effects have been found in 54.2 %, 22 % and 33 % of the case studies on phytophagous arthropods, arthropod natural enemies and pollinators, respectively. Few behavioral studies have been documented on arthropod pollinators. The majority of cases reporting behavioral effects have derived from target phytophagous arthropods. Among them, locomotion and feeding behavior were the most frequently affected. For arthropod natural enemies, the cases using target prey/host in tri-trophic studies only accounted for a small proportion of behavioral effects observed on foraging behavior (host/ prey selection). Overall, the effects through tri-trophic pathways on behaviors of natural enemies are limited. To conclude, while attention needs to be paid to several behavioral effects that may undermine the efficacy of IRGM crops in sustainable pest management, the behavioral effects generally do not disrupt the role of IRGM crops in achieving the goal of integrated pest management and crop production.

**Keywords** Behavior · Parasitoids · Predators · Risk assessment · *Bacillus thuringiensis* (Bt) · IPM

#### Kev message

- No literature has reviewed the behavioral effect of insect-resistant genetically modified (IRGM) crops on phytophagous or beneficial arthropods in agroecosystems.
- In this review, we found that behavioral effects have been mostly evident on phytophagous arthropods, less on natural enemies and arthropod pollinators.
- Behavioral effects of IRGM crops on arthropods may not disrupt the role of such a technology in integrated pest management and crop production.

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### Introduction

Since the advent of genetically modified (GM) crops in the late 1970s, we have witnessed an unprecedented increase in the development and commercial use of this technology worldwide. The dominant GM traits nowadays include insect resistance, expressing *Bacillus thuringiensis* (Bt) and



digestive enzyme inhibitors, and herbicide tolerance, tolerating applications of particular herbicides (James 2014). IRGM crops that aim at controlling agricultural pests while reducing the use of insecticides have been extensively cultivated. They have been regarded as cost-efficient and environmentally sound (Ferry et al. 2006; Brookes and Barfoot 2006; Park et al. 2011). While IRGM crops are developed to act directly on the target phytophagous pests, the unintentional effects on non-target arthropods have been and continue to be a subject of much debate (Andow and Zwahlen 2006; Babendreier et al. 2006; Rose 2007; Romeis et al. 2006, 2008; Naranjo 2009; Desneux and Bernal 2010; Desneux et al. 2010; EFSA 2010; Romeis et al. 2011, 2012). Notably, arthropod natural enemies (the arthropod species from higher trophic levels) and arthropod pollinators have been the subject of risk assessment studies owing to their importance in agro-ecosystems (Duan et al. 2008: Lu et al. 2012).

Several review articles have discussed the effects of IRGM crops on non-target arthropods (O'Callaghan et al. 2005; Marvier et al. 2007; Romeis et al. 2008; Wolfenbarger et al. 2008; Duan et al. 2010), specifically on natural enemies (Lövei and Arpaia 2004; Romeis et al. 2006; Lövei et al. 2009; Lundgren et al. 2009) and arthropod pollinators (Duan et al. 2008). While these reviews have thoroughly discussed the potential effects of IRGM crops on non-target arthropods, they have not specifically addressed the behavioral effects of IRGM crops on either target or non-target arthropods. The importance of how IRGM crops affect arthropod behavior should not be underestimated for at least three reasons. First, like other animals, arthropod behaviors are susceptible to external (environmental variations) and internal factors (physiological requirements). Arthropods must behave in the right way to survive and develop (Wajnberg et al. 2007). Second, IRGM crops may interfere with behavioral patterns of phytophagous arthropods, natural enemies and arthropod pollinators, and a good knowledge of these effects may favor sustainable pest management (Lundgren et al. 2009). Taking phytophagous arthropods as examples, the behavior of caterpillars needs to be assessed when designing refuge strategies. Knowledge of the dispersal and movement behaviors of those caterpillars is required when designing so-called "refuge-in-the-bag" tactics that are valued to delay the development of pest resistance to IRGM crops (Mallet and Porter 1992; Goldstein et al. 2010; Ramalho et al. 2014). Lastly, from the perspective of behavioral ecology, identifying how IRGM crops influence arthropod behavior and consequently how these effects translate into population levels may help understanding the outcomes of species interactions in arthropod communities (DiRienzo et al. 2013). In summary, the sustainable use of IRGM crops within the framework of integrated pest management (IPM) would be highly dependent on how these crops affect arthropod behaviors.

Interestingly, many studies have examined the sublethal behavioral effects of IRGM crops on arthropods, especially the behavioral traits that are highly ecologically relevant. For example, they have evaluated the feeding and oviposition behavior of phytophagous arthropods, foraging behavior of natural enemies, as well as foraging and flight activities of arthropod pollinators. Despite this, to our knowledge, no review article has been developed on this specific subject.

The scope of the current review is to systematically and critically survey the published, peer-reviewed literature regarding the behavioral effects of IRGM crops on phytophagous and beneficial arthropods (arthropod natural enemies and arthropod pollinators). The selection of literature for review not only includes the ones that involve solely the behavioral studies, but also the ones where the concomitant behavioral bioassays have been conducted with the measurements of other life-history traits (articles reviewed were selected using the ISI Web of Science database). Totally 115 case studies have been reviewed with 59 for phytophagous arthropods, 50 for arthropod natural enemies and 6 for arthropod pollinators. We point out two important aspects for the selection of the literature. First, we did not cover the studies using artificial diets contaminated with insecticidal proteins in our review. We only focused on the behavioral studies using whole plant or plant materials in field trials or laboratory bioassays, which are more ecologically relevant. Second, to enhance the quality appraisal of the covered studies in this review, the appropriateness of the non-IRGM plant material (i.e., negative control) used in the studies has been thoroughly investigated. The negative control should be as closely related to the IRGM plant as possible in order to obtain a robust comparison. To this end, we classified the negative control into three categories: "near isoline," "experimental plant" or "not specified." The negative control plant that was not clearly indicated as near the isoline or near-isogenic hybrid, but indicated as the conventional or nontransformed line, should be considered as the "experimental plants." This is also the case for the studies using cultivars that genetically modified to produce lectin, protease inhibitor or other insect-resistant feature, which have not been fully characterized and yet commercialized (e.g., Beale et al. 2006; Chen et al. 2008; Loivamäki et al. 2008; Aasen and Hågvar 2012). These studies thus need to be taken with caution when behavioral effects have been observed. The reason is that the effects observed in the cases where genetic transformation was undergone using different cultivars (instead of using near isolines) are likely to be caused by other unknown



differences in the characteristics of the two cultivars and not due to the insecticidal proteins.

The goal of this review is to provide the state-of-the-art of current literature addressing behavioral effects of IRGM crops on phytophagous and beneficial arthropods, with the aim to enhance the understanding of the compatibility of IRGM crops in crop production.

# Behavioral effects of IRGM crops on phytophagous arthropods

The high effectiveness of IRGM crops against major agricultural pests has made this technology extensively adopted by growers worldwide. However, the pest susceptibility could alter its behavior and in turn have considerable ecological consequences (Ramalho et al. 2014). Behavioral effects of IRGM crops have been extensively examined either on target or non-target phytophagous arthropods (summarized in Table 1). The behavioral traits most studied are feeding, oviposition and locomotion. These behaviors have been studied on target pests (e.g., Helicoverpa armigera, Spodoptera frugiperda, Ostrinia nubilalis) and non-target pests (e.g., Aphis gossypii, Myzus persicae, Bemisia tabaci, Adelphocoris suturalis).

Locomotion (spatial distribution, escape behavior and dispersal)

Movement and spatial distribution patterns of phytophagous insects across the plant may vary with plant quality. For phytophagous insects, aggregation in a given part of the plant indicates a preference toward this part or an avoidance of the other parts. In IRGM crops, this behavior may be linked to variations in expression levels of insecticidal proteins among those parts. For example, H. armigera larvae were mostly found on Bt cotton flowers and least on leaves, while on non-Bt cotton the larvae were equally found in flowers and leaves (Lu et al. 2011). In addition, this species has been found to move twice as far down on Bt cotton compared to non-Bt cotton (Gore et al. 2002a). Besides target phytophagous arthropods, the aggregation patterns of non-target arthropods are also affected by IRGM crops and the effects are case-dependent. For example, Bt cotton influenced the spatial distribution pattern of A. gossypii, but not for B. tabaci (Rojas et al. 2010). The aphid A. gossypii showed altered spatial distribution patterns on the crop with more alate and apterous aphids in the middle or bottom parts of Bt cotton, while the aphid population peaks took place on the nodes from the middle parts of non-Bt cotton (Fernandes et al. 2012).

There are many cases in which target arthropods tend to avoid the toxins present in IRGM crops and display an escape behavior from IRGM crops (Ramachandran et al. 1998a; Stapel et al. 1998; Gore et al. 2002b; Zhang et al. 2004; Men et al. 2005; Goldstein et al. 2010; Razze and Mason 2012). These findings suggest that a seed mixture strategy may not be an effective strategy to prevent or reduce the rate of resistance in these target pests. However, there are some exceptions that the target insects do not exhibit escape behaviors. For example, *H. zea* stayed in Bt cotton with minimal movement, which may be due to the fact that they lack the recognition of harmful substances present in the crop tissues (Jackson et al. 2010). Variations in this behavior could explain their various capabilities of developing resistance to Bt crops.

Dispersal capacity of phytophagous species reflect their colonization potential within a given habitat. Potato producing snowdrop lectin (Galanthus nivalis agglutinin, GNA) has been less likely to be colonized by alate aphid M. persicae, and the results may benefit controlling the aphids by altering the colonization behavior of alates (Aasen and Hågvar 2012). The presence of the Cry1Ac protein in Bt cotton and its probable detection by Alabama argillacea larvae increase the probability of dispersion from the plant where the larvae began (Ramalho et al. 2014). This study provided important information on how Bt cotton affects the dispersal behavior of target arthropods over time. Consequently, such information is helpful for predicting the long-term effectiveness of the "refuge-inthe-bag" tactic in management of resistance in target arthropods.

Foraging (orientation, host plant preference and feeding)

The first step of foraging is orientation, which is defined as a process through which organisms move to the sites with potential resources (Schone and Strausfeld 1984). In this process, some insects rely on visual and olfactory cues to determine their orientation (Han et al. 2010a). We could expect that orientation response in arthropods may vary with changes in host plant features. Indeed, IRGM crops may alter their chemical or physical features compared to their isogenic lines (Sun et al. 2013). However, so far, no study has been able to monitor how phytophagous arthropods locate their habitat and host plants because of the difficulties in measuring their orientation under field conditions. Still, how IRGM crops modify the orientation of phytophagous insects can be indirectly assessed by identifying where the insects perform their oviposition (see "Mating and oviposition" below).

After the orientation decision has been made, phytophagous arthropods must select host plants to feed. They are able to detect and confirm the suitability of hosts in terms of plant nutritional quality and defense (Schoonhoven et al. 2005) and then select the plants that represent



Table 1 A summary of behavioral effects of IRGM crops on phytophagous arthropods

Potato CDA   Pot	•			)					
Potato/GNA   Experimental   Myzus persicae   Target   Choice   Colonization   Delawior	References	Testing crop/gene	Negative control	Testing species	Target/non- target	Choice/non-choice	Behavior types	Effect on behavior?	Outcome
Potato/Cry3A         Not specified decentineatia         Leptinotarsa decentineatia         Target         Non-choice         Hight behavior           Eggplant/Cry3Bb         Near isoline         Leptinotarsa decentineatia         Target         Choice         Oviposition behavior           Ambidopsis/ sesquiterpene-β- plant famesene (Eβf)         Experimental Mycus persicae         Target         Choice and Choice and Beavior non-choice         Escape behavior non-choice           2A, Cry9C         plant plant         Diabrotica virgifera         Target         Choice and Choice and Beavior non-choice         Feeding behavior non-choice           Amize/Cry3Bb1         Near isoline         Diabrotica virgifera         Target         Non-choice         Feeding behavior non-choice           Rice/Bt         Experimental incernulas         Scirpophaga         Target         Non-choice         Feeding behavior non-choice           Rice/Bt         Experimental incernulas         Target         Choice         Dispersal behavior non-choice           Rice/Bt         Experimental incernulas         Scirpophaga         Target         Choice         Dispersal behavior non-choice           Cotton/Cry1Ac         Near isoline         Oxrinia nubilatis         Target         Non-choice         Spatial distribution behavior hon-choice	Aasen and Hågvar (2012)	Potato/GNA	Experimental plant	Myzus persicae	Target	Choice	Colonization behavior	Yes	Less likely to be colonized by potato producing GNA compared to near isoline
Eggplant/Cry3Bb         Near isoline         Leptinotarsa decemlineata         Target         Choice         Oviposition behavior           Potato/Cry1Ab         Near isoline         Leptinotarsa         Non-target         Choice         Oviposition behavior           Arabidopsis/ sesquiterpene-B- famesene (Eβf)         Experimental         Myzus persicae         Target         Choice and choice         Escape behavior           RiceCry1Ac, Cry plant         Cry9C         plant         Chilo suppressalis         Target         Choice and choice         Feeding behavior           A. Cry9C         plant         Diabrotica virgifera         Target         Choice and choice         Feeding behavior           A. Cry9C         plant         Diabrotica virgifera         Target         Non-choice         Feeding behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Maing behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Dispersal behavior           Rice/Bt         Recding behavior         Scirpophaga         Target         Choice         Dispersal behavior           Aphis gossypii         Non-target         Non-choice         Spatial distribution           Aphis gossypii         Target	Alyokhin and Ferro (1999)	Potato/Cry3A	Not specified	Leptinotarsa decemlineata	Target	Non-choice	Flight behavior	Yes	Resistant beetles feeding on Bt potato took longer time to initiate flight
Potato/Cry1Ab         Near isoline         Leptinotarsa decendinectua         Non-target accendinectua         Choice and accendinectua         Oviposition behavior accendinectua           Arabidopsis/ sequirepene-β- squirepene-β- famesene (Εβf)         Experimental         Myzus persicae         Target         Choice and choice         Excape behavior non-choice           2A, Cry9C         plant         Diabrotica virgifera         Target         Choice and choice         Feeding behavior non-choice           Actry9C         plant         Diabrotica virgifera         Target         Non-choice         Feeding behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Mating behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Dispersal behavior           Rice/Bt         Roar isoline         Aphiis gosxypii         Non-thoice         Spatial distribution           Maize/Cry1Ab         Near isoline         Aphiis gosxypii         Non-thoice         Spatial distribution	Arpaia et al. (2009)	Eggplant/Cry3Bb	Near isoline	Leptinotarsa decemlineata	Target	Choice	Oviposition behavior	No	The spatial distribution of egg masses were similar between IRGM and control plot
Arabidopsis/ sesquiterpene-β- famesene (Eβf) Rice/Cry1Ac, Cry Experimental Chilo suppressalis (Choice and Peeding behavior non-choice plant plant incertulas (Chilo suppressalis)  Rice/Bt Experimental Scirpophaga (Chilo suppressalis)  Rice/Bt (Chilo suppressalis		Potato/Cry1Ab	Near isoline	Leptinotarsa decemlineata	Non-target	Choice	Oviposition behavior	No	As above
Rice/Cry1Ac, Cry         Experimental         Chilo suppressalis         Target         Choice and non-choice         Feeding behavior           Aaize/Cry3Bb1         Near isoline         Diabrotica virgifera         Target         Non-choice         Feeding behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Mating behavior           Rice/Bt         Experimental         Scirpophaga         Target         Choice         Dispersal behavior           Rice/Bt         Scirpophaga         Target         Choice         Dispersal behavior           Rotton/Cry1Ac         Near isoline         Aphis gossypii         Non-target         Non-choice         Spatial distribution           Maize/Cry1Ab         Near isoline         Ostrinia nubitalis         Target         Non-choice         Movement/dispersal behavior	Beale et al. (2006)	Arabidopsis/ sesquiterpene- $\beta$ -farnesene (E $\beta$ f)	Experimental plant	Myzus persicae	Target	Choice and non-choice	Escape behavior	Yes	Repellent response in the aphids
Maize/Cry3Bb1       Near isoline       Diabrotica virgifera       Target       Non-choice       Feeding behavior         Rice/Bt       Experimental plant incertulas       Target       Choice       Mating behavior         Chilo suppressadis       Scirpophaga       Target       Choice       Dispersal behavior         Cotton/Cry1Ac       Near isoline       Aphis gossypii       Non-target       Non-choice       Spatial distribution         Maize/Cry1Ab       Near isoline       Ostrinia nubilalis       Target       Non-choice       Movement/dispersal behavior	Chen et al. (2008)	Rice/Cry1Ac, Cry 2A, Cry9C	Experimental plant	Chilo suppressalis	Target	Choice and non-choice	Feeding behavior	Yes	Preferentially fed on non-Bt culm cuttings of rice
Rice/Bt       Experimental plant       Scirpophaga incertulus       Target       Choice       Mating behavior         Chilo suppressalis       Scirpophaga incertulus       Target       Choice       Dispersal behavior         Cotton/Cry1Ac       Near isoline       Aphis gossypii       Non-target       Non-choice       Spatial distribution         Maize/Cry1Ab       Near isoline       Ostrinia nubilalis       Target       Non-choice       Movement/dispersal behavior	Clark et al. (2006)	Maize/Cry3Bb1	Near isoline	Diabrotica virgifera	Target	Non-choice	Feeding behavior	Yes	Larvae exposed to Bt maize roots consumed significantly less tissue than larvae on non-Bt maize
Scirpophaga Target Choice Dispersal behavior incertulas Chilo suppressalis Cotton/Cry1Ac Near isoline Aphis gossypii Non-target Non-choice Spatial distribution Maize/Cry1Ab Near isoline Ostrinia nubilalis Target Non-choice Movement/dispersal behavior	Cuong and Cohen (2003)	Rice/Bt	Experimental plant	Scirpophaga incertulas Chilo suppressalis	Target	Choice	Mating behavior	No	Mating occurred randomly among individuals emerging from Bt rice fields and nearby refuge fields
Cotton/Cry1Ac Near isoline Aphis gossypii Non-target Non-choice Spatial distribution  Maize/Cry1Ab Near isoline Ostrinia nubilalis Target Non-choice Movement/dispersal behavior				Scirpophaga incertulas Chilo suppressalis	Target	Choice	Dispersal behavior	N <sub>O</sub>	Dispersal capacity not affected by Bt rice
Maize/Cry1Ab Near isoline Ostrinia nubilalis Target Non-choice Movement/dispersal behavior	Fernandes et al. (2012)	Cotton/Cry1Ac	Near isoline	Aphis gossypii	Non-target	Non-choice	Spatial distribution	Yes	The distribution pattern of aphids on Bt cotton was different from those on non-Bt cotton
	Goldstein et al. (2010)	Maize/Cry1Ab	Near isoline	Ostrinia nubilalis	Target	Non-choice	Movement/dispersal behavior	Yes	Neonates abandoned more frequently Bt maize compared to non-Bt maize



Table 1 continued								
References	Testing crop/gene	Negative control	Testing species	Target/non- target	Choice/non- choice	Behavior types	Effect on behavior?	Outcome
Gore et al. (2002a)	Cotton/Cry1Ac	Near isoline	Helicoverpa armigera	Target	Choice	Movement	Yes	Movement twice as far down on Bt cotton compared to non-Bt cotton
Gore et al. (2002b)	Cotton/Cry1Ac	Near isoline	Helicoverpa zea	Target	Non-choice	Movement	Yes	More bollworms moved from the site of infestation on Bt cotton than on non-Bt cotton
Halcomb et al. (2000)	Cotton/Cry1Ac	Not specified	Helicoverpa ten	Target	Non-choice	Feeding behavior	No	Feeding indiscriminately on BTK and non-BTK cotton
			Heliothis virescens	Target	Non-choice	Feeding behavior	No	As above
Hardke et al. (2012)	Bt cotton/Cry1Ac, Cry2Ab, Cry1F	Not specified	Spodoptera frugiperda	Target	Choice	Oviposition behavior	No	Random oviposition on Bt and non-Bt cotton
Jackson et al. (2010)	Cotton/Cry1Ac, Cry1F	Not specified	Helicoverpa zea	Target	Non-choice	Movement	No	
Kumar (2004)	Cabbage/Cry1Ab	Not specified	Plutella xyllostella	Target	Choice	Oviposition/foraging behavior	No	
Lei et al. (2009)	Cotton/Cry1Ac, Cry1Ab, Cry2Ab, Cry1Fa	Near isoline	Liriomyza trifolii	Non-target	Choice and non- choice	Feeding/oviposition behavior	Yes	More eggs oviposited and more feeding on non-Bt cotton
Li et al. (2007)	Cotton/Cry1Ac, Cry2Ab	Near isoline	Trichoplusia ni	Target	Choice and non- choice	Foraging behavior	Yes	More first instars moved to non-Bt leaves, whereas the third and fifth instars did not show significant differences in the first 8 h, but eventually more moved to non-Bt leaves
Li et al. (2010)	Cotton/ Cry1Ac + CpTI	Near isoline	Adelphocoris suturalis	Non-target	Choice and non- choice	Oviposition behavior	No	
Liu et al. (2005)	Cotton/ Cry1Ac + CpTI	Near isoline	Aphis gossypii	Non-target	Non-choice	Feeding behavior	Yes	Feeding waveforms were lower; frequencies of moving and finding sites were higher on Cry1Ac + CpTI cotton compared to isogenic line
López et al. (2013)	Maize/Cry1Ab	Near isoline	Sesamia nonagrioides	Target	Chice and Non- choice	Dispersal behavior	Yes	Larvae that hatched on Bt plants tended to disperse more than those hatched on non-Bt plants



Table 1 continued								
References	Testing crop/gene	Negative control	Testing species	Target/non- target	Choice/non-choice	Behavior types	Effect on behavior?	Outcome
Lu et al. (2011)	Cotton/Cry1Ac, Cry2Ab	Near isoline	Helicoverpa armigera	Target	Choice	Spatial distribution	No	
Luong et al. (2016)	Cotton/Cry1Ac	Near isoline	Helicoverpa armigera	Target	Choice	Oviposition behavior	No	No preference for laying eggs between Bt and non-Bt cotton
Marquardt and Kruple (2009)	Maize/Cry3Bb1	Not specified	Diabrotica virgifera	Target	Non-choice	Mating behavior/ dispersal behavior	oN o	No difference in the number of captured males from refuge and Bt crops; females do not move far away from the site of emergence
Men et al. (2005)	Cotton/Cry1Ac	Not specified	Helicoverpa armigera	Target	Choice and non- choice	Feeding behavior/movement	Yes	Feeding frequency decreased and movement frequency increased in larvae
Obonyo et al. (2008)	Maize/CryAb	Near isoline	Chilo partellus Sesamia calamistis	Target	Choice and non- choice	Oviposition behavior	No	
Orr and Landis (1997)	Maize/Cry1Ab	Near isoline	Ostrinia nubilalis	Target	Choice	Oviposition	No	
Petzold-Maxwell et al. (2012)	Maize/Cry3Bb1	Near isoline	Diabrotica virgifera (Susceptible strain)	Target	Choice	Feeding behavior	No Vo	
			Diabrotica virgifera (Resistant strain)	Target	Choice	Feeding behavior	No	
Rojas et al. (2010)	Cotton/Cry1Ac	Not specified	Aphis gossypii	Non-target	Non-choice	Spatial distribution	Yes	Bt cotton altered the normal pattern of aphid distribution in the crop
			Bemisia tabaci	Non-target	Non-choice	Spatial distribution	No	
Ramachandran et al. (1998a)	Canola/Cry IAc	Near isoline	Plutella xylostella	Target	Choice	Movement	Yes	The larvae moved from Bt to non-Bt canola before acquiring lethal doses of insecticidal protein
Ramachandran et al. (1998b)	Canola/Cry1Ac	Near isoline	Plutella xylostella	Target	Choice	Oviposition behavior	No	
Ramalho et al. (2014)	Cotton/Cry1Ac	Near isoline	Alabama argillacea	Target	Non-choice	Feeding behavior/ dispersal behavior	Yes	Low acceptance of Bt plant stimulated their dispersal
Rao and Rao (2008)	Cotton/Bt	Not specified	Helicoverpa armigera	Target	Non-choice	Orientation/feeding behavior	Yes	Restless and minimum feeding behavior on Bt cotton; No orientation to Bt cotton plants



References	Testing crop/gene	Negative control	Testing species	Target/non- target	Choice/non-choice	Behavior types	Effect on behavior?	Outcome
Rovenská et al. (2005)	Eggplant/Cry3Bb	Experimental plant	Tetranychus uticae	Non-target	Choice	Feeding behavior	Yes	The spider mite preferred feeding on Bt eggplants
Rovenská and Zemek (2006)	Potato/GNA	Experimental plant	Aulacorthum solani, Thrips tabaco and Tetranychus urticae	Non-target	Choice	Foraging behavior	Yes	GNA potato less preferred by the three species in a choice test
Razze and Mason (2012)	Maize/Cry1F, Cry1Ab	Near isoline	Ostrinia nubilalis	Target	Choice	Movement/ Dispersal behavior	Yes	Movement and dispersal were higher on Bt maize
Spencer et al. (2003)	Maize/Cry3Bb1	Near isoline	Diabrotica vigifera	Target	Choice	Movement	Yes	Adults movement to adjacent soybean crops
Stapel et al. (1998)	Cotton/Bt	Experimental plant	Spodoptera exigua	Target	Non-choice	Feeding behavior	Yes	More larvae on non-Bt cotton and more damage observed on non-Bt cotton
Sun et al. (2013)	Rice/Cry2A	Experimental plant	Cnaphalocrocis medinalis	Target	Choice	Oviposition behavior	No	
Swamy et al. (2008)	Pigeonpeas/ Cry1Ab, SBTI	Experimental plant	Helicoverpa armigera	Target	Choice	Oviposition/feeding behavior	No	
Telléz-Rodríguez et al. (2014)	Maize/Cry1F	Near isoline	Spodoptera frugiperda	Target	Choice	Oviposition behavior	Yes	Oviposition preference for undamaged Bt maize over damaged conventional maize
Van den Berg and Van Wyk (2007)	Maize/Cry1Ab	Near isoline	Sesamia calamistis	Target	Choice	Oviposition behavior	No	
Whitehouse et al. (2007)	Cotton/Vegetative insecticidal protein (Vip)	Experimental plant	Mirid bugs pollen beetles	Non-target	Choice	Feeding behavior	Yes	Vip cotton attracted more bugs and beetles
Zhang et al. (2004)	Cotton/CpTI-Bt	Not specified	Helicoverpa armigera	Target	Choice and non- choice	Feeding behavior	Yes	Lepidoptera neonate larvae avoided transgenic plants
Zhao et al. (2009)	Cotton/Cry1Ac	Not specified	Helicoverpa armigera	Target	Non-choice	Calling behavior	Yes	Calling was reduced, and the age of calling was delayed in resistant strains



the suitable sites for population growth (Bernays and Chapman 1994). IRGM crops have been shown to disrupt the feeding behavior of target phytophagous arthropods in both choice and non-choice tests (Zhang et al. 2004; Men et al. 2005; Chen et al. 2008). Target species may avoid or feed less on Bt crops, even though several species have exhibited no feeding preference between Bt and non-Bt tissues in choice bioassays (Swamy et al. 2008; Petzold-Maxwell et al. 2012). The results often vary with the protocols adopted. For example, the study by Petzold-Maxwell et al. (2012) used a choice test, whereas a nonchoice test was used in an earlier study by Clark et al. (2006). Notably, many studies have exhibited that the nontarget species prefer feeding on Bt crops (Liu et al. 2005; Rovenská et al. 2005; Whitehouse et al. 2007). One mechanism that could explain such preferences is that IRGM crops (e.g., Bt crops) suffer significantly less damage by the target pests; as a result, they are often more healthy (e.g., lower levels of induced secondary metabolites that defend against the herbivorous insects) and thus could be more attractive and/or more suitable for feeding by non-target herbivores (Whitehouse et al. 2007; Hagenbucher et al. 2013).

# Mating and oviposition

Mating behavior plays an important role in mediating the effectiveness of the refuge strategy, which was designed to mitigate the development of insect resistance to IRGM crops (Gould 1998). To obtain high efficacy of such a strategy, random mating is expected to occur between the pool of the homozygous susceptible individuals from non-Bt crops and the homozygous resistant individuals from Bt crop fields (Cuong and Cohen 2003; Zhao et al. 2008; Marquardt and Kruple 2009). Alteration in mating or other related behaviors (e.g., calling behavior) of phytophagous insects could occur in IRGM crops. However, few studies have examined this subject. Zhao et al. (2009) observed reduced time spent on calling behavior in Bt-resistant H. armigera, and it may reduce the mating frequency of the resistant strain. Such behavioral changes may reduce the likelihood of hybridization, which might compromise the effectiveness of the refuge strategy.

Theory predicts that there should be a strong selection on female moths to avoid oviposition on the IRGM crops (e.g., Bt crops), which has become a sink for the main target pest. However, most of the studies have shown that the insects did not show any oviposition preference between Bt and non-Bt crops (Ramachandran et al. 1998b; Kumar 2004; Van den Berg and Van Wik 2007; Obonyo et al. 2008; Li et al. 2010; Hardke et al. 2012; Zalucki et al. 2012; Sun et al. 2013). These results suggest that the adults are not able to perceive Bt proteins, or the potential

alterations in plant features due to genetic modification do not interfere with their oviposition. By contrast, few species (either target or non-target) have shown oviposition preference for non-Bt plants (Lei et al. 2009), or Bt plants (Telléz-Rodríguez et al. 2014), when they have opportunities to choose. Notably, the latter case has provided compelling evidence that a major pest of maize, *S. frugiperda*, had a strong oviposition preference for Bt maize. Such an oviposition preference may undermine the effectiveness of refuge strategy in delaying the evolution of resistance.

#### Other behaviors

Phytophagous arthropods exhibit many other sophisticated behaviors among which learning behavior is one of the most important aspects. Learning and memory, a process defined as the acquisition and retention of neuronal representations of new information, are ubiquitous among insects (Dukas 2008). Indeed, learning is essential for host plant searching and selection because the species must learn to adjust their host plant preference in order to have higher fitness. However, little is known about how IRGM crops influence the learning behavior of phytophagous arthropods, with only few studies documenting the effects of Bt proteins on arthropod pollinators (e.g., Ramirez-Romero et al. 2008a; Han et al. 2010a).

# Behavioral effects of IRGM crops on arthropod natural enemies

Within the framework of IPM, the compatibility between IRGM crops and natural enemies has become a major subject (Lundgren et al. 2009). Natural enemies are expected to exert strong top-down control on major pest insects in IRGM crop agro-ecosystems (Han et al. 2014). However, natural enemy arthropods can be susceptible to IRGM crops. In terms of exposure pathways, IRGM crops can affect natural enemies (1) directly through exposure to insecticidal proteins via feeding on IRGM crops tissues (e.g., omnivorous predators) (Torres and Ruberson 2006), (2) indirectly because IRGM crops may induce changes in crop environment such as the quantity or nutritional quality of non-prey foods, as well as plant cues that natural enemies rely on searching for food or shelter (Lundgren et al. 2009) or (3) indirectly owing to changes in "plant-herbivore-parasitoid/predator" tri-trophic interactions in food webs (Hilbeck et al. 1998; Zwahlen et al. 2000; Couty et al. 2001; Prütz and Dettner 2004; Obrist et al. 2005; Torres et al. 2006; Walker et al. 2007; Ramirez-Romero et al. 2007; García et al. 2010). The importance of this latter pathway has been critically



reviewed (Romeis et al. 2006; Naranjo 2009; Lundgren et al. 2009). Predator and parasitoid behaviors are vital to their success in suppressing herbivorous insects (Wajnberg et al. 2007). Behavioral traits such as mobility, foraging (host/prey location, selection and suitability), mating/oviposition, orientation/associative learning and other behaviors related to specific species are susceptible to diverse biotic or abiotic constraints in realistic settings (Landis et al. 2000; Heimpel and Casas 2007). So far, numerous studies have addressed the behavioral effects of IRGM crops on natural enemies (summarized in Table 2), and the findings have been extensively discussed in the context of biological control.

#### Locomotion (mobility)

Locomotion behavior (or mobility) is characterized as a set of parameters including the total activity period, mean velocity, total distance moved or the duration of movement. Locomotion of natural enemies reflects somehow the degree of activeness during pursuit of prey/host. Taking the predatory beetle as an example, mobility is usually measured as walking speeds and the durations until the beetles twirled after being placed on their dorsum (i.e., flip time) (Lundgren and Wiedenmann 2002, 2005; Ferry et al. 2007). Several studies have examined the effects of IRGM crops on these behaviors. The spotted lady beetle Coleomegilla maculata exhibited a similar mobility when feeding on the aphids that had consumed Cry3Bb1 or non-Bt maize even though the Bt maize-fed aphids exhibited lower biomass and thus of lower prey quality (Lundgren and Wiedenmann 2005). Similarly, the potato expressing Cry3A did not affect the locomotion behavior of the ladybeetle Harmonia axyridis or the ground beetle Nebria brevicollis (Ferry et al. 2007). Furthermore, indirect evidence was found in a field study showing that three predator species exhibited no difference in abundances before, during and after pollen shed in both Bt maize and conventional maize (Pilcher et al. 1997). Overall, the trait of locomotion alone has not received much interest. Studies often prefer to measure this trait in a more ecological-relevant context such as "foraging" (the section below). In other words, the mobility has been measured when a given natural enemy has an objective to target or risk to avoid in the habitat. In contrast with predator mobility, no study has specifically assessed the effect of IRGM crops on locomotion of parasitoids. However, locomotion involves in the orientation behavior of parasitoids in response to semiochemicals [e.g., herbivore-induced plant volatiles (HIPVs)] (Kessler and Baldwin 2001), which is discussed in the following section.

#### **Foraging**

The foraging process of natural enemies involves mainly, but not exclusively, several distinct and consecutive processes: habitat location, host/prey location, host acceptance and suitability (for parasitoids), or prey handling and consumption activities (for predators) (Hågvar and Hofsvang 1991). Parasitoids and predators rely on plant-derived cues (e.g., HIPVs) as well as host/prey-derived cues (e.g., odor and frass) to locate host or prey (Dicke et al. 2000; Dicke 2009; Pareja et al. 2009; Desneux et al. 2010). After the location being made, they select the most suitable host or prey for parasitism and predation.

Parasitoids Numerous studies have documented the effect of IRGM crops on the foraging behavior of parasitoids. We firstly summarize the studies that have compared foraging between Bt and non-Bt crops. Due to the increasing evolution of pest resistance to Bt proteins (Carriere and Tabashnik 2001; Tabashnik et al. 2003; Gassmann et al. 2011; Zhang et al. 2011; Wan et al. 2012), we then compare the foraging preference between susceptible and resistant host genotypes.

There has been much evidence showing that parasitoids do not discriminate foraging on Bt or non-Bt crops (Orr and Landis 1997; Schuler et al. 2003; Turlings et al. 2005; Sanders et al. 2007; Himanen et al. 2009; Liu et al. 2011; Moraes et al. 2011; Dutra et al. 2012; Liu et al. 2015). Such an indiscrimination may be attributed to the fact that the volatile blend has not been altered in IRGM crops compared to the isogenic line (Moraes et al. 2011; Liu et al. 2015). The indiscrimination was even observed in the case where the blend of volatiles was actually altered in Bt crops (Turlings et al. 2005). However, discrimination in parasitoid foraging between Bt and non-Bt crops may still occur when the environmental condition has changed (Himanen et al. 2009). While IRGM crops themselves might not directly influence parasitoid foraging behavior, host insects feeding on these plants may experience physiological changes (e.g., modifying certain cues in frass), which may influence the attractiveness of parasitoids (Desneux et al. 2010). Such an effect may be linked to the detrimental effects of Bt protein on the bacterial community and the modification of odors that are attractive to the parasitoid. Besides the case study on Bt crops, other types of IRGM crops have exhibited strong effects on parasitoids. For instance, the parasitoid Diaeretiella rapae exhibited strong arrestment responses on the plants genetically modified to emit aphid alarm pheromone (Beale et al. 2006). In addition, no disruption in parasitoid foraging was found on crops producing GNA (Setamou et al. 2002).

Parasitoids are likely to be exposed to the co-existence of susceptible, heterozygous resistance or resistant hosts. In



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References	Testing crop/gene	Negative control	Testing species	Host/prey species	Choice/ non- choice	Behavior types	Effect on behavior?	Outcome
Parasitoids								
Beale et al. (2006)	Arabidopsis/ sesquiterpene -β-farnesene (Eβt)	Experimental plant	Diaeretiella rapae	Myzus persicae (NT)	Non- choice	Arrestant reponse	Yes	Stronger arresting response of parasitoids
Desneux et al. (2010)	Maize/Cry1Ab	Near isoline	Cotesia marginiventris	Spodoptera frugiperda (T)	Choice	Foraging behavior	Yes	The foraging behavior was disrupted by the frass derived from Bt maize-fed host
Himanen et al. (2009)	Oilseed rape/ Cry1Ac	Not specified	Cotesia vestalis	Plutella xylostella (T)	Choice	Foraging behavior/ orientation	Yes	Parasitoids orientated more to non-Bt than Bt oilseed rape under elevated O <sub>3</sub> concentrations; Such an effect was not found under elevated CO <sub>2</sub> concentrations
Liu et al. (2011)	Brocoli/Cry1Ac	Experimental plant	Diade gma insulare	Plutella xylostella (T)	Choice	Foraging behavior	No	Parasitoids did not discriminate between Bt and non-Bt Brocoli as foraging habitat
						Foraging behavior	No	Parasitoids did not discriminate among Bt (S), Bt (R) or heterozygous host genotypes
Liu et al. (2015)	Rice/CryAb	Experimenal plant	Cotesia chilonis	Chilo suppressalis (T)	Choice	Foraing behavior	No	Parasitoids did not differentiate between caterpillar-infested Bt rice and non-Bt rice
Loivamäki et al. (2008)	Arabidopsis/ PcISPS (Isoprene- emetting)	Experimental plant	Diadegma semiclausum	Unknown	Choice	Foraging behavior/ orientation	Yes	A repellent effect of GM isoprene-emitting plants on <i>D. semiclausum</i>
			Cotesia rubecula	Unknown	Choice	Foraging behavior	No	The effect above was not found on C. rubecula
Moraes et al. (2011)	Cotton/Cry1Ac	Near isoline	Trchiogramma pretiosum	Spodoptera frugiperda (T)	Choice	Foraging behavior	No	
Orr and Landis (1997)	Maize/Cry1Ab	Near isoline	Eriborus terebrans Macrocentrus grandii	Ostrinia nubilalis (T)	Choice	Foraing behavior	No No	
Sanders et al. (2007)	Maize/Cry1Ab	Near isoline	Campoletis sonorensis	Spodoptera frugiperda (T)	Choice	Foraging/host preference	No	
Schuler et al. (2003)	Oilseed rape/ Cry1Ac	Near isoline	Cotesia Plutellae	Unknown	Choice	Foraging behavior	No	Parasitoids did not discriminate between Bt and wild plants
				Plutella xylostella (T)	Choice	Foraging behavior	Yes	More attracted to Bt plants damaged by Bt (R) hosts than Bt (S) hosts



Table 2 continued									
References	Testing	Negative	Testing species	Host/prey	Choice/	Behavior types	Effect on	Outcome	
	crop/gene	control		species	-uou		behavior?		
				1	00.040				

References	Testing crop/gene	Negative control	Testing species	Host/prey species	Choice/ non- choice	Behavior types	Effect on behavior?	Outcome
Setamou et al. (2002)	Sugarcane/ GNA	Experimental plant	Cotesia flavipes	Diatraea saccharalis (NT)	Choice and non-choice	Host preference/ Location	°Z	
Turlings et al. (2005)	Maize/Cry1Ab	Near isoline	Cotesia marginiventris Microplitis rufiventris	Unknown	Choice	Foraging/ Orientation	No oN	
Predators								
Arpaia et al. (2009)	Potato/ Cry1Ab	Near isoline	Unknown generalist predators	Leptinotarsa decemlineata (NT)	Choice	Predation behavior	No	No effect on predation in Bt-potato field
	Eggplant/ Cry3Bb	Near isoline	Unknown generalist predators	Leptinotarsa decemlineata (T)	Choice	Predation behavior	No	Prey egg consumption was similar between Bt and non-Bt eggplant
Dogan et al. (1996)	Potato / Bt	Not specified	Hippodamia convergens	Myzus persicae (NT)	Non- choice	Predation behavior	No	
Dutra et al. (2012)	Maize/ Cry1Ab	Near isoline	Harmonia axyridis	Spodoptera frugiperda (T)	Choice	Predation behavior	No	
Esteves et al. (2010)	Cotton/ Cry1Ac	Near isoline	Phytoseiulus macropilis	Tetranychus urticae (NT)	Non- choice	Predation behavior	No	
	Cotton/ Cry1Ac	Near isoline	Phytoseiulus macropilis	Tetranychus urticae (NT)	Choice	Oviposition behavior	No	
Fеrry et al. (2006)	Oilseed rape/ Cry1Ac	Near isoline	Pterostichus madidus	Plutella xylostella (T)	Choice	Predation behavior	Yes	Predators preferred selecting/consuming non-Bt fed prey or Bt (R) prey than Bt (S) prey
Fетту et al. (2007)	Potato/ Cry3A	Not specified	Harmonia axyridis Nebria brevicollis	Acyrthosiphon pisum (NT) Lacanobia oleracea (T)	Non- choice Non- choice	Lotomotion Predation behavior Locomotion Predation behavior	° ° ° ° ° ° ° ° ° ° ° ° ° ° ° ° ° ° °	
Li et al. (2011)	Rice/ Cry1Ab	Near isoline	Pirata subpiraticus Tetragnatha maxillosa	Unknown	Choice	Predation behavior	No No	



No effect on the predation activity of spider The predator consumed non-Bt spider mites Spent more time in the vicinity of non-Bt Predators preferred consuming prey from maize fed prey than Bt maize-fed prey spider mites than near Bt spider mites No evidence of predators avoiding non-Predators preferred consuming non-Bt at a faster rate than Bt spider mites No effect on the spider reaction time No effect on the spider web-building non-Bt eggplant than Bt eggplant Not avoid Bt (S) prey toward prey toward prey target prey behavior Outcome Effect on behavior? Yes Yes Yes Yes Š Š s s Š Š 8 N ž å ž ž å Š å Web-building behavior Oviposition behavior Predation behavior Behavior types Mobility Mobility Mobility choice choice Choice/ choice Choice Choice choice Choice Choice Choice Choice choice Non-Non-Non-Nonnon-Rhopalosiphum Rhopalosiphum cinnabarinus littoralis (T) decemlineata littoralis (T) urticae (NT) Leptinotarsa **Tetranychus Tetranychus** Spodoptera padi (NT) Spodoptera Unknown Unknown Host/prey Unknown species (NT)Coleomegilla Chrysoperla Coleomegilla Coleomegilla Chrysoperla **Phytoseiulus** Testing species diadematus Chrysoperla **Phytoseiulus** insidiosus maculata persimilis persimilis punctipes rufilabris maculate **Micromus** maculat cupreus **Seocoris** Poecilus carneacarnea Araneus Orius spec. Experimental plant Negative control Not specified Not specified Not specified Near isoline Near isoline Near isoline Near isoline Near isoline Cry3Bb1, Cry1Ab Cry3Bb1 Eggplant/ Cry3Bb Cry1Ab crop/gene Cry1Ab Cry1Ab Cry1Ab Cry1Ac Cry3A Testing Maize/ Maize/ Maize/ Cotton/ Maize/ Maize/ Maize/ Potato/ Rovenská et al. (2005) Wiedenmann (2005) Riddick and Barbosa Torres and Ruberson Meissle et al. (2005) Pilcher et al. (1997) Meier and Hilbeck Prager et al. (2014) Fable 2 continued Ludy and Lang Lundgren and (2006b)References



Table 2 continued								
References	Testing crop/gene	Negative control	Testing species	Host/prey species	Choice/ non- choice	Choice/ Behavior types E non- b choice	Effect on behavior?	Effect on Outcome behavior?
Toschki et al. (2007) Maize/ Cry1al	Maize/ Cry1ab	Near isoline	Spiders and carabid beetles	Unknown	Non- choice	Phenological behavior	No	
Zhang et al. (2006)	Cotton/ Cry1Ac, Cry1Ab	Near isoline	Propylaea japonica	Aphis gossypii (NT)	Non- choice	Mating behavior	Yes	Predators preying on Bt-fed prey mated more frequently

T host/prey targeted by IRGM crops, NT host/prey not targeted by IRGM crops

this context, parasitoids have to make a choice among them when foraging. The decision made by parasitoids may influence how the pest population is structured, i.e., the relative abundance of the co-existing three genotypes. As a result, foraging behavior may have considerable effects on the evolution of resistance to IRGM crops in target pests (Lundgren et al. 2009), which has been demonstrated in a modeling study on *Plutella xylostella* (Onstad et al. 2013). Few empirical studies have addressed this specific issue. For example, the parasitoid Cotesia plutellae preferred to forage Bt-resistant diamondback moth than the Bt-susceptible host on Bt oil-seed rape (Schuler et al. 2003). This foraging preference may benefit the overall management of the diamondback moth as biocontrol service provided by the parasitoid can add up to the effect of the Bt crop. However, this was not always the case. For example, the parasitoid Diadegma insulare did not discriminate among different genotypes of P. xylostella feeding on Bt broccoli (Liu et al. 2011). In this case, the effects of parasitism on Bt-resistant host genotypes were diluted by the allocation of parasitism on Bt-susceptible hosts. In other words, the parasitism is actually considered redundant with the effects of Bt proteins. Furthermore, the non-discrimination in parasitoid foraging may not help reducing the selection of resistant pest populations in Bt crops.

Predators Predator foraging behavior is a set of activities that are closely associated with each other, including prey location and selection as well as the following prey handling and consumption activities. Paired-choice assays in the "Bt/non-Bt crop-prey-predator" tri-trophic systems have been largely used to test the effects of Bt crops on prey preference and consumption by predators. The effects of Bt crops on predators are discussed (1) when the predator itself belongs to the targeted insect order (e.g., Coleoptera) (direct effect via Bt protein toxicity), (2) when the prey species is targeted by Bt protein or (3) when the prey are non-target species but they can accumulate Bt proteins (e.g., spider mites).

Effects on predators may occur through direct ingestion of IRGM crop materials (e.g., pollen and tissues). Phytophagy is thought to sustain a certain group of predators (i.e., omnivorous predators) during the period of low prey availability. Thus, their fitness may be influenced via exposure to Bt protein (Lundgren et al. 2009). However, little evidence has been found for the effects of Bt crops on omnivorous predators even though feeding on Bt plants was directly observed (Moser et al. 2008). For example, the ladybird beetle *H. axyridis* and the carabid beetle *Nebria brevicollis* exhibited similar behaviors of prey selection and prey consumption on both Bt potato and conventional line (Ferry et al. 2007). The highly polyphagous predators from the Carabidae family appear to make the prey



selection decision mainly based on prey quality (see below), not based on whether the crop produces Bt protein.

In the case where prey is a target species, exposure to IRGM crops may cause lethal and sublethal effects on the prey species, which may in turn make them unsuitable for the predator. Predators have been documented to show behavioral avoidance of low-quality food (e.g., poisoned or sick prey), for example the Bt-fed susceptible (Bt (S)) prey, compared to the non-Bt fed prey (Riddick and Barbosa 1998; Meier and Hilbeck 2001; Ferry et al. 2006; but see Meissle et al. 2005) or the Bt-fed resistant (Bt (R)) prey (Ferry et al. 2006). Indeed, generalist predators are able to exhibit behavioral plasticity to cope with highly variable prey quality of numerous prey types (Symondson et al. 2002). Occasionally, they have to include the Bt (S) prey into their diet mixture to support their reproduction when the Bt (R) is scare in the habitat (Ferry et al. 2006). The predation preference for Bt (R) prey can reduce the population size of this group and avoid delivering a redundant suppression force on Bt (S) prey. When the pest is consumed in the egg stage, however, prey quality is less likely to be affected by Bt crops. Then, it is readily understandable that generalist predators consumed the eggs of target pests from the Bt and their isogenic conventional lines equally (Orr and Landis 1997; Arpaia et al. 2009).

In the case where prey is a non-target species, we should stress the importance of Hemipteran pests even though Bt proteins exhibit negligible impact on sap-feeding insects (Chougule and Bonning 2012). Specifically, aphids, as one of the most important pest groups, do not ingest considerable amounts of insecticidal proteins when feeding on Bt crops (Ramirez-Romero et al. 2008b; Romeis and Meissle 2011). Thus, they generally do not represent a major route of exposure of Bt proteins to predators as well as other natural enemies. The lack of behavioral effects of Bt-fed aphid on predators has been documented (Dogan et al. 1996; Meissle et al. 2005). By contrast, spider mites are known to acquire and accumulate higher concentrations of Bt proteins than those present in Bt crops (Dutton et al. 2002; Obrist et al. 2006; Esteves et al. 2010; Torres and Ruberson 2008; Meissle and Romeis 2009; Li and Romeis 2010). Such an exposure pathway to Bt proteins via spider mite may put their predators at risk. However, most of the studies lack the behavioral assessment of the exposed predators, or the predator was found exhibiting a similar predation rate on the prey feeding on Bt and non-Bt crops (Esteves et al. 2010). Interestingly, predators have been shown to avoid foraging a given plant type that actually has attracted abundant prey. For example, the Cry3Bb eggplants attracted more spider mites, while this type of Bt-fed prey was less attractive to the predatory mite Phytoseiulus persimilis. Such an effect has been attributed to the fact that Phytoseiulus persimilis intended to avoid the Crycontaining prey (Rovenská et al. 2005). A lower predation rate on Bt-fed spider mites by predatory mites has also been observed on Bt maize (Prager et al. 2014). Generally, predators often adopt active prey selection to compromise among maximizing energy intake, balancing the nutrient composition of the prey body and minimizing toxin consumption (Toft 1999). In the two cases described above, the disruption in predation behavior of the predatory mite, i.e., avoiding consumption of Bt-fed prey, may compromise the compatibility of Bt eggplants and predatory mites in managing spider mites.

### Mating and oviposition

Few studies have assessed the effects of IRGM crops on mating and oviposition behavior of natural enemies. Zhang et al. (2006) observed a more frequent mating in the ladybeetle Propylaea japonica feeding on Bt cotton than on non-Bt cotton, whereas such an effect was not found in the Coccinellid, H. axyridis (Ferry et al. 2007). Although the transmission of Bt proteins through non-target prey to the ladybeetles may pose sublethal effects on their behavior, the mechanisms underlying such effects remain to be explored. After mating, predators should select a suitable plant species or plant part as oviposition site. It has been shown that predators rely upon physical traits such as substrate sickness, trichome density and shape when selecting an oviposition site (Shapiro and Ferkovich 2006; Lundgren et al. 2008; Seagraves et al. 2011). Despite the unintentional effects that may arise in the process of genetic modification in plants (Haslberger 2003; Cellini et al. 2004), these changes may not be able to disrupt the oviposition behavior of arthropod predators. Indeed, Torres and Ruberson (2006) showed that the spatial distribution pattern of predator eggs was similar between Bt and conventional cotton plots, indicating no difference in their site selection for oviposition. Similar results were found with the predator mite Phytoseiulus macropilis ovipositing on Bt and non-Bt cotton (Esteves et al. 2010).

In the case of parasitoids, mating behavior has been experimentally observed in the field studies (Antolin and Strand 1992; Fauvergue et al. 1999). However, no comparative study has been conducted to test the effects of IRGM crops on their mating behavior. The following oviposition behavior often refers to laying eggs inside the host bodies (larvae or eggs) or on the surface of the bodies, which is actually one part of the foraging process. For example, *Parallorhogas pyralophagus* females preferred to probe, drill and parasitize *Eoreuma loftini* larvae, the Mexican rice borer, fed on GNA-free sugarcanes than those fed on GNA-containing sugarcanes (Tomov et al. 2003). Such a parasitism preference may benefit the control of *E. loftini*. However, the ecological relevance needs to be



further assessed as GNA-containing sugarcane materials mixed in artificial diets was used in this study. Another case study has reported a neutral effect that the larval parasitoid *D. insulare* exhibited a similar parasitism rate between Bt broccoli-fed resistant and non-Bt broccoli fed *P. xylostella* larvae (Liu et al. 2011). Besides the direct observation of oviposition behavior, we can assess the oviposition behavior indirectly by examining the outcome of oviposition behavior (i.e., parasitism rate). However, this approach may be misleading since the parasitism rate can be influenced by host acceptance and suitability (Desneux et al. 2012).

#### Other behaviors

Orientation and foraging of natural enemies rely on an important activity, i.e., the associative learning of visual cues and oviposition/herbivory- induced plant volatiles (Lewis and Tumlinson 1988; Vet and Groenewold 1990; Meiners et al. 2003; De Boer and Dicke 2006; Schroder et al. 2008; Desouhant et al. 2010; Benelli and Canale 2012). There has been an increasing recognition of associative learning as an important capacity in foraging success, either for predators or parasitoid wasps (De Boer et al. 2005; Dukas 2008). For example, the parasitoid Leptopilina heterotoma can learn to respond to a novel odor of a given pest species that can be used in the subsequent host location. Such an association can be stronger when the parasitoid is rewarded with an oviposition experience (Vet and Groenewold 1990). Odor learning is of adaptive importance for this generalist parasitoid, since it helps enhance host location efficiency by reducing the time allocated to the decision on where to search for hosts (Canale et al. 2014). To our knowledge, however, no study so far has been conducted to assess the effects of IRGM crops on associative learning in arthropod natural enemies.

Other specific behaviors of concern are mainly exhibited by the species from the class Arachnida. Spiders exert strong predation pressures on a wide range of insect taxa and thus considerably contribute to the biocontrol services in the garden or cropping field (Peterson et al. 2011). Although spiders are exposed to Bt proteins by feeding pollen containing the proteins or infected prey (Ludy and Lang 2006a), such an exposure did not affect their fitness and predation activity on prey (Ludy and Lang 2006b; Meissle and Romeis 2009; also see review by Peterson et al. 2011). For example, the garden spider Araneus diadematus spent similar reaction times toward prey fed with Bt and conventional maize pollen. Moreover, orb-web geometry, the most direct parameter of the spider's webbuilding behavior, was not disrupted when they consumed Bt maize pollen (Ludy and Lang 2006b). A 3-year field study also demonstrated that the phenological behavior of spiders and carabid beetles was not affected by the Bt maize (Toschki et al. 2007).

# Behavioral effects of IRGM crops on arthropod pollinators

Since the debate over the potential risk of Bt plant pollen on the survival and feeding behavior of the monarch butterfly (Losey et al. 1999; Hansen Jesse and Obrycki 2000; Hellmich et al. 2001), concerns have been increasingly raised over the negative effects of IRGM crops on arthropod pollinators. Honeybees, bumblebees as well as other non-bee pollinators are the most known functional group of arthropod pollinators in nature (Rader et al. 2016). In Bt crops, for example, the cotton nectar or maize pollen attracts honeybee foragers (Babendreier et al. 2004; Duan et al. 2008) and thus becomes a potential route of exposure to the Bt proteins for honeybees (EFSA 2013). In certain cultivars, a relatively high expression level of Bt proteins has been reported (Han et al. 2010a, b). This scenario becomes even worse for the foraging bees if a given crop variety is extensively cultivated in an area where alternative nectar/pollen-producing plants are unavailable or out of season (Haydak 1970). Therefore, honeybee has been considered a key non-target arthropod surrogate in the framework of environmental risk assessments of IRGM crops (Romeis et al. 2008). Two review articles have concluded that Bt proteins did not induce lethal effects on honeybee larvae or adults (Duan et al. 2008; Malone and Burgess 2009), and the neutral effects have been documented in the more recent literature (Liu et al. 2009; Han et al. 2010a, b, 2012; Lima et al. 2011, Hendriksma et al. 2011; Dai et al. 2012a, b, 2015; Niu et al. 2013).

Several studies have examined the behavioral effects of Bt plant materials on feeding behavior (Han et al. 2010b), foraging behavior (Dai et al. 2012a) and olfactory/visual learning behavior (Han et al. 2010a; Dai et al. 2012a) in the honeybee Apis mellifera as well as larvae feeding behavior, movement (Prasifka et al. 2007) and oviposition behavior (Tschenn et al. 2001) in the monarch butterfly Danaus plexippus (summarized in Table 3). The deleterious effects of Bt pollen on honeybee feeding behavior may lead to malnutrition of the colony, though the mechanisms underneath such effects and the ecological consequences remain to be explored. The monarch butterfly was found to exhibit similar oviposition behavior on milkweed plant dusted pollen from Bt maize or the conventional maize (Tschenn et al. 2001). However, they behaved differently when exposed to Cry1Ab-expressing maize anthers in which they exhibited increased wandering and more movement under laboratory conditions (Prasifka et al. 2007). Still, whether such behavioral alterations could translate into population effects in the field remains to be



known. The results may be case dependent. In a larger scale experiment in greenhouses, however, the pollination activities of bumblebees were found to be compatible with Bt eggplant (Arpaia et al. 2011).

Other behavioral assays on honeybees or bumblebees have used artificial diets contaminated with insecticidal Cry protein or other toxic proteins (e.g., SBTI, BBI, CpTI) regarding feeding behavior (Malone and Pham-Delègue 2001; Ramirez-Romero et al. 2005, 2008a; Babendreier et al. 2008), flight activity (Malone et al. 2001), foraging (Picard-Nizou et al. 1995; Ramirez-Romero et al. 2005; Mommaerts et al. 2010) and olfactory learning behavior (Picard-Nizou et al. 1997; Pham-Delègue et al. 2000; Ramirez-Romero et al. 2008a). Most studies have performed the bioassays using artificial diets contaminated with high doses of those insecticidal proteins. Negative effects on olfactory learning and feeding behavior in honeybees have been reported (Picard-Nizou et al. 1997; Ramirez-Romero et al. 2005, 2008a). However, the ecological relevance of those behavioral effects needs to be further explored (Ramirez-Romero et al. 2008a).

# The importance of behavioral aspects for sustainable use of IRGM crops

As part of the environmental risk assessment process that takes place before the commercial release of IRGM crops, the potential adverse effects of IRGM on non-target arthropods, notably the natural enemies and arthropod pollinators, have been considered an extremely important issue for the scientific community, public and regulatory agencies (Raybould 2006; Raybould et al. 2007; Rose 2007; Romeis et al. 2008, 2011; Hilbeck et al. 2012). A tiered approach has been internationally recognized, which focuses on problem formulation including policy goals, scope, assessment endpoints and methodology, testing of clearly defined risk hypotheses and progressing between testing tiers (i.e., laboratory, semi-field and open field conditions) (Romeis et al. 2008; Álvarez-Alfageme et al. 2011). To increase the robustness and confidence of data gathered under laboratory conditions, Romeis et al (2011) provided recommendations on the experimental design concerning the selection of surrogate species and lifestages, test substance, measurement endpoints, test duration, control substances and statistical considerations under the defined standards of good laboratory practice. Currently, the primary measurement endpoints include mortality, fecundity, developmental duration of each life stage, growth, predation rate, parasitism rate and the percentage of testing individuals reaching a certain life stage. Behavioral aspects have been rarely considered in the legislation of IRGM crops. So far, no behavioral effect that later caused significant harm (e.g., failure of crop yield or pest outbreaks) in the field has been missed in the pre-market risk assessment as it was conducted. However, we claim that special attention needs to be paid to several key behavioral traits either before or after the release of the IRGM crop line, such as locomotion/oviposition of the target Lepidopterans, olfactory/visual associate learning of honeybees and foraging behavior (e.g., prey/host selection) of arthropod natural enemies. Such an assertion is in line with the guidance document by the European Food Safety Authority (EFSA) regarding the risk assessment of IRGM crops on phytophagous arthropods and arthropod pollinators (EFSA 2011, 2013).

One major concern is that behavioral effects on phytophagous arthropods (especially the target species) might have important implications for management of resistance evolution. This issue becomes crucial as the resistance to Cry1F maize has been documented on S. frugiperda (Storer et al. 2010). One possible mechanism is that the isogenic conventional maize that was heavily damaged by S. frugiperda becomes less attractive to the ovipositing adults than Cry 1F maize. Such a behavioral change, i.e., the damage-avoiding oviposition as populations increase, may accelerate the resistance evolution and either lead to requirements for a larger refuge or undermine resistance management altogether (Telléz-Rodríguez et al. 2014). Besides oviposition, an increased larval dispersal rate with less exposure to Bt crops may also result in a lower efficiency for the refuge strategy (López et al. 2013). Indeed, physiologically mediated resistance is considered to develop more slowly when the species shows avoidance of food containing Bt proteins (Jallow and Hoy 2007; Onstad 2008). All these behavioral effects should be of major concern to the scientific community.

Pollinators such as honeybees exhibited many behavioral traits namely foraging, orientation, feeding, coordination, waggling dances, nestmate recognition and repellency, which are highly valued by apiculture and agriculture. Among them, foraging is a critical behavior for the nutritional status and health of the bee population (Malone and Pham-Delègue 2001; Chittka et al. 2003; Decourtye et al. 2010). When exposed to risky abiotic factors, the relevant traits, such as associate olfactory/visual learning capacities, are expected to be carefully assessed (Greggers and Menzel 1993; Ramirez-Romero et al. 2005, 2008a, b; Han et al. 2010a; Dai et al. 2012a). Furthermore, more realistic field studies on pollinators can follow laboratory evaluation procedures (Thompson 2003; Decourtye et al. 2004, 2013; Mommaerts et al. 2009; Gill et al. 2012; Rondeau et al. 2014; Dively et al. 2015). Once some behavioral effect has been observed, some procedures should be further tested, renewed and standardized. Numerous promising tools can be used and improved in future behavioral studies (summarized in Table 4).



**Table 3** A summary of behavioral effects of IRGM crops on arthropod pollinators

References	Crop/gene	Control crop	Arthropod species	Behavior types	Effect on behavior?	Outcome
Dai et al. (2012a)	Maize/Cry 1Ah	Near isoline	Apis mellifera	Foraging behavior	No	
				Olfactory learning behavior	No	
Han et al.	Cotton/	Near isoline	Apis mellifera	Olfactory learning/	No	
(2010a)	Cry1Ac + CpTI			Visual learning behavior		
Han et al. (2010b)	Cotton/ Cry1Ac + CpTI	Near isoline	Apis mellifera	Feeding behavior	Yes	Antifeedant effect on honeybees
Prasifka et al. (2007)	Maize/Cry1Ab	Near isoline	Danaus plexippus	Movement/feeding behavior	Yes	Larvae exposed to Bt anthers spent more time moving and less time for feeding on anthers
Tschenn et al. (2001)	Maize/Cry1Ab	Near isoline	Danaus plexippus	Oviposition behavior	No	

While behavioral effects on arthropod natural enemies are less advocated by researchers and regulators, we do advocate that understanding natural enemy foraging preferences among various genotypes of pest insects (susceptible, resistance or heterozygous resistance) is vital to increase the synergy between IRGM crops and the biocontrol agents in pest management. For instance, we could improve IPM if natural enemies prefer to attack resistant pest genotypes (Riddick and Barbosa 1998; Meier and Hilbeck 2001; Schuler et al. 2003; Ferry et al. 2006). Morever, contrast, the unexpected foraging preferences may undermine the goal of IPM when predators do not prefer to locate on Bt crops where pests have been attracted (Rovenská et al. 2005). Other important behavioral traits in natural enemies such as mating and oviposition should also be investigated for the potential risks of IRGM crops, following the experience of studying sublethal effects of insecticides on arthropod natural enemies (Desneux et al. 2007; Biondi et al. 2012).

Furthermore, the behavioral aspects need to be carefully assessed for the following reasons. First, the process of behavioral effects assessment is likely to offer new insights into examining associated physiological aspects that have not been considered before. Behavioral effects often coincide with physiological change in arthropods (Desneux et al. 2007), and any evidence of behavioral disruption may trigger additional tests on physiological changes. For example, the antifeedant effect observed in honeybees during an oral exposure to cotton pollen containing Cry1Ac (Han et al. 2010b) has triggered further investigations into the possible effects of antifeedant behavior on hypopharyngeal gland development and into the mechanisms underlying such an antifeedant effect (Han et al. 2012). Sublethal behavioral effects are often recorded by detailed

observation; once verified, the mechanisms underlying the effect are expected to be explored. The data from the following in-depth studies can not only enhance the reliability of the observed behavior effect, but also propose new questions. Second, rapid advances in insect behavioral ecology from theoretical approaches to field applications have been achieved with a vision of multi-trophic interactions (Wajnberg et al. 2007). One of the longstanding goals of behavioral ecology is to understand how individual behavioral changes translate into population processes and to unravel the role of these traits in shaping community structure (Werner and Peacor 2003; Vet and Godfray 2007). Based on this point, with the increasing cultivation of IRGM crops worldwide, the agro-ecosystem landscapes are undergoing tremendous changes in community structure and functioning of biological interactions. If the assumptions above are true, more attention needs to be paid to the responses of insect behaviors (mainly major pest species, key natural enemies and arthropod pollinators) in future studies.

# **Conclusions**

The widespread cultivation of IRGM crops creates an agricultural environment conserving the ecological services of resident and immigrant natural enemies (Lundgren et al. 2009; Lu et al. 2012) and arthropod pollinators (Johnson et al. 2010). The concurrent benefit is that the reduction in insecticide applications, may favor the fitness of beneficial arthropods (Thompson 2003; Desneux et al. 2007). However, behavioral effects of IRGM crops on phytophagous arthropods (mainly major agricultural pest), natural enemies and arthropod pollinators need to be



Table 4 An overview of promising tools/procedures for assessing behavioral effects of IRGM crops on arthropod pollinators and natural enemies

Arthropod group	Behavioral trait	Evaluation tools/procedures	References
Pollinators	Olfactory learning	Conditioned-PER (proboscis extension response)	Picard-Nizou et al. (1997)
			Pham-Delègue et al. (2000)
			Ramirez-Romero et al. (2005, 2008a)
			Han et al. (2010a)
	Visual learning	T-maze	Han et al. (2010a)
	Foraging behavior/	Artificial flower feeder	Pierre et al. (2003)
	Pollination	Field parallel strips observation	Decourtye et al. (2004)
		Evaluation of foraging bees returning to hives	Ramirez-Romero et al. (2005)
			Dai et al. (2012a)
	Flight activity	Evaluation of tagged bees	Malone et al. (2001)
Parasitoids	Foraging behavior	"Y-tube" dual-choice olfactometer	Moraes et al. (2011)
		Four-arm olfactometer	Vet and Groenewold (1990)
			Setamou et al. (2002)
			Desneux et al. (2010)
		Wind tunnel	Schuler et al. (1999, 2003)
		Six-arm olfactometer	Turlings et al. (2005)
	Olfactory learning	Conditioning using host frass/ordor and subsequent testing	Meiners et al. (2003)
Predators	Locomotion/foraging	Visual-observation or video-recording in arena	Meier and Hilbeck (2001)
	behavior		Meissle et al. (2005)
			Ferry et al. (2006, 2007)
			Dutra et al. (2012)
	Predation behavior	DNA-based gut-content analysis	Harwood et al. (2007)
	Predation behavior	PCR-ligase detection reaction (LDR)	Li et al. (2011)
	Predation behavior	Modified Munger cell	Rovenská et al. (2005)
	Predation/ mobility/web building	Wood frames with spider web exposure to IRGM crop pollen in the field and feeding by spiders	Ludy and Lang (2006b)

carefully assessed in order to maintain the potential benefits of IRGM crops in agriculture. Key results of this review are outlined:

- 1. Among the three groups, phytophagous arthropods are the most affected by the IRGM crops (54.2 % out of total cases) (Table 5). Locomotion and foraging behaviors are frequently affected by IRGM crops, while mating and oviposition behaviors are less affected. Target species are more likely to experience behavioral changes (e.g., locomotion and feeding), and the implications of these behavioral effects relevant to insect resistance evolution need to be considered with caution. The altered behavior in non-target arthropods (preference to Bt crops) may contribute to higher damage to Bt crops.
- 2. The minority of the studies (22 %) exhibited behavioral effects on arthropod natural enemies (Table 5). The effects mainly refer to the behavioral preference

when they select their host or prey. Generally, they avoid the host or prey of lower quality because of the lethal and/or sublethal effects of IRGM crops. However, we should notice that these behavioral effects may be unnecessarily due to the presence of plant insect-resistant traits. In around 30% of the total case studies reporting behavioral effects, the genetic background of negative control plants has not been fully characterized, and the plant counterparts are likely to differ in many other unknown characteristics as discussed above. Thus, caution needs to be taken when referring to these studies. Overall, we conclude that behavioral effects of IRGM crops on arthropod natural enemies via tri-trophic links are limited. However, we should not underestimate the unintentional behavioral effects that may undermine their biocontrol services in agro-ecosystems (e.g., predation behaviors, Rovenská et al. 2005; Prager et al. 2014).



Table 5 A summary of existing case studies on behavioral effects of IRGM crops on phytophagous arthropods, arthropod natural enemies and arthropod pollinators

		Effect (yes)			Effect (no)		Total
Phytophagous arthropods	Target		Non-target	Target		Non-target	
Number of studies	22		10	25		2	59
Percentage		54.2% (yes)			45.8% (no)		
Arthropod natural enemies	Target prey/host	Non-target prey/host	Unknown	Target prey/host	Non-target prey/host	Unknown	
Number of studies	5	5	1	15	8	16	50
Percentage		22% (yes)			78% (no)		
Arthropod pollinators							
Number of studies		2			4		6
Percentage		33% (yes)			67% (no)		

Effect (yes) or effect (no) indicates the presence or absence of behavioral effects. The effects on phytophagous arthropods have been separated as the species is targeted (or not) by the genetically modified insect resistance. The effects on natural enemies have been separated as the prey/host species is targeted (or not) by the genetically modified insect resistance or unknown. The proportions of observations of behavioral effects for each arthropod group were calculated based on the number of case studies

3. Behavioral effects of IRGM crops on arthropod pollinators are relatively limited (Table 5). Negative effects on honeybee foraging behavior have been found in several studies using artificial diets containing transgenic plant materials. More realistic bioassays are required to better understand the ecological relevance.

Understanding the behavioral effects of IRGM crops on phytophagous and beneficial arthropods is critical for the compatibility among genetically modified insect resistance, biological control and beneficial pollinators in securing crop production. In general, behavioral alterations in arthropods due to IRGM crops seem not deleterious to the goal of IPM. However, the current knowledge is far from telling us how those arthropods are actually influenced and what the real ecological consequences are. We face various constraints in enhancing our understanding on this subject. For example, current studies have been mostly carried out in laboratory settings with semi-field and field tests largely lagging behind, which is especially the case for parasitoids (Heimpel and Casas 2007). The techniques available for behavioral tests have been well developed but are still limited (Table 4). Future efforts can be made to design more reliable tools for behavioral studies under field conditions. Furthermore, it provides valuable insights about the effects of behavioral changes on the community dynamics of interacting species (Vet and Godfray 2007). It is important to understand the indirect interactions triggered by behavioral traits, for example, the trait-mediated indirect interactions among the natural enemies and other organisms from different trophic levels within the arthropod community (Werner and Peacor 2003; Schmitz et al. 2004). As IRGM crops become increasingly widespread worldwide, we believe that more in-depth behavioral studies on plant-inhabiting arthropods are going to considerably improve the sustainability of agriculture using IRGM crops.

#### **Author contribution statement**

RRR and ND conceptualized the review article. PH, MCVH, RRR and ND wrote the manuscript.

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#### Compliance with ethical standards

Conflict of interest The authors declare no conflict of interest.

Ethical approval This article does not contain any studies with human participants or animals performed by any of the authors.



### Glossary

Associative The behavior involves the

learning behavior establishment, through experience, of

an association between two stimuli or between a stimulus and a response.

Sublethal The behavioral alteration in an behavioral effects arthropod individual that survives an

exposure to plant materials containing genetically modified insect-resistant proteins or other

toxic compounds.

for the purpose of attracting mates

and repelling rivals.

Dispersal behavior The capacity of an arthropod in

moving/flight and the potential for its

spatial distribution.

Foraging behavior The behavior of an arthropod

searching for food, host or prey.

Refuge-in-the-bag A tactic that planting of mixed Bt

and non-Bt crops for preventing or delaying resistance evolution in

target arthropod species.

#### References

- Aasen SS, Hågvar EB (2012) Effect of potato plants expressing snowdrop lectin (GNA) on the performance and colonization behavior of the peach-potato aphid *Myzus persicae*. Acta agriculturae Scandinavica 62:352–361
- Álvarez-Alfageme F, Bigler F, Romeis J (2011) Laboratory toxicity studies demonstrating no adverse effects of Cry1Ab and Cry3Bb1 to larvae of *Adalia bipunctata* (Coleoptera:Coccinellidae): the importance of study design. Transgenic Res 20:467–479
- Alyokhin AV, Ferro DN (1999) Modifications in dispersal and oviposition of BT-resistant and Bt-susceptible Colorado potato beetles as a result of exposure to *Bacillus thuringiensis* subsp. Tenebrionis Cry3A toxin. Entomol Exp Appl 90:93–101
- Andow DA, Zwahlen C (2006) Assessing environmental risks of transgenic plants. Ecol Lett 9:196–214
- Antolin MF, Strand MR (1992) Mating system of *Bracon hebetor* (Hymenoptera: Braconidae). Ecol Entomol 17:1–7
- Arpaia SJ, Schmidt EU, Di Leo GM, Fiore MC (2009) Oviposition of the Colorado potato beetle (*Leptinotarsa decemlineata*) and natural predation on its egg masses in Bt-expressing fields. Biocontrol Sci Technol 19:971–984
- Arpaia S, De Cristofaro A, Guerrieri E, Bossi S, Cellini F, Di Leo GM et al (2011) Foraging activity of bumblebees (*Bombus terrestris* L.) on Bt-expressing eggplants. Arthropod-Plant Interact 5:255–261
- Babendreier D, Kalberer NM, Romeis J, Fluri P, Bigler F (2004) Pollen consumption in honey bee larvae: a step forward in the risk assessment of transgenic plants. Apidologie 35:293–300
- Babendreier D, Bigler F, Kuhlmann U (2006) Current status and constraints in the assessment of non-target effects. In Bigler et al: Environmental Impact of Invertebrates for Biological Control of Arthropods. CAB International, pp. 1–14

Babendreier D, Reichhart B, Romeis J, Bigler F (2008) Impact of insecticidal proteins expressed in transgenic plants on bumblebee microcolonies. Entomol Exp Appl 126:148–157

- Beale MH, Birkett MA, Bruce TJ, Chamberlain K, Field LM, Huttly AK et al (2006) Aphid alarm pheromone produced by GM plants affects aphid and parasitoid behavior. Proc Natl Acad Sci 103:10509–10513
- Benelli G, Canale A (2012) Learning of visual cues in the fruit fly parasitoid *Psyttalia concolor* (Szepligeti) (Hymenoptera: Braconidae). Biocontrol 57:767–777
- Bernays EA, Chapman RF (1994) Host-plant selection by phytophagous insects. Chapman & Hall, New York
- Biondi A, Mommaerts V, Smagghe G, Vinuela E, Zappala L, Desneux N (2012) The non-target impact of spinosyns on beneficial arthropods. Pest Manag Sci 68:1523–1536
- Brookes G, Barfoot P (2006) Global impact of biotech cops: Socioeconomic and environmental effects in the first 10 years of commercial use. AgBioForum 9:139–151
- Canale A, Geri S, Benelli G (2014) Associative learning for host-induced fruit volatiles in *Psyttalia concolor* (Hymenoptera: Braconidae), a koinobiont parasitoid of tephritid flies. Bull Entomol Res 104:774–780
- Carriere Y, Tabashnik BE (2001) Reversing insect adaptation to transgenic insecticidal plants. Proc R Soc B—Biol Sci 268:1475–1480
- Cellini F, Chesson A, Colquhoun I, Constable A, Davies HV, Engel KH et al (2004) Unintended effects and their detection in genetically modified crops. Food Chem Toxicol 42:1089–1125
- Chen H, Mang G, Zhang QF, Lin YJ (2008) Effect of transgenic *Bacillus thuringiensis* rice lines on mortality and feeding behavior of rice stem borers (Lepidoptera: Crambidae). J Econ Entomol 101:182–189
- Chittka L, Dyer AG, Bock F, Dornhaus A (2003) Bees trade off foraging speed for accuracy. Nature 424:388
- Chougule NP, Bonning BC (2012) Toxins for transgenic resistance to Hemipteran pests. Toxins 4:405–429
- Clark PL, Vaughn TYT, Meinke LJ, Molina-Ochoa J, Foster JE (2006) *Diabrotica virgifera virgifera* (Coleoptera: Chrysomelidae) larval feeding behavior on transgenic maize (MON 863) and its isoline. J Econ Entomol 99:722–727
- Couty A, Clark SJ, Poppy GM (2001) Are fecundity and longevity of female Aphelinus abdominalis affected by development in GNAdosed Macrosiphum euphorbiae? Physiol Entomol 26:287–293
- Cuong NL, Cohen MB (2003) Mating and dispersal behavior of Scirpophaga incertulas and Chilo suppressalis (Lepidoptera: Pyralidae) in relation to resistance management for rice transformed with Bacillus thuringiensis toxin genes. Int J Pest Manag 49:275–279
- Dai PL, Zhou W, Zhang J, Cui HJ, Wang Q, Jiang WY et al (2012a) Field assessment of Bt cry1Ah corn pollen on the survival, development and behavior of *Apis mellifera* ligustica. Ecotoxicol Environ Saf 79:232–237
- Dai PL, Zhou W, Zhang J, Jiang WY, Wang Q, Cui HJ et al (2012b)
  The effects of Bt Cry1Ah toxin on worker honeybees (*Apis mellifera ligustica* and *Apis cerana cerana*). Apidologie 43:384–391
- Dai PL, Zhou W, Zhang J, Lang ZH, Zhou T, Wang Q et al (2015) Effects of Bt cry1Ah corn pollen on immature workers of Apis cerana cerana. J Apicult Res 54:72–76
- De Boer JG, Dicke M (2006) Olfactory learning by predatory arthropods. Anim Biol 56:143–155
- De Boer JG, Snoeren TAL, Dicke M (2005) Predatory mites learn to discriminate between plant volatiles induced by prey and nonprey herbivores. Anim Behav 69:869–879
- Decourtye A, Devillers J, Cluzeau S, Charreton M, Pham-Delègue MH (2004) Effects of imidacloprid and deltamethrin on



associative learning in honeybee under semi-field and laboratory conditions. Ecotoxicol Environ Saf 57:410-419

- Decourtye A, Mader E, Desneux N (2010) Landscape scale enhancement of floral resources for honey bees in agroecosystems. Apidologie 41:264–277
- Decourtye A, Henry M, Desneux N (2013) Overhaul pesticide testing on bees. Nature 497:188
- Desneux N, Bernal JS (2010) Genetically modified crops deserve greater ecotoxicological scrutiny. Ecotoxicology 19:1642–1644
- Desneux N, Decourtye A, Delpuech JM (2007) The sublethal effects of pesticides on beneficial arthropods. Annu Rev Entomol 52:81–106
- Desneux N, Ramirez-Romero R, Bokonon-Ganta AH, Bernal JS (2010) Attraction of the parasitoid *Cotesia marginiventris* to host frass is affected by GM maize. Ecotoxicology 19:1183–1192
- Desneux N, Roger Blahnik, Delebecque CJ, Heimpel GE (2012) Host phylogeny and specialization in parasitoids. Ecol Lett 15:452–460
- Desouhant E, Navel S, Foubert E, Fischbein D, Théry M, Bernstein C (2010) What matters in the associative learning of visual cues in foraging parasitoid wasps: colour or brightness? Anim Cogn 13:535–543
- Dicke M (2009) Behavioural and community ecology of plants that cry for help. Plant Cell Environ 32:654-665
- Dicke M, Schütte C, Dijkman H (2000) Change in behavioral response to herbivore-induced plant volatiles in a predatory mite population. J Chem Ecol 26:1497–1514
- Dively GP, Embrey MS, Kamel A, Hawthorne DJ, Pettis JS (2015) Assessment of chronic sublethal effects of imidacloprid on honey bee colony health. PLoS One 10(3):e0118748. doi:10. 1371/journal.pone.0118748
- Dogan EB, Berry RE, Reed GL, Rossignol PA (1996) Biological parameters of convergent lady beetles (Coleoptera: Coccinellidae) feeding on aphids (Homoptera: Aphididae) on GM potato. J Econ Entomol 89:1105–1108
- Duan JJ, Marvier M, Huesing J, Dively G, Huang ZY (2008) A metaanalysis of effects of Bt crops on honey bees (Hymenoptera: Apidae). PLoS One 3(1):e1415. doi:10.1371/journal.pone. 0001415
- Duan JJ, Lundgren JG, Naranjo S, Marvier M (2010) Extrapolating non-target risk of Bt crops from laboratory to field. Biol Lett 6:74–77
- Dukas R (2008) Evolutionary biology of insect learning. Annu Rev Entomol 53:145–160
- Dutra CC, Koch RL, Burkness EC, Meissle M, Romeis J, Hutchison WD et al (2012) Harmonia axyridis (Coleoptera: Coccinellidae) exhibits no preference between Bt and on-Bt maize fed Spodoptera frugiperda (Lepidoptera: Noctuidae). PLoS One 7(9):e44867. doi:10.1371/journal.pone.0044867
- Dutton A, Klein H, Romeis J, Bigler F (2002) Uptake of Bt-toxin by herbivores feeding on transgenic maize and consequences for the predator *Chrysoperla carnea*. Ecol Entomol 27:441–447
- EFSA (2010) Guidance on the environmental risk assessment of genetically modified Plants. EFSA J 8:1879
- EFSA (2011) Scientific Opinion updating the evaluation of the environmental risk assessment and risk management recommendations on insect resistant genetically modified maize 1507 for cultivation. EFSA J 9:2429
- EFSA (2013) EFSA Guidance Document on the risk assessment of plant protection products on bees (Apis mellifera, Bombus spp. and solitary bees). EFSA J 11:3295
- Esteves AB, de Oliveira JV, Torres JB, Gondim MGC (2010) Compared biology and behavior of *Tetranychus urticae* Koch (Acari: Tetranychidae) and *Phytoseiulus macropilis* (Banks) (Acari: Phytoseiidae) on Bollgard (TM) and non-Transgenic Isoline Cotton. Neotropical Entomol 39:338–344

- Fauvergue X, Fleury F, Lemaitre C, Allemand R (1999) Parasitoid mating structures when hosts are patchily distributed: field and laboratory experiments with *Leptopilina boulardi* and *L. heterotoma*. Oikos 86:344–356
- Fernandes FS, Ramalho FS, Nascimento J, Malaquias JB, Nascimento ARB, Silva CAD et al (2012) Within-plants distribution of cotton aphids, *Aphis gossypii* Glover (Hemiptera: Aphididae) in Bt and non-Bt cotton fields. Bull Entomol Res 102:79–87
- Ferry N, Mulligan EA, Stewart CN, Tabashnik BE, Port GR Gatehouse AMR (2006) Prey-mediated effects of GM canola on a beneficial, non-target, carabid beetle. Transgenic Res 35:501–514
- Ferry N, Mulligan EA, Majerus MEN, Gatehouse AMR (2007) Bitrophic and tritrophic effects of Bt Cry3A GM potato on beneficial, non-target, beetles. Transgenic Res 16:795–812
- García M, Ortego F, Castañera P, Farinós GP (2010) Effects of exposure to the toxin Cry1Ab through Bt maize fed-prey on the performance and digestive physiology of the predatory rove beetle Atheta coriaria. Biol Control 55:225–233
- Gassmann AJ, Petzold-Maxwell JL, Keweshan RS, Dunbar MW (2011) Field-evolved resistance to Bt maize by western corn rootworm. PLoS One 6(7):e22629. doi:10.1371/journal.pone. 0022629
- Gill RJ, Ramos-Rodriguez O, Raine NE (2012) Combined pesticide exposure severely affects individual- and colony-level traits in bees. Nature 491:105–119
- Goldstein JA, Mason CE, Pesek J (2010) Dispersal and movement behavior of neonate European corn borer (Lepidoptera: Crambidae) on non-Bt and Transgenic Bt corn. J Econ Entomol 103:331–339
- Gore J, Leonard BR, Church GE, Cook DR (2002a) Behavior of bollworm (Lepidoptera: Noctuidae) larvae on genetically engineered cotton. J Econ Entomol 95:763–769
- Gore J, Leonard R, Church G (2002b) Bollworm larval behavior on Bollgard cotton: findings may change scouting procedures. Louisiana Agric 45:7–9
- Gould F (1998) Sustainability of transgenic insecticidal cultivars: integrating pest genetics and ecology. Annu Rev Entomol 43:701–726
- Greggers U, Menzel R (1993) Memory dynamics and foraging strategies of honeybees. Behav Ecol Sociobiol 32:17–29
- Hagenbucher S, Wäckers FL, Wettestein FE, Olson DM, Ruberson JR, Romeis J (2013) Pest trade-offs in technology: reduced damage by caterpillars in Bt cotton benefits aphids. Proc R Soc B 280:20130042
- Hågvar EB, Hofsvang T (1991) Aphid parasitoids (Hymenoptera, Aphidiidae): biology, host selection and use in biological control. Biocontrol News Inf 12:13–41
- Halcomb JL, Benedict JH, Cook B, Ring DR, Correa JC (2000) Feeding behavior of bollworm and tobacco budworm (Lepidoptera: Noctuidae) larvae in mixed stands of nontransgenic and transgenic cotton expressing an intsecticidal protein. J Econ Entomol 93:1300–1307
- Han P, Niu CY, Lei CL, Cui JJ, Desneux N (2010a) Use of an innovative T-tube maze assay and the Proboscis Extension Response assay to assess sublethal effects of GM products and pesticides on learning capacity of the honey bee *Apis mellifera* L. Ecotoxicology 19:1612–1619
- Han P, Niu CY, Lei CL, Cui JJ, Desneux N (2010b) Quantification of toxins in a Cry1Ac + CpTI cotton cultivar and its potential effects on the honey bee *Apis mellifera* L. Ecotoxicology 19:1452–1459
- Han P, Niu CY, Biondi A, Desneux N (2012) Does transgenic Cry1Ac + CpTI cotton pollen affect hypopharyngeal gland development and midgut proteolytic enzyme activity in the



honey bee *Apis mellifera* L. (Hymenoptera Apidae)? Ecotoxicology 21:2214–2221

- Han P, Niu CY, Desneux N (2014) Identification of top-down forces regulating cotton aphid population growth in transgenic Bt cotton in Central China. PLoS One 9(8):e102980. doi:10.1371/ journal.pone.0102980
- Hansen Jesse LC, Obrycki JJ (2000) Field deposition of Bt transgenic corn pollen: lethal effects on the monarch butterfly. Oecologia 125:241–248
- Hardke JT, Leonard BR, Temple JH (2012) Fall armyworm oviposition on cotton plants expressing wide strike(TM) Bollgar (R), and Bollgard II (R) Cry proteins. Southwest Entomol 37:295–303
- Harwood JD, Desneux N, Yoo HYS, Rowley D, Greenstone MH, Obrycki JJ et al (2007) Tracking the role of alternative prey in soybean aphid predation by *Orius insidiosus*: a molecular approach. Mol Ecol 16:4390–4400
- Haslberger AG (2003) Codex guidelines for GM foods include the analysis of unintended effects. Nat Biotechnol 21:739–741
- Haydak MH (1970) Honey bee nutrition. Annu Rev Entomol 15:143–156
- Heimpel GE, Casas J (2007) Parasitoid foraging and oviposition behavior in the field. In: Wajnberg E, Bernstein C, van Alphen J (eds) Behavioral ecology of insects parasitoids, from theoretical approaches to field applications. Wiley, New York, pp 51–70
- Hellmich RL, Siegfried BD, Sears MK, Stanley-Horn DE, Daniels MJ, Mattila HR, Spencer T, Bidne KG, Lewis LC (2001) Monarch larvae sensitivity to *Bacillus thuringiensis*-purified proteins and pollen. Proc Natl Acad Sci 98:11925–11930
- Hendriksma HP, Hartel S, Steffan-Dewenter I (2011) Testing pollen of single and stacked insect-resistant Bt-maize on in vitro reared honey bee larvae. PLoS One 6(12):e28174. doi:10.1371/journal.pone.0028174
- Hilbeck A, Baumgartner M, Fried PM, Bigler F (1998) Effects of transgenic *Bacillus thuringiensis* corn-fed prey on mortality and development time of immature *Chrysoperla carnea* (Neuroptera: Chrysopidae). Environ Entomol 27:480–487
- Hilbeck A, McMillan JM, Meier M, Humbel A, Schlaepfer-Miller J, Trtikova M (2012) A controversy re-visited: is the coccinellid Adalia bipunctata adversely affected by Bt toxins? Environ Sci Europe 24:10
- Himanen SJ, Nerg A, Nissinen A, Pinto DM, Stewart CNJr, Poppy GM et al (2009) Effects of elevated carbon dioxide and ozone on volatile terpenoid emissions and multitrophic communication of transgenic insecticidal oilseed rape (*Brassica napus*). New Phytol 181:174–186
- Jackson RE, Gore J, Abel C (2010) Bollworm (Lepidoptera: Noctuidae) behavior on transgenic cotton expressing Cry1Ac and Cry1F proteins. J Entomol Sci 45:252–261
- Jallow MFA, Hoy CW (2007) Indirect selection for increased susceptibility to permethrin in diamondback moth (Lepidoptera: Plutellidae). J Econ Entomol 100:526–533
- James (2014) Global Status of Commercialized Biotech/GM Crops: 2014. ISAAA Brief No. 49. ISAAA, Ithaca
- Johnson RM, Ellis MD, Mullin CA, Frazier M (2010) Pesticides and honey bee toxicity-USA. Apidologie 41:312–331
- Kessler A, Baldwin IT (2001) Defensive function of herbivoreinduced plant volatile emissions in nature. Science 291:2141– 2144
- Kumar H (2004) Orientation, feeding and ovipositional behavior of Diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae), on transgenic cabbage expressing Cry1Ab toxin of *Bacillus thuringiensis* (Berliner). Environ Entomol 33:1025–1031
- Landis DA, Wratten SD, Gurr GM (2000) Habitat management to conserve natural enemies of arthropod pests in agriculture. Annu Rev Entomol 45:175–201

- Lei Z, Liu TX, Greenberg SM (2009) Feeding, oviposition and survival of *Liriomyza trifolii* (Diptera: Agromyzidae) on Bt and non-Bt cottons. Bull Entomol Res 99:253–261
- Lewis WJ, Tumlinson JH (1988) Host detection by chemically mediated associative learning in a parasitoid wasp. Nature 331:257–259
- Li YH, Romeis J (2010) Bt maize expressing Cry3Bb1 does not harm the spider mite, *Tetranychus urticae*, or its ladybird beetle predator, *Stethorus punctillum*. Biol Control 53:337–344
- Li YX, Greenberg SM, Liu TX (2007) Orientation behavior, development and survival of *Trichoplusia ni* (Lepidoptera: Noctuidae) larvae on cotton expressing Cry1Ac and Cry2Ab and conventional cotton. J Insect Behav 20:473–488
- Li GP, Feng HQ, Chen PY, Wu SY, Liu B, Qiu F (2010) Effects of transgenic Bt cotton on the population density, oviposition behavior, development, and reproduction of a nontarget pest, *Adelphocoris suturalis* (Hemiptera: Miridae). Environ Entomol 39:1378–1387
- Li K, Tian J, Wang QX, Chen Q, Chen M, Wang H et al (2011) Application of a novel method PCR-ligase detection reaction for tracking predator-prey trophic links in insect-resistant GM rice ecosystem. Ecotoxicology 20:2090–2100
- Lima MAP, Pires CSS, Guedes RNC, Nakasu EYT, Lara MS, Fontes EMG et al (2011) Does Cry1Ac Bt-toxin impair development of worker larvae of Africanized honey bee? J Appl Entomol 135:415–422
- Liu XD, Zhai BP, Zhang XX, Zong JM (2005) Impact of transgenic cotton plants on a non-target pest, Aphis gossypii Glover. Ecol Entomol 30:307–315
- Liu B, Shu C, Xue K, Zhou KX, Li XG, Liu DD et al (2009) The oral toxicity of the transgenic Bt-CpTI cotton pollen to honey bees (*Apis mellifera*). Ecotoxicol Environ Saf 72:1163–1169
- Liu XX, Chen M, Onstad D, Roush R, Shelton AM (2011) Effect of Bt broccoli and resistant genotype of *Plutella xylostella* (Lepidoptera: Plutellidae) on development and host acceptance of the parasitoid *Diadegma insulare* (Hymenoptera: Ichneumonidae). Transgenic Res 20:887–897
- Liu QS, Romeis J, Yu HL, Zhang YJ, Li YH, Peng YF (2015) Bt rice does not disrupt the host-searching behavior of the parasitoid Cotesia chilonis. Sci Rep 5:15295
- Loivamäki M, Mumm R, Dicke M, Schnitzler JP (2008) Isoprene interferes with the attraction of bodyguards by herbaceous plants. Proc Natl Acad Sci 105:17430–17435
- Losey JE, Rayor LS, Carter ME (1999) Transgenic pollen harms monarch larvae. Nature 399:214
- Lu B, Downes S, Wilson L, Gregg P, Knight K, Kauter G et al (2011) Preferences of field bollworm larvae for cotton plant structures: impact of Bt and history of survival on Bt crops. Entomol Exp Appl 140:17–27
- Lu YH, Wu KM, Jiang YY, Guo YY, Desneux N (2012) Widespread adoption of Bt cotton and insecticide decrease promotes biocontrol services. Nature 487:362–365
- Ludy C, Lang A (2006a) Bt maize pollen exposure and impact on the garden spider, *Araneus diadematus*. Entomol Exp Appl 118:145–156
- Ludy C, Lang A (2006b) A 3-year field-scale monitoring of foliagedwelling spiders (Araneae) in transgenic Bt maize fields and adjacent field margins. Biol Control 38:314–324
- Lundgren JG, Wiedenmann RN (2002) Coleopteran-specific Cry3Bb toxin from transgenic corn pollen does not affect the fitness of a nontarget species, *Coleomegilla maculata* DeGeer (Coleoptera: Coccinellidae). Environ Entomol 31:1213–1218
- Lundgren JG, Wiedenmann RN (2005) Tritrophic interactions among Bt (Cry3Bb1) corn, aphid prey, and the predator *Coleomegilla maculata* (Coleoptera: Coccinellidae). Environ Entomol 34:1621–1625



Lundgren JG, Fergen JK, Riedell WE (2008) The influence of plant anatomy on oviposition and reproductive success of the omnivorous bug, *Orius insidiosus*. Anim Behav 75:1495–1502

- Lundgren JG, Gassmann AJ, Bernal J, Duan JJ, Ruberson J (2009) Ecological compatibility of GM crops and biological control. Crop Prot 28:1017–1030
- Luong TTA, Downes SJ, Cribb B, Perkins LE, Zalucki MP (2016) Oviposition site selection and survival of susceptible and resistant larvae of *Helicoverpa armigera* (Lepidoptera: Noctuidae) on Bt and non-Bt cotton. Bull Entomol Res. doi:10.1017/ S0007485316000328
- López C, Hernández-Escareño G, Eizaguirre M, Albajes R (2013) Antixenosis and larval and adult dispersal in the Mediterranean corn borer, *Sesamia nonagrioides*, in relation to Bt maize. Entomol Exp Appl 149:256–264
- Lövei GL, Arpaia S (2004) The impact of transgenic plants on natural enemies: a critical review of laboratory studies. Entomol Exp Appl 114:1–14
- Lövei GL, Andow DA, Arpaia S (2009) Transgenic insecticidal crops and natural enemies: a detailed review of laboratory studies. Environ Entomol 38:293–306
- Mallet J, Porter P (1992) Preventing insect adaptation to insectresistant crops: are seed mixes or refuge the best stratege? Proc R Soc Lond B Biol 250:165–169
- Malone LA, Burgess EPJ (2009) Impact of genetically modified crops on pollinators. In: Gatehouse AMR, Ferry N (eds) Environmental Impact of Genetically Modified Crops. CAB International, Wallingford, pp 199–222
- Malone LA, Pham-Delègue MH (2001) Effects of transgene products on honey bees (*Apis mellifera*) and bumblebees (*Bombus* sp.). Apidologie 32:287–304
- Malone LA, Burgess EPJ, Gatehouse HS, Voisey CR, Tregidga EL, Philip B (2001) Effects of ingestion of a *Bacillus thuringiensis* toxin and a trypsin inhibitor on honey bee flight activity and longevity. Apidologie 32:57–68
- Marquardt PT, Kruple CH (2009) Dispersal and mating behavior of Diabrotica virgifera virgifera (Coleoptera: Chrysomelidae). Environ Entomol 38:176–182
- Marvier M, McCreedy C, Regetz J, Kareiva P (2007) A meta-analysis of effects of Bt cotton and maize on nontarget invertebrates. Science 316:1475–1477
- Meier M, Hilbeck A (2001) Influence of transgenic Bacillus thuringiensis corn-fed prey on prey preference of immature *Chrysoperla carnea* (Neuroptera: Chrysopidae). Basic Appl Ecol 2:35–44
- Meiners T, Wackers F, Lewis WJ (2003) Associative learning of complex odours in parasitoid host location. Chem Senses 28:231–236
- Meissle M, Romeis J (2009) The web-building spider *Theridion impressum* (Araneae: Theridiidae) is not adversely affected by Bt maize resistant to corn rootworms. Plant Biotech J 7:645–656
- Meissle M, Vojtech E, Poppy GM (2005) Effects of Bt maize-fed prey on the generalist predator *Poecilus cupretis* L. (Coleoptera: Carabidae). Transgenic Res 14:123–132
- Men X, Ge F, Yardim EN, Parajulee MN (2005) Behavioral response of *Helicoverpa armigera* (Lepidoptera: Noctuidae) to cotton with and without expression of the Cry1Ac δ-Endotoxin protein of *Bacillus thuringiensis* Berlier. J Insect Behav 18:33–50
- Mommaerts V, Reynders S, Boulet J, Besard L, Sterk G, Smagghe G (2009) Risk assessment for side-effects of neonicotinoids against bumblebees with and without impairing foraging behavior. Ecotoxicology 19:207–215
- Mommaerts V, Jans K, Smagghe G (2010) Impact of Bacillus thuringiensis strains on survival, reproduction and foraging behaviour in bumblebees (*Bombus terrestris*). Pest Manag Sci 66:520–525

- Moraes MCB, Laumann RA, Aquino MFS, Paula DP, Borges M (2011) Effect of Bt genetic engineering on indirect defense in cotton via a tritrophic interaction. Transgenic Res 20:99–107
- Moser SE, Harwood JD, Obrycki JJ (2008) Larval feeding on Bt hybrid and non-Bt corn seedlings by *Harmonia axyridis* (Coleoptera: Coccinellidae) and *Coleomegilla maculata* (Coleoptera: Coccinellidae). Environ Entomol 37:525–533
- Naranjo (2009) Impacts of Bt crops on non-target invertebrates and insecticide use patterns. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources 2009 (4), No. 011
- Niu L, Ma Y, Mannakkara A, Zhao Y, Ma WH, Lei CL (2013) Impact of single and stacked insect-resistant Bt-cotton on the honey bee and silkworm. PLoS One 9:e72988. doi:10.1371/journal.pone. 0072988
- O'Callaghan M, Glare TR, Burgess EPJ, Malone LA (2005) Effects of plants genetically modified for insect resistance on nontarget organisms. Annu Rev Entomol 50:271–292
- Obonyo DN, Songa JM, Oyieke FA, Nyamasyo GHN, Mugo SN (2008) Bt-transgenic maize does not deter oviposition by two important African cereal stem borers, *Chilo partellus* Swinhoe (Lepidoptera: Crambidae) and *Sesamia calamistis* Hampson. J Appl Biosci 10:424–433
- Obrist LB, Klein H, Dutton A, Bigler F (2005) Effects of Bt maize on Frankliniella tenuicornis and exposure of thrips predators to prey-mediated Bt toxin. Entomol Exp Appl 115:409–416
- Obrist LB, Klein H, Dutton A, Bigler F (2006) Assessing the effects of Bt maize on the predatory mite *Neoseiulus cucumeris*. Exp Appl Acarol 38:125–139
- Onstad DW (2008) Major issues in insect resistance management. In:
  Onstad DW (ed) Insect resistance management: biology,
  economics and prediction. Academic, San Diego, pp 1–16
- Onstad DW, Liu X, Chen M, Roush R, Shelton AM (2013) Modeling the integration of parasitoid, insecticide, and transgenic insecticidal crop for the long-term control of an insect pest. J Econ Entomol 106:1103–1111
- Orr DB, Landis DA (1997) Oviposition of European corn borer (Lepidoptera: Pyralidae) and impact of natural enemy populations in transgenic versus isogenic corn. J Econ Entomol 90:905–909
- Pareja M, Mohib A, Birkett MA, Dufour S, Glinwood RT (2009) Multivariate statistics coupled to generalized linear models reveal complex use of chemicals cues by parasitoid. Anim Behav 77:901–909
- Park JR, McFarlane I, Phipps RH, Ceddia G (2011) The role of transgenic crops in sustainable development. Plant Biotechnol J 9:2–21
- Peterson JA, Lundgren JG, Harwood JD (2011) Interactions of transgenic *Bacillus thuringiensis* insecticidal crops with spiders (Aranae). J Arachnol 39:1–21
- Petzold-Maxwell JL, Cibils-Stewart X, French BW, Gassmann AJ (2012) Adaptation by Western corn rootworm (Coleoptera: Chrysomelidae) to Bt maize: inheritance, fitness, costs, and feeding preference. J Econ Entomol 105:1407–1418
- Pham-Delègue MH, Girard C, Le Metayer M, Picard-Nizou AL, Hennequet C, Pons O et al (2000) Long-term effects of soybean protease inhibitors on digestive enzymes, survival and learning abilities of honeybees. Entomol Exp Appl 95:21–29
- Picard-Nizou AL, Pham-Delègue HM, Kerguelen V, Douault P, Marilleau R, Olsen L et al (1995) Foraging behaviour of honey bees (*Apis mellifera* L.) on transgenic oilseed rape (*Brassica napus* L. var. oleifera). Transgenic Res 4:270–276
- Picard-Nizou AL, Grison R, Olsen L, Pioche C, Arnold G, Pham-Delègue HM (1997) Impact of proteins used in plant genetic engineering: toxicity and behavioral study in the honeybee. J Econ Entomol 90:1710–1716



- Pierre J, Marsault D, Genecque E, Renard M, Champolivier J, Pham-Delègue MH (2003) Effects of herbicide-tolerant transgenic oilseed rape genotypes on honey bees and other pollinating insects under field conditions. Entomol Exp Appl 108:159–168
- Pilcher CD, Obrycki JJ, Rice ME, Lewis LC (1997) Preimaginal development, survival, and field abundance of insect predators on GM Bacillus thuringiensis corn. Environ Entomol 26:446–454
- Prager SM, Martini X, Guvvala H, Nansen C, Lundgren J (2014) Spider mite infestations reduce *Bacillus thuringiensis* toxin concentration in corn leaves and predators avoid spider mites that have fed on *Bacillus thuringiensis* corn. Ann Appl Biol 165:108–116
- Prasifka PL, Hellmich RL, Prasifka JR, Lewis LC (2007) Effects of Cry1Ab-expressing corn anthers on the movement of monarch butterfly larvae. Environ Entomol 36:228–233
- Prütz G, Dettner K (2004) Effect of Bt corn leaf suspension on food consumption by *Chilo partellus* and life history parameters of its parasitoid *Cotesia flavipes* under laboratory conditions. Entomol Exp Appl 111:179–187
- Rader R, Bartomeus I, Garibaldi LA, Garratt MPD, Howlett BG, Winfree R et al (2016) Non-bee insects are important contributors to global crop pollination. Proc Natl Acad Sci 113:146–151
- Ramachandran S, Buntin GD, All JN, Raymer PL, Stewart CN (1998a) Movement and survival of diamondback moth (Lepidoptera: Plutellidae) larvae in mixtures of nontransgenic and transgenic canola containing a cryIA (c) gene of *Bacillus thuringiensis*. Environ Entomol 27:649–656
- Ramachandran S, Buntin GD, All JN, Tabashnik BE, Raymer PL, Adang MJ et al (1998b) Survival, development, and oviposition of resistant diamondback moth (Lepidoptera: Plutellidae) on transgenic canola producing a *Bacillus thuringiensis* Toxin. J Econ Entomol 91:1239–1244
- Ramalho FS, Pachú JKS, Lira ACS, Malaquias JB, Zanuncio JC, Fernandes FS (2014) Feeding and dispersal behavior of the cotton leafworm, *Alabama argillacea* (Hübner) (Lepidoptera: Noctuidae), on Bt and non-Bt cotton: implications for evolution and resistance management. PLoS One 9(11):e111588. doi:10. 1371/journal.pone.0111588
- Ramirez-Romero R, Josette C, Pham-Delègue MH (2005) Effects of Cry1Ab protoxin, deltamethrin and imidacloprid on the foraging activity and the learning performances of the honeybee *Apis mellifera*, a comparative approach. Apidologie 36:601–611
- Ramirez-Romero R, Bernal JS, Chaufaux J, Kaiser L (2007) Impact assessment of Bt-maize on a moth parasitoid, *Cotesia marginiventris* (Hymenoptera: Braconidae), via host exposure to purified Cry1Ab protein or Bt-plants. Crop Prot 26:953–962
- Ramirez-Romero R, Desneux N, Decourtye A, Chaffiol A, Pham-Delègue MH (2008a) Does Cry1Ab protein affect learning performance of the honey bee *Apis mellifera* L. (Hymenoptera, Apidae)? Ecotoxicol Environ Saf 70:327–333
- Ramirez-Romero R, Desneux N, Chaufaux J, Kaiser L (2008b) Bt-maize effects on biological parameters of the non-target aphid Sitobion avenae (Homoptera: Aphididae) and Cry1Ab toxin detection. Pesticide Biochem Phys 91:110–115
- Rao NS, Rao PA (2008) Behavioral and physiological effects of Bt cotton on cotton bollworm, *Helicoverpa armigera* (Hub.). J Entomol Res 32:273–277
- Raybould A (2006) Problem formulation and hypothesis testing for environmental risk assessments of genetically modified crops. Environ Biosaf Res 5:119–125
- Raybould A, Stacey D, Vlachos D, Graser G, Li X, Joseph R (2007) Non-target organisms risk assessment of MIR604 maize expressing Cry3A for control of corn rootworms. J Appl Entomol 131:391–399

- Razze JM, Mason CE (2012) Dispersal behavior of neonate European corn borer (Lepidoptera: Crambidae) on Bt corn. J Econ Entomol 105:1214–1223
- Riddick EW, Barbosa P (1998) Impact of Cry3A-intoxicated Leptinotarsa decemlineata (Coleoptera: Chrysomelidae) and pollen on consumption, development, and fecundity of Coleomegilla maculata (Coleoptera: Coccinellidae). Ann Entomol Soc Am 91:303–307
- Rojas RT, Gino FM, Roberto SH (2010) Spatial distribution of Aphis gossypii (Glover) (Hemiptera: Aphididae) and Bemisia tabaci (Gennadius) biotype B (Hemiptera, Aleyrodidae) on Bt and non-Bt cotton. Rev Bras Entomol 54:136–143
- Romeis J, Meissle M (2011) Non-target risk assessment of Bt crops— Cry protein uptake by aphids. J Appl Eotomol 135:1–6
- Romeis J, Meissle M, Bigler F (2006) GM crops expressing *Bacillus* thuringiensis toxins and biological control. Nat Biotechnol 24:63–71
- Romeis J, Bartsch D, Bigler F, Candolfi MP, Gielkens MMC, Hartley SE et al (2008) Assessment of risk of insect-resistant transgenic crops to non-target arthropods. Nat Biotechnol 26:203–208
- Romeis J, Hellmich RL, Candolfi MP, Carstens K, De Schrijver A, Gatehouse AMR et al (2011) Recommendations for the design of laboratory studies on non-target arthropods for risk assessment of genetically engineered plants. Transgenic Res 20:1–22
- Romeis J, Álvarez-Alfageme F, Bigler F et al (2012) Putative effects of Cry1Ab to larvae of Adalia bipunctata—reply to Hilbeck et al. (2012). Environ Sci Europe 24:18
- Rondeau G, Sánchez-Bayo F, Tennekes HA, Decourtye A, Ramírez-Romero R, Desneux N (2014) Delayed and time-cumulative toxicity of imidacloprid in bees, ants and termites. Sci Rep 4:5566
- Rose RI (2007) White paper on tier-based testing for the effects of proteinaceous insecticidal plant-incorporated protectants on non-target invertebrates for regulatory risk assessment. USDA-APHIS and US Environmental Protection Agency, Washington, DC
- Rovenská GZ, Zemek R (2006) Host plant preference of aphids, thrips and spider mites on GNA-expressing and control potatoes. Phytoparasitica 34:139–148
- Rovenská GZ, Zemek R, Schmidt JEU, Hilbeck A (2005) Altered host plant preference of *Tetranychus urticae* and prey preference of its predator *Phytoseiulus persimilis* (Acari: Tetranychidae, Phytoseiidae) on transgenic Cry3Bb-eggplants. Biol Control 33:293–300
- Sanders CJ, Pell JK, Poppy GM, Raybould A, Garcia-Alonso M, Schuler TH (2007) Host-plant mediated effects of GM maize on the insect parasitoid *Campoletis sonorensis* (Hymenoptera: Ichneumonidae). Biol Control 40:362–369
- Schmitz OJ, Krivan V, Ovadia O (2004) Trophic cascades: the primacy of trait-mediated indirect interactions. Ecol Lett 7:153–163
- Schone H, Strausfeld C (1984) Spatial orientation, the spatial control of behavior in animals and man. University Press, New Jersey
- Schoonhoven LM, van Loon JJA, Dicke M (2005) Insect-plant biology. Oxford University Press, Oxford
- Schroder R, Wurm L, Varama M, Meiners T, Hilker M (2008) Unusual mechanisms involved in learning of ovipositioninduced host plant odours in an egg parasitoid? Anim Behav 75:1423–1430
- Schuler TH, Potting RPJ, Denholm I, Poppy GM (1999) Parasitoid behaviour and Bt plants. Nature 400:825–826
- Schuler TH, Potting RPJ, Denholm I, Clark SJ, Clark AJ, Stewart CN et al (2003) Tritrophic choice experiments with Bt plants, the diamondback moth (*Plutella xylostella*) and the parasitoid *Cotesia plutellae*. Transgenic Res 12:351–361



Seagraves MP, Riedell WE, Lundgren JG (2011) Oviposition preference for water-stressed plants in *Orius insidiosus* (Hemiptera: Anthocoridae). J Insect Behav 24:132–143

- Setamou M, Bernal JS, Legaspi JC, Mirkov TE (2002) Parasitism and location of sugarcane borer (Lepidoptera: Pyralidae) by *Cotesia flavipes* (Hymenoptera: Braconidae) on transgenic and conventional sugarcane. Environ Entomol 31:1219–1225
- Shapiro JP, Ferkovich SM (2006) Oviposition and isolation of viable eggs from *Orius insidiosus* in aparafilm and water substrate: comparison with green beans and use in enzyme-linked immunosorbant assay. Ann Entomol Soc Am 99:586–591
- Spencer JL, Mabry TR, Vaughn TT (2003) Use of transgenic plants to measure insect herbivore movement. J Econ Entomol 96:1738–1749
- Stapel JO, Waters DJ, Ruberson JR, Lewis WJ (1998) Development and behavior of *Spodoptera exigua* (Lepidoptera: noctuidae) larvae in choice tests with food substrates containing toxins of *Bacillus thuringiensis*. Biol Control 11:29–37
- Storer NP, Babcock JM, Schlenz M, Meade T, Thompson GD, Bing JW et al (2010) Discovery and characterization of field resistance to Bt maize: Spodoptera frugiperda (Lepidoptera: Noctuidae) in Puerto Rico. J Econ Entomol 103:1031–1038
- Sun X, Zhou W, Liu H, Zhang A, Ai CR, Zhou SS et al (2013) Transgenic Bt rice does not challenge host preference of the target pest of rice leaffolder, *Cnaphalocrocis medinalis* (Lepidoptera: Pyralidae). PLoS One 8(11):e79032. doi:10.1371/ journal.pone.0079032
- Swamy SVSG, Sharma HC, Subbaratman GV, Vijay MP (2008) Ovipositional and feeding preferences of *Helicoverpa armigera* towards putative transgenic and non-transgenic pideonpeas. Resistant Pest Manag Newslett 17:50–52
- Symondson WOC, Sunderland KD, Greenstone MH (2002) Can generalist predators be effective biocontrol agents? Annu Rev Entomol 47:561–594
- Tabashnik BE, Carrière Y, Dennehy TJ, Morin S, Sisterson MS, Roush RT et al (2003) Insect resistance to transgenic *Bacillus thuringiensis* crops: lessons from the laboratory and field. J Econ Entomol 96:1031–1038
- Telléz-Rodríguez P, Raymond B, Morán-Bertot I, Rodríguez-Cabrera L, Wright DJ, Borroto CG et al (2014) Strong oviposition preference for Bt over non-Bt maize in *Spodoptera frugiperda* and its implications for the evolution of resistance. BMC Biol 12:48
- Thompson HM (2003) Behavioural effects of pesticides in bees-their potential for use in risk assessment. Ecotoxicology 12:317–330
- Toft S (1999) Prey choice and spider fitness. J Arachnol 27:301–307
  Tomov BW, Bernal JS, Vinson SB (2003) Impacts of transgenic sugarcane expressing GNA lectin on parasitism of Mexican rice borer by *Parallorhogas pyralophagus* (Marsh) (Hymenoptera: Braconidae). Environ Entomol 32:866–872
- Torres JB, Ruberson JR (2006) Interactions of Bt-cotton and the omnivorous big-eyed bug *Geocoris punctipes* (Say), a key predator in cotton fields. Biol Control 39:47–57
- Torres JA, Ruberson JR (2008) Interactions of *Bacillus thuringiensis* Cry1Ac toxin in genetically engineered cotton with predatory heteropterans. Transgenic Res 17:345–354
- Torres JB, Ruberson JR, Adang MJ (2006) Expression of *Bacillus thuringiensis* Cry1Ac protein in cotton plants, acquisition by pests and predators: a tritrophic analysis. Agric Forest Entomol 8:191–202
- Toschki A, Hothorn LA, Ross-Nickoll M (2007) Effects of cultivation of genetically modified Bt maize on epigeic arthropods (Araneae: Carabidae). Environ Entomol 36:967–981
- Tschenn J, Losey JE, Jesse LH, Obrycki JJ, Hufbauer R (2001) Effects of corn plants and corn pollen on monarch butterfly

- (Lepidoptera: Danaidae) oviposition behavior. Environ Entomol 30:495–500
- Turlings TCJ, Jeanbourquin PM, Held M, Degen T (2005) Evaluating the induced-odour emission of a Bt maize and its attractiveness to parasitic wasps. Transgenic Res 14:807–816
- Van den Berg J, Van Wik A (2007) The effect of Bt maize on Sesamia calamistis in South Africa. Entomol Exp Appl 122:45–51
- Vet LEM, Godfray HCJ (2007) Multitrophic interactions and parasitoid behavioural ecology. In: Wajnberg E, Bernstein C, Van Alphen J (eds) Behavioral ecology of insect parasitoids from theoretical approaches to field applications. Blackwell, Oxford, pp 231–252
- Vet LEM, Groenewold AW (1990) Semiochemicals and learning in parasitoids. J Chem Ecol 16:3119–3135
- Wajnberg E, Bernstein C, van Alphen J (2007) Behavioral ecology of insects parasitoids, from theoretical approaches to field applications. Wiley, New York
- Walker GP, Cameron PJ, MacDonald FM, Madhusudhan VV, Wallace AR (2007) Impacts of *Bacillus thuringiensis* toxins on parasitoids (Hymenoptera: Braconidae) of *Spodoptera litura* and *Helicoverpa armigera* (Lepidoptera: Noctuidae). Biol Control 40:142–151
- Wan P, Huang YX, Wu HH, Huang MS, Cong SB, Tabashnik BE et al (2012) Increased frequency of pink bollworm resistance to Bt toxin Cry1Ac in China. PLoS One 7(1):e29975. doi:10.1371/journal.pone.0029975
- Werner EE, Peacor SD (2003) A review of trait-mediated indirect interactions in ecological communities. Ecology 84:1083–1100
- Whitehouse MEA, Wilson LJ, Constable GA (2007) Target and nontarget effects on the invertebrate community of Vip cotton, a new insecticidal transgenic. Aust J Agric Res 58:273–285
- Wolfenbarger LL, Naranjo SE, Lundgren JG, Bitzer RJ, Watrud LS (2008) Bt crop effects on functional guilds of non-target arthropods: a meta-analysis. PLoS One 3(5):e2118. doi:10. 1371/journal.pone.0002118
- Zalucki MP, Cunningham JP, Downes S, Ward P, Lange C, Meissle M et al (2012) No evidence for change in oviposition behaviour of *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae) after widespread adoption of transgenic insecticidal cotton. Bull Entomol Res 102:468–476
- Zhang JH, Wang CZ, Qin JD, Guo SD (2004) Feeding behavior of Helicoverpa armigera larvae on insect-resistant transgenic cotton and non-transgenic cotton. J Appl Entomol 128:218–225
- Zhang GF, Wan FH, Lovei GL, Liu WX, Guo JY (2006) Transmission of Bt toxin to the predator *Propylaea japonica* (Coleoptera: Coccinellidae) through its aphid prey feeding on transgenic Bt cotton. Environ Entomol 35:143–150
- Zhang HN, Yin W, Zhao J, Jin L, Yang YH, Wu SW et al (2011) Early warning of cotton bollworm resistance associated with intensive planting of Bt cotton in China. PLoS One 6(8):e22874. doi:10.1371/journal.pone.0022874
- Zhao XC, Wu KM, Liang GM, Guo YY (2008) Altered mating behaviour in a Cry1Ac-resistant strain of *Helicoverpa armigera* (Lepidoptera: Noctuidae). J Appl Entomol 132:360–365
- Zhao XC, Wu KM, Liang GM, Guo YY (2009) Modified female calling behavior in Cry1Ac-resistant *Helicoverpa armigera* (Lepidoptera: Noctuidae). Pest Manag Sci 65:353–357
- Zwahlen C, Nentwig W, Bigler F, Hilbeck A (2000) Tritrophic interactions of transgenic *Bacillus thuringiensis* corn, *Anapho-thrips obscurus* (Thysanoptera: Thripidae), and the predator *Orius majusculus* (Heteropera: Anthocoridae). Environ Entomol 29:846–850

