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Revision of the Nomenclature for the *Bacillus thuringiensis* Pesticidal Crystal Proteins

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BACKGROUND AND HISTORY OF PESTICIDAL CRYSTAL PROTEIN NOMENCLATURE

Since the first cloning of an insecticidal crystal protein gene from *Bacillus thuringiensis* (91), many other such genes have been isolated. Initially, each newly characterized gene or protein received an arbitrary designation from its discoverers: *icp* (64); *cry* (21, 121); *kurhd1* (31); Bta (88); bt1, bt2, etc. (40); type B and type C (43); and 4.5 kb, 5.3 kb, and 6.6 kb (55). The first systematic attempt to organize the genetic nomenclature relied on the insecticidal activities of crystal proteins for the primary ranking of their corresponding genes (44). The *cryI* genes encoded proteins toxic to lepidopterans; *cryII* genes encoded proteins toxic to both lepidopterans and dipterans; *cryIII* genes encoded proteins toxic to coleopterans; and *cryIV* genes encoded proteins toxic to dipterans alone.

This system provided a useful framework for classifying the ever-expanding set of known genes. Inconsistencies existed in the original scheme, however, due to attempts to accommodate genes that were highly homologous to known genes but did not encode a toxin with a similar insecticidal spectrum. The *cryIIB* gene, for example, received a place in the lepidopteran-dipteran class with *cryIIA*, even though toxicity against dipterans could not be demonstrated for the toxin designated CryIIB. Other anomalies arose after the nomenclature was established. The protein named CryIC, for example, was reported to be toxic to both dipterans and lepidopterans (103), while the protein designated CryIB was reported to be toxic to both lepidopterans and coleopterans (8). Because the nomenclature system provided no central committee or database to maintain standardization, new genes encoding a diverse set of proteins without a common insecticidal activity each received the name *cryV*, based on the next available Roman numeral (32, 46, 67, 100, 102, 108).

PROPOSED NOMENCLATURE

We propose in this review a revised nomenclature for the *cry* and *cyt* genes. To organize the wealth of data produced by genomic sequencing efforts, a new nomenclatural paradigm is emerging, exemplified by the internationally recognized cyto-

chrome P-450 superfamily nomenclature system (68a, 122a). Our proposal conforms closely to this model both in conceptual basis and in nomenclature format. The underlying basis of this type of system is to assign names to members of gene superfamilies according to their degree of evolutionary divergence as estimated by phylogenetic tree algorithms. The nomenclature format in such a system is designed to convey rich informational content about these relationships by appending to the mnemonic root a series of numerals and letters assigned in a hierarchical fashion to indicate degrees of phylogenetic divergence. This change from a function-based to a sequence-based nomenclature allows closely related toxins to be ranked together and removes the necessity for researchers to bioassay each new protein against a growing series of organisms before assigning it a name.

In our proposed revision, Roman numerals have been exchanged for Arabic numerals in the primary rank (e.g., Cry1Aa) to better accommodate the large number of expected new proteins. The mnemonic Cyt to designate crystal proteins showing a general cytolytic activity *in vitro* has been retained because of its historical precedent and entrenchment in the research literature. Our definition of a Cry protein is rather broad: a parasporal inclusion (crystal) protein from *B. thuringiensis* that exhibits some experimentally verifiable toxic effect to a target organism, or any protein that has obvious sequence similarity to a known Cry protein. Similarly, Cyt denotes a parasporal inclusion (crystal) protein from *B. thuringiensis* that exhibits hemolytic activity, or any protein that has obvious sequence similarity to a known Cyt protein. By these criteria, the nontoxic 40-kDa crystal protein from *B. thuringiensis* subsp. *thompsoni*, for example, has been excluded from our list, but the lepidopteran-active 34-kDa protein (now Cry15A) encoded by an adjacent gene has been included (11).

The freely available software applications CLUSTAL W (110) and PHYLIP (27) define the sequence relationships among the toxins to form the framework of the new nomenclature. In the first step, CLUSTAL W aligns the deduced amino acid sequences of the full-length toxins and produces a distance matrix, quantitating the sequence similarities among the set of toxins. CLUSTAL W default settings are employed, except that the "delay divergent sequences" setting in the multiple-alignment parameter menu is reduced from 40 to 0%. The NEIGHBOR application within the PHYLIP package then constructs a phylogenetic tree from the distance matrix by an unweighted pair-group method using arithmetic averages

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(UPGMA) algorithm. The TREEVIEW application (73), with the “phylogenetic tree” and “ladderize left” options selected, produces a graphic presentation of the resulting tree.

We have applied this procedure to the set of holotype sequences given in Table 1 to produce the phylogenetic tree presented in Fig. 1. Vertical lines drawn through the tree show the boundaries used to define the various nomenclatural ranks. The name given to any particular toxin depends on the location of the node where the toxin enters the tree relative to these boundaries. A new toxin that joins the tree to the left of the leftmost boundary will be assigned a new primary rank (an Arabic number). A toxin that enters the tree between the left and central boundaries will be assigned a new secondary rank (an uppercase letter). It will have the same primary rank as the other toxins within that cluster. A toxin that enters the tree between the central and right boundaries will be assigned a new tertiary rank (a lowercase letter). Finally, a toxin that joins the tree to the right of the rightmost boundary will be assigned a new quaternary rank (another Arabic number). Toxins with identical sequences but isolated independently will receive separate quaternary ranks.

By this method each toxin will be assigned a unique name incorporating all four ranks. A completely novel toxin would currently be assigned the name Cry23Aa1. For the sake of convenience, however, we propose that the inclusion of the tertiary rank *a* and quaternary rank 1 be optional, their use dictated only by a need for clarity. This new toxin could therefore simply be referred to as Cry23A.

In choosing locations for rank boundaries, we attempted to construct a nomenclature reflecting significant evolutionary relationships while at the same time minimizing changes from the gene names assigned under the old system. In the resulting system, proteins with a common primary rank are similar enough that the percent identity can be defined with some confidence. Proteins with the same primary rank often affect the same order of insect; those with different secondary and tertiary ranks may have altered potency and targeting within an order. At the tertiary rank, differences can be due to the accumulation of dispersed point mutations, but often they appear to have resulted from ancestral recombination events between genes differing at a lower rank level (9). The quaternary rank was established to group “alleles” of genes coding for known toxins that differ only slightly, either because of a few mutational changes or an imprecision in sequencing. To avoid confusion, however, the reader should bear in mind the differences between the quaternary rank number and the classical concept of the allele. Any *cry* gene specified with a quaternary rank is a natural isolate. No assumption about functionality is implied by the presence of this rank number in the gene name. In contrast, an allele number would be assumed, unless parenthetical or subscripted information indicated otherwise, to denote a nonfunctional mutant form of a wild-type gene found at a discrete genetic locus. Because of the somewhat modular nature of the Cry proteins and the effect that various segmental relationships could have on the clustering algorithm, it is likely that these boundaries will move slightly or even bend as the addition of new sequences changes the topology of the phylogenetic tree. Currently the boundaries represent approximately 95, 78, and 45% sequence identity.

A *B. thuringiensis* Pesticidal Crystal Protein Nomenclature Committee, consisting of the authors of this paper, will remain as a standing committee of the *Bacillus* Genetic Stock Center (BGSC) to assist workers in the field of *B. thuringiensis* genetics in assigning names to new Cry and Cyt toxins. The corresponding gene or protein sequences must first be deposited into a publicly accessible database (GenBank, EMBL, or PIR) and

released by the repository for electronic publication in the database so that the scientific community may conduct an independent analysis. Researchers should submit new sequences directly to the BGSC director (D. R. Zeigler), either by electronic mail (zeigler.1@osu.edu) or on computer diskette. The director will analyze the amino acid sequence as described above and suggest the appropriate name, subject to the approval of the committee. The committee will periodically review the literature of the Cry and Cyt toxins and publish a comprehensive list. This list, alongside other relevant information, will also be available via the Internet at the following URL: http://www.biols.susx.ac.uk/Home/Neil_Crickmore/Bt/.

The current list of *cry* and *cyt* genes (including quaternary ranks) is given in Table 1. New gene names are listed with their previous names, their GenBank accession numbers, and published references. The quaternary ranks were assigned in the order that the gene sequences were discovered in the literature or submitted to the committee. Genes assigned the quaternary rank 1 represent holotype sequences.

The boundaries shown in Fig. 1 allow most *cry* genes to retain the names they received under the system of Höfte and Whiteley (44), after a substitution of Arabic for Roman numerals. There are a few notable exceptions: *cryIG* becomes *cry9A*, *cryIIIC* becomes *cry7Aa*, *cryIIID* becomes *cry3C*, *cryIVC* becomes *cry10A*, *cryIVD* becomes *cry11A*, *cytA* becomes *cyt1A*, and *cytB* becomes *cyt2A* (Table 1). Under the revised system, the known Cry and Cyt proteins fall into 24 sets at the primary rank—Cyt1, Cyt2, and Cry1 through Cry22.

ROBUSTNESS OF THE NOMENCLATURE

The robustness of the current naming process was assessed by a number of additional analyses. The choice of clustering algorithm (unweighted pair-group method using arithmetic averages) was driven largely by the consistent location of a root and constant branch lengths, resulting in a common vertical alignment of sequence names and essentially allowing a “ruler across the tree” approach to naming. It has the drawback of imposing a common evolutionary clock on the clustering process, an assumption that cannot be assured. The distance metric related to percent identity (essentially 1 minus the fraction of identical residues of the total compared without gaps) is the one most commonly found as the output of sequence comparison programs, including CLUSTAL W. For phylogenetic analysis, a more usual distance metric relates to the number of substitutions per site to convert one sequence to the other (e.g., Dayhoff’s point accepted mutation [PAM]) and accounts for the possibility of multiple substitutions per site as the sequences are more divergent. The latter method has the drawback of being more computationally intensive, and, for very divergent sequences, requiring too large a value, resulting in numeric computation failures. They also differ in the way sequences of unequal length are handled, with the percent identity method typically ignoring excess sequence and the other methods assigning a penalty. This is particularly important for crystal proteins, since a number of them lack the C-terminal protoxin segments yet are quite related to some longer toxins in the N-terminal toxin segment; we feel that the stronger association of such relationships found by the percent identity method is preferred.

To assess the effect of using the neighbor-joining method to generate an unrooted tree, CLUSTAL W routines were used to generate such a tree with 1,000 bootstraps of the sequence alignment we used for Fig. 1. When an appropriate outgroup was chosen, the resulting tree (not shown) resembled our Fig. 1. The bootstrap values indicated that the tree thus generated

TABLE 1. Known *cry* and *cyt* gene sequences with revised nomenclature assignments

Revised gene name	Original gene or protein name	Accession no.	Coding region ^a	Reference	Revised gene name	Original gene or protein name	Accession no.	2125–3990>	Reference
<i>cryIAa1</i>	<i>cryIA(a)</i>	M11250	527–4054	92	<i>cry2Ab2</i>	<i>cryIIB</i>	X55416	874–2775	17
<i>cryIAa2</i>	<i>cryIA(a)</i>	M10917	153–>2955	98	<i>cry2Ac1</i>	<i>cryIIC</i>	X57252	2125–3990	124
<i>cryIAa3</i>	<i>cryIA(a)</i>	D00348	73–3600	99	<i>cry3Aa1</i>	<i>cryIIIA</i>	M22472	25–1956	39
<i>cryIAa4</i>	<i>cryIA(a)</i>	X13535	1–3528	62	<i>cry3Aa2</i>	<i>cryIIIA</i>	J02978	241–2172	93
<i>cryIAa5</i>	<i>cryIA(a)</i>	D17518	81–3608	113	<i>cry3Aa3</i>	<i>cryIIIA</i>	Y00420	566–2497	41
<i>cryIAa6</i>	<i>cryIA(a)</i>	U43605	1–>1860	63	<i>cry3Aa4</i>	<i>cryIIIA</i>	M30503	201–2132	65
<i>cryIAb1</i>	<i>cryIA(b)</i>	M13898	142–3606	119	<i>cry3Aa5</i>	<i>cryIIIA</i>	M37207	569–2500	22
<i>cryIAb2</i>	<i>cryIA(b)</i>	M12661	155–3622	111	<i>cry3Aa6</i>	<i>cryIIIA</i>	U10985	569–2500	1
<i>cryIAb3</i>	<i>cryIA(b)</i>	M15271	156–3620	31	<i>cry3Ba1</i>	<i>cryIIIB2</i>	X17123	25–>1977	101
<i>cryIAb4</i>	<i>cryIA(b)</i>	D00117	163–3627	50	<i>cry3Ba2</i>	<i>cryIIIB</i>	A07234	342–2297	85
<i>cryIAb5</i>	<i>cryIA(b)</i>	X04698	141–3605	40	<i>cry3Bb1</i>	<i>cryIIIBb</i>	M89794	202–2157	24
<i>cryIAb6</i>	<i>cryIA(b)</i>	M37263	73–3537	37	<i>cry3Bb2</i>	<i>cryIIIC(b)</i>	U31633	144–2099	23
<i>cryIAb7</i>	<i>cryIA(b)</i>	X13233	1–3465	36	<i>cry3Ca1</i>	<i>cryIIID</i>	X59797	232–2178	59
<i>cryIAb8</i>	<i>cryIA(b)</i>	M16463	157–3621	69	<i>cry4Aa1</i>	<i>cryIVA</i>	Y00423	1–3540	121
<i>cryIAb9</i>	<i>cryIA(b)</i>	X54939	73–3537	13	<i>cry4Aa2</i>	<i>cryIVA</i>	D00248	393–3935	95
<i>cryIAb10</i>	<i>cryIA(b)</i>	A29125	— ^b	28	<i>cry4Ba1</i>	<i>cryIVB</i>	X07423	157–3564	16
<i>cryIaC1</i>	<i>cryIA(c)</i>	M11068	388–3921	3	<i>cry4Ba2</i>	<i>cryIVB</i>	X07082	151–3558	112
<i>cryIaC2</i>	<i>cryIA(c)</i>	M35524	239–3769	117	<i>cry4Ba3</i>	<i>cryIVB</i>	M20242	526–3930	125
<i>cryIaC3</i>	<i>cryIA(c)</i>	X54159	339–>2192	18	<i>cry4Ba4</i>	<i>cryIVB</i>	D00247	461–3865	95
<i>cryIaC4</i>	<i>cryIA(c)</i>	M73249	1–3534	84	<i>cry5Aa1</i>	<i>cryVA(a)</i>	L07025	1–>4155	102
<i>cryIaC5</i>	<i>cryIA(c)</i>	M73248	1–3531	83	<i>cry5Ab1</i>	<i>cryVA(b)</i>	L07026	1–>3867	67
<i>cryIaC6</i>	<i>cryIA(c)</i>	U43606	1–>1821	63	<i>cry5Ac1</i>		I34543	1–>3660	76
<i>cryIaC7</i>	<i>cryIA(c)</i>	U87793	976–4509	38	<i>cry5Ba1</i>	PS86Q3	U19725	1–>3735	76
<i>cryIaC8</i>	<i>cryIA(c)</i>	U87397	153–3686	71	<i>cry6Aa1</i>	<i>cryVIA</i>	L07022	1–>1425	68
<i>cryIaC9</i>	<i>cryIA(c)</i>	U89872	388–3921	33	<i>cry6Ba1</i>	<i>cryVIB</i>	L07024	1–>1185	67
<i>cryIaC10</i>	<i>cryIA(c)</i>	AJ002514	388–3921	107	<i>cry7Aa1</i>	<i>cryIIIC</i>	M64478	184–3597	58
<i>cryIAd1</i>	<i>cryIA(c)</i>	M73250	1–3537	79	<i>cry7Ab1</i>	<i>cryIIIC(b)</i>	U04367	1–>3414	75
<i>cryIAe1</i>	<i>cryIA(e)</i>	M65252	81–3623	60	<i>cry7Ab2</i>	<i>cryIIIC(c)</i>	U04368	1–>3414	75
<i>cryIAf1</i>	<i>icp</i>	U82003	172–>2905	49	<i>cry8Aa1</i>	<i>cryIIIE</i>	U04364	1–>3471	29
<i>cryIBa1</i>	<i>cryIB</i>	X06711	1–3684	10	<i>cry8Ba1</i>	<i>cryIIIG</i>	U04365	1–>3507	66
<i>cryIBa2</i>		X95704	186–3869	105	<i>cry8Ca1</i>	<i>cryIIIF</i>	U04366	1–3447	70
<i>cryIBb1</i>	ET5	L32020	67–3753	25	<i>cry9Aa1</i>	<i>cryIG</i>	X58120	5807–9274	104
<i>cryIBc1</i>	<i>cryIB(c)</i>	Z46442	141–3839	6	<i>cry9Aa2</i>	<i>cryIG</i>	X58534	385–>3837	32
<i>cryIBd1</i>	<i>cryE1</i>	U70726		12	<i>cry9Ba1</i>	<i>cryX</i>	X75019	26–3488	97
<i>cryICa1</i>	<i>cryIC</i>	X07518	47–3613	45	<i>cry9Ca1</i>	<i>cryIH</i>	Z37527	2096–5569	57
<i>cryICa2</i>	<i>cryIC</i>	X13620	241–>2711	88	<i>cry9Da1</i>	N141	D85560	47–3553	4
<i>cryICa3</i>	<i>cryIC</i>	M73251	1–3570	79	<i>cry9Da2</i>		AF042733	<1–>1937	122
<i>cryICa4</i>	<i>cryIC</i>	A27642	234–3800	114	<i>cry10Aa1</i>	<i>cryIVC</i>	M12662	941–2965	111
<i>cryICa5</i>	<i>cryIC</i>	X96682	1–>2268	106	<i>cry11Aa1</i>	<i>cryIVD</i>	M31737	41–1969	21
<i>cryICa6</i>	<i>cryIC</i>	X96683	1–>2268	106	<i>cry11Aa2</i>	<i>cryIVD</i>	M22860	<1–235	2
<i>cryICa7</i>	<i>cryIC</i>	X96684	1–>2268	106	<i>cry11Ba1</i>	Jeg80	X86902	64–2238	19
<i>cryICb1</i>	<i>cryIC(b)</i>	M97880	296–3823	48	<i>cry11Bb1</i>	94 kDa	AF017416		72
<i>cryIDa1</i>	<i>cryID</i>	X54160	264–3758	42	<i>cry12Aa1</i>	<i>cryVB</i>	L07027	1–>3771	67
<i>cryIDb1</i>	<i>prtB</i>	Z22511	241–3720	56	<i>cry13Aa1</i>	<i>cryVC</i>	L07023	1–2409	90
<i>cryIEa1</i>	<i>cryIE</i>	X53985	130–3642	115	<i>cry14Aa1</i>	<i>cryVD</i>	U13955	1–3558	77
<i>cryIEa2</i>	<i>cryIE</i>	X56144	1–3513	7	<i>cry15Aa1</i>	34kDa	M76442	1036–2055	11
<i>cryIEa3</i>	<i>cryIE</i>	M73252	1–3513	82	<i>cry16Aa1</i>	<i>cbm71</i>	X94146	158–1996	5
<i>cryIEa4</i>		U94323	388–3900	47	<i>cry17Aa1</i>	<i>cbm72</i>	X99478	12–1865	5
<i>cryIEb1</i>	<i>cryIE(b)</i>	M73253	1–3522	81	<i>cry18Aa1</i>	<i>cryBPI</i>	X99049	743–2860	126
<i>cryIFa1</i>	<i>cryIF</i>	M63897	478–3999	14	<i>cry19Aa1</i>	Jeg65	Y07603	719–2662	86
<i>cryIFa2</i>	<i>cryIF</i>	M73254	1–3525	80	<i>cry19Ba1</i>		D88381		87
<i>cryIFb1</i>	<i>prtD</i>	Z22512	483–4004	56	<i>cry20Aa1</i>	86kDa	U82518	60–2318	61
<i>cryIGa1</i>	<i>prtA</i>	Z22510	67–3564	56	<i>cry21Aa1</i>		I32932	1–3501	74
<i>cryIGa2</i>	<i>cryIM</i>	Y09326	692–4210	96	<i>cry22Aa1</i>		I34547	1–2169	76
<i>cryIGb1</i>	<i>cryH2</i>	U70725		12					
<i>cryIHa1</i>	<i>prtC</i>	Z22513	530–4045	56					
<i>cryIhb1</i>		U35780	728–4195	53					
<i>cryIIa1</i>	<i>cryV</i>	X62821	355–2511	108					
<i>cryIIa2</i>	<i>cryV</i>	M98544	1–2157	34	<i>cyt1Aa1</i>	<i>cytA</i>	X03182	140–886	118
<i>cryIIa3</i>	<i>cryV</i>	L36338	279–2435	100	<i>cyt1Aa2</i>	<i>cytA</i>	X04338	509–1255	120
<i>cryIIa4</i>	<i>cryV</i>	L49391	61–2217	54	<i>cyt1Aa3</i>	<i>cytA</i>	Y00135	36–782	26
<i>cryIIa5</i>	<i>cryV159</i>	Y08920	524–2680	94	<i>cyt1Aa4</i>	<i>cytA</i>	M35968	67–813	30
<i>cryIIb1</i>	<i>cryV465</i>	U07642	237–2393	100	<i>cyt1Ab1</i>	<i>cytM</i>	X98793	28–777	109
<i>cryIIa1</i>	ET4	L32019	99–3519	25	<i>cyt1Ba1</i>		U37196	1–795	78
<i>cryIIb1</i>	ET1	U31527	177–3686	116	<i>cyt2Aa1</i>	<i>cytB</i>	Z14147	270–1046	51
<i>cryIKa1</i>		U28801	451–4098	52	<i>cyt2Ba1</i>	“ <i>cytB</i> ”	U52043	287–655	35
<i>cry2Aa1</i>	<i>cryIIA</i>	M31738	156–2054	20	<i>cyt2Bb1</i>		U82519	416–1204	15
<i>cry2Aa2</i>	<i>cryIIA</i>	M23723	1840–3738	123					
<i>cry2Aa3</i>		D86064	2007–3911	89					
<i>cry2Ab1</i>	<i>cryIIB</i>	M23724	1–1899	123					

^a The symbols < and > indicate that the coding region extends up- or downstream, respectively, from the known sequence data.^b Only the polypeptide sequence has been reported.

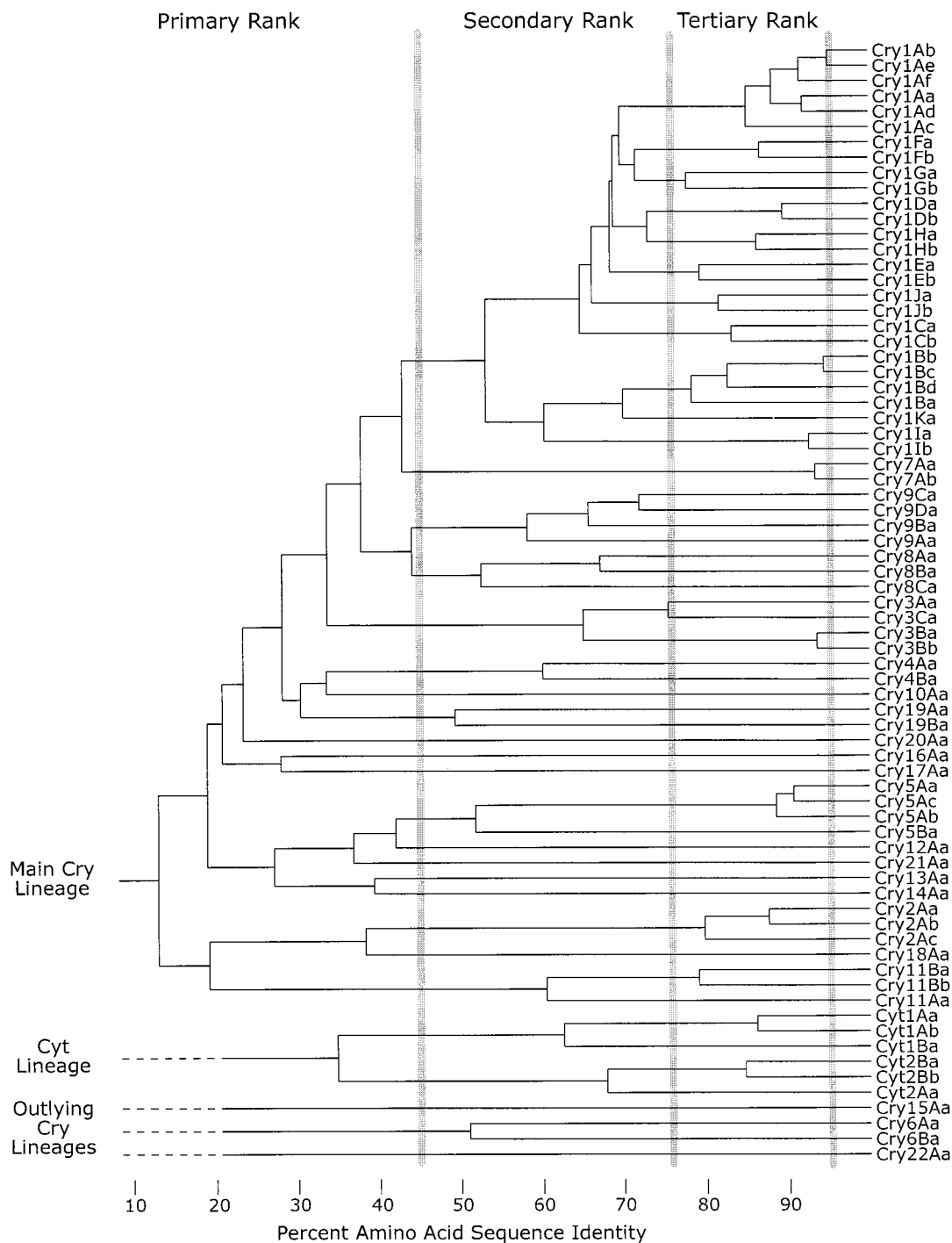


FIG. 1. Phylogram demonstrating amino acid sequence identity among Cry and Cyt proteins. This phylogenetic tree is modified from a TREEVIEW visualization of NEIGHBOR treatment of a CLUSTAL W multiple alignment and distance matrix of the full-length toxin sequences, as described in the text. The gray vertical bars demarcate the four levels of nomenclature ranks. Based on the low percentage of identical residues and the absence of any conserved sequence blocks in multiple-sequence alignments, the lower four lineages are not treated as part of the main toxin family, and their nodes have been replaced with dashed horizontal lines in this figure.

had significant branch points deeper in the tree than the chosen primary rank in the nomenclature. This sort of analysis was rejected as unsuitable for the purposes of Cry nomenclature due to the generally ragged branch lengths it produced and the requirement for the careful choice of an outgroup.

An alternative method of clustering protein sequences, ca-

pable of handling sequences that are quite diverse, is parsimony analysis. A consensus tree generated from 100 bootstraps of such an analysis displaces the two incomplete Cry1 sequences (Cry1Bd and Cry1Af) and the two Cry1 sequences lacking the C-terminal protoxin segments (Cry1Ia and Cry1Ib) into a region of the tree populated with such shortened se-

quences (not shown). With the further exceptions of Cry12A being interjected into the Cry5 cluster and a number of sequences besides Cry6B clustering higher in the tree than Cry6A, the proposed nomenclature successfully reflects the grouping of sequences provided by this method of analysis as well.

As noted above, the usual distance metrics for phylogenetic analysis account for multiple substitutions per site; most commonly, the Dayhoff PAM metric is used. When this distance metric was applied to the alignment used to make Fig. 1, a large number of the sequence pairs were found to have infinite distance. Therefore, the main Cry lineage and the Cyt lineage were separately aligned, the distances were calculated, and the distance matrices were clustered by using the FITCH program (of the PHYLIP software package). This method of analysis revealed several strongly associated groups of sequences (>90% of trees) in the main Cry lineage that extend deeper into the tree than the primary rank assigned in the proposed nomenclature: Cry1; Cry3; Cry4; Cry7; the Cry5, Cry12-Cry13-Cry14-Cry21 group; the Cry8-Cry9 group; the Cry10-Cry19 group; the Cry16-Cry17 group; and the Cry2-Cry11-Cry18 group. Many of these groups, however, were separated by branch points that were either nonmajority or were found <60% of the time; thus, the arrangement of these groups would be likely to change with additional sequence additions. At the secondary rank, the only anomaly with respect to the proposed nomenclature was the interjection of the Cry1Ia and Cry1Ib sequences into the Cry1B group. This effect may be due to an artificially reduced distance between the Cry1I sequences and the incomplete Cry1Bd sequence caused by the particular distance metric used. The Cyt lineage sequences were separated into the expected two primary rank groups that separate into the expected secondary rank groupings. This more standard phylogenetic approach also suffers from an accentuated visual disorientation of uneven branch lengths and shortening of the more closely related branches, especially at the tertiary rank (lowercase letter), where a great deal of comparative work has been done among the Cry1 toxins.

In summary, the proposed nomenclature uses readily available software that can be easily interpreted by investigators in the field and meets their needs as well as, or better than, alternative methods of analysis and presentation. When the holotype toxins were analyzed by alternative phylogenetic methods, the hierarchy implied by the nomenclature was essentially consistent with the resulting phylogenetic clustering, and the few exceptions were largely explainable by known properties of the sequences in question.

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REFERENCES

- Adams, L. F., S. Mathewes, P. O'Hara, A. Petersen, and H. Gürtler. 1994. Elucidation of the mechanism of CryIIIA overproduction in a mutagenized strain of *Bacillus thuringiensis* var. *tenebrionis*. *Mol. Microbiol.* **14**:381-389.
- Adams, L. F., J. E. Visick, and H. R. Whiteley. 1989. A 20-kilodalton protein is required for efficient production of the *Bacillus thuringiensis* subsp. *israelensis* 27-kilodalton crystal protein in *Escherichia coli*. *J. Bacteriol.* **171**:521-530.
- Adang, M. J., M. J. Staver, T. A. Rocheleau, J. Leighton, R. F. Barker, and D. V. Thompson. 1985. Characterized full-length and truncated plasmid clones of the crystal protein of *Bacillus thuringiensis* subsp. *kurstaki* HD-73 and their toxicity to *Manduca sexta*. *Gene* **36**:289-300.
- Asano, S. I., Y. Nukumizu, H. Bando, T. Iizuka, and T. Yamamoto. 1997. Cloning of novel enterotoxin genes from *Bacillus cereus* and *Bacillus thuringiensis*. *Appl. Environ. Microbiol.* **63**:1054-1057.
- Barloy, F., A. Delécluse, L. Nicolas, and M.-M. Lecadet. 1996. Cloning and expression of the first anaerobic toxin gene from *Clostridium bifementans* subsp. *malaysia*, encoding a new mosquitocidal protein with homologies to *Bacillus thuringiensis* delta-endotoxins. *J. Bacteriol.* **178**:3099-3105.
- Bishop, A. H. 1994. Unpublished observation.
- Bossé, M., L. Masson, and R. Brousseau. 1990. Nucleotide sequence of a novel crystal protein gene isolated from *Bacillus thuringiensis* subspecies *kenyae*. *Nucleic Acids Res.* **18**:7443.
- Bradley, D., M. A. Harkey, M.-K. Kim, D. Biever, and L. S. Bauer. 1995. The insecticidal CryIB protein of *Bacillus thuringiensis* ssp. *thuringiensis* has dual specificity to coleopteran and lepidopteran larvae. *J. Invertebr. Pathol.* **65**:162-173.
- Bravo, A. 1997. Phylogenetic relationships of *Bacillus thuringiensis* δ -endotoxin family proteins and their functional domains. *J. Bacteriol.* **179**:2793-2801.
- Brizzard, B. L., and H. R. Whiteley. 1988. Nucleotide sequence of an additional crystal protein gene cloned from *Bacillus thuringiensis* subsp. *thuringiensis*. *Nucleic Acids Res.* **16**:2723-2724.
- Brown, K. L., and H. R. Whiteley. 1992. Molecular characterization of two novel crystal protein genes from *Bacillus thuringiensis* subsp. *thompsoni*. *J. Bacteriol.* **174**:549-557.
- Chak, K. F. 1996. Unpublished observation.
- Chak, K. F., and J. C. Chen. 1993. Complete nucleotide sequence and identification of a putative promoter region for the expression in *Escherichia coli* of the *cryIA(b)* gene from *Bacillus thuringiensis* var. *aizawai* HD133. *Proc. Natl. Sci. Coun. Repub. China* **17**:7-14.
- Chambers, J. A., A. Jelen, M. P. Gilbert, C. S. Jany, T. B. Johnson, and C. Gawron-Burke. 1991. Isolation and characterization of a novel insecticidal crystal protein gene from *Bacillus thuringiensis* subsp. *aizawai*. *J. Bacteriol.* **173**:3966-3976.
- Cheong, H., and S. S. Gill. 1997. Cloning and characterization of a cytolytic and mosquitocidal δ -endotoxin from *Bacillus thuringiensis* subsp. *jegathesan*. *Appl. Environ. Microbiol.* **63**:3254-3260.
- Chungjatupornchai, W., H. Höfte, J. Seurinck, C. Angsuthanasombat, and M. Vaeck. 1988. Common features of *Bacillus thuringiensis* toxins specific for *Diptera* and *Lepidoptera*. *Eur. J. Biochem.* **173**:9-16.
- Dankocsik, C., W. P. Donovan, and C. S. Jany. 1990. Activation of a cryptic crystal protein gene of *Bacillus thuringiensis* subspecies *kurstaki* by gene fusion and determination of the crystal protein insecticidal specificity. *Mol. Microbiol.* **4**:2087-2094.
- Dardenne, F., J. Seurinck, B. Lambert, and M. Peferoen. 1990. Nucleotide sequence and deduced amino acid sequence of a *cryIA(c)* gene variant from *Bacillus thuringiensis*. *Nucleic Acids Res.* **18**:5546.
- Delécluse, A., M.-L. Rosso, and A. Ragni. 1995. Cloning and expression of a novel toxin gene from *Bacillus thuringiensis* subsp. *jegathesan* encoding a highly mosquitocidal protein. *Appl. Environ. Microbiol.* **61**:4230-4235.
- Donovan, W. P., C. C. Dankocsik, M. P. Gilbert, W. C. Gawron-Burke, R. R. Groat, and B. C. Carlton. 1988. Amino acid sequence and entomocidal activity of the P2 crystal protein. An insect toxin from *Bacillus thuringiensis* var. *kurstaki*. *J. Biol. Chem.* **263**:561-567. (Author's correction, **263**:4740.)
- Donovan, W. P., C. Dankocsik, and M. P. Gilbert. 1988. Molecular characterization of a gene encoding a 72-kilodalton mosquito-toxic crystal protein from *Bacillus thuringiensis* subsp. *israelensis*. *J. Bacteriol.* **170**:4732-4738.
- Donovan, W. P., J. M. González, Jr., M. P. Gilbert, and C. Dankocsik. 1988. Isolation and characterization of EG2158, a new strain of *Bacillus thuringiensis* toxic to coleopteran larvae, and nucleotide sequence of the toxin gene. *Mol. Gen. Genet.* **214**:365-372.
- Donovan, W. P., M. J. Rugar, and A. C. Slaney. January 1995. U.S. patent 5,378,625.
- Donovan, W. P., M. J. Rugar, A. C. Slaney, T. Malvar, M. C. Gawron-Burke, and T. B. Johnson. 1992. Characterization of two genes encoding *Bacillus thuringiensis* insecticidal crystal proteins toxic to *Coleoptera* species. *Appl. Environ. Microbiol.* **58**:3921-3927.
- Donovan, W. P., Y. Tan, C. S. Jany, and J. M. González, Jr. June 1994. U.S. patent 5,322,687.
- Earp, D. J., and D. J. Ellar. 1987. *Bacillus thuringiensis* var. *morrisoni* strain PG14: nucleotide sequence of a gene encoding a 27 kDa crystal protein. *Nucleic Acids Res.* **15**:3619.
- Felsenstein, J. 1989. PHYLIP—phylogeny inference package (version 2). *Cladistics* **5**:164-166.
- Fischhoff, D. A., K. S. Bowdish, F. J. Perlak, P. G. Marrone, S. H. McCormick, J. G. Niedermeyer, D. A. Dean, K. Kusano-Kretzmer, E. J. Mayer, D. E. Rochester, S. G. Rogers, and R. T. Fraley. 1987. Insect tolerant transgenic tomato plants. *Bio/Technology* **5**:807-813.
- Foncerrada, L., A. J. Sick, and J. M. Payne. August 1992. European Patent Office no. EP 0498537.
- Galjart, N. J., N. Sivasubramanian, and B. A. Federici. 1987. Plasmid location, cloning and sequence analysis of the gene encoding a 23-kilodalton cytolytic protein from *Bacillus thuringiensis* subsp. *morrisoni* (PG-14). *Curr. Microbiol.* **16**:171-177.
- Geiser, M., S. Schweitzer, and C. Grimm. 1986. The hypervariable region in the genes coding for entomopathogenic crystal proteins of *Bacillus thurin-*

- giensis*: nucleotide sequence of the kurhd1 gene of subsp. *kurstaki* HD1. Gene **48**:109–118.
32. Gleave, A. P., R. J. Hedges, and A. H. Broadwell. 1992. Identification of an insecticidal crystal protein from *Bacillus thuringiensis* DSIR517 with significant sequence differences from previously described toxins. J. Gen. Microbiol. **138**:55–62.
 33. Gleave, A. P., R. J. Hedges, A. H. Broadwell, and P. J. Wigley. 1992. Cloning and nucleotide sequence of an insecticidal crystal protein gene from *Bacillus thuringiensis* DSIR732 active against three species of leafroller Lepidoptera *Tortricidae*. N. Z. J. Crop Hortic. Sci. **20**:27–36.
 34. Gleave, A. P., R. Williams, and R. J. Hedges. 1993. Screening by polymerase chain reaction of *Bacillus thuringiensis* serotypes for the presence of *cryV*-like insecticidal protein genes and characterization of a *cryV* gene cloned from *B. thuringiensis* subsp. *kurstaki*. Appl. Environ. Microbiol. **59**:1683–1687.
 35. Guerchicoff, A., R. U. Ugalde, and C. P. Rubinstein. 1997. Identification and characterization of a previously undescribed *cyt* gene in *Bacillus thuringiensis* subsp. *israelensis*. Appl. Environ. Microbiol. **63**:2716–2721.
 36. Haider, M. Z., and D. J. Ellar. 1988. Nucleotide sequence of a *Bacillus thuringiensis* aizawai ICI entomocidal crystal protein gene. Nucleic Acids Res. **16**:10927.
 37. Hefford, M. A., R. Brousseau, G. Préfontaine, Z. Hanna, J. A. Condie, and P. C. K. Lau. 1987. Sequence of a lepidopteran toxin gene of *Bacillus thuringiensis* subsp. *kurstaki* NRD-12. J. Biotechnol. **6**:307–322.
 38. Herrera, G., S. J. Snyman, and J. A. Thomson. 1994. Construction of a bioinsecticidal strain of *Pseudomonas fluorescens* active against the sugarcane borer, *Eldana saccharina*. Appl. Environ. Microbiol. **60**:682–690.
 39. Herrnstadt, C., T. E. Gilroy, D. A. Sobieski, B. D. Bennett, and F. H. Gaertner. 1987. Nucleotide sequence and deduced amino acid sequence of a coleopteran-active delta-endotoxin gene from *Bacillus thuringiensis* subsp. *san diego*. Gene **57**:37–46.
 40. Höfte, H., H. de Greve, J. Seurinck, S. Jansens, J. Mahillon, C. Ampe, J. Vandekerckhove, M. van Montagu, M. Zabeau, and M. Vaeck. 1986. Structural and functional analysis of a cloned delta endotoxin of *Bacillus thuringiensis berliner* 1715. Eur. J. Biochem. **161**:273–280.
 41. Höfte, H., J. Seurinck, A. Van Houtven, and M. Vaeck. 1987. Nucleotide sequence of a gene encoding an insecticidal protein of *Bacillus thuringiensis* var. *tenebrionis* toxic against Coleoptera. Nucleic Acids Res. **15**:7183.
 42. Höfte, H., P. Soetaert, S. Jansens, and M. Peferoen. 1990. Nucleotide sequence and deduced amino acid sequence of a new Lepidoptera-specific crystal protein gene from *Bacillus thuringiensis*. Nucleic Acids Res. **18**:5545.
 43. Höfte, H., J. Van Rie, S. Jansens, A. Van Houtven, H. Vanderbruggen, and M. Vaeck. 1988. Monoclonal antibody analysis and insecticidal spectrum of three types of lepidopteran-specific insecticidal crystal proteins of *Bacillus thuringiensis*. Appl. Environ. Microbiol. **54**:2010–2017.
 44. Höfte, H., and H. R. Whiteley. 1989. Insecticidal crystal proteins of *Bacillus thuringiensis*. Microbiol. Rev. **53**:242–255.
 45. Honée, G., T. van der Salm, and B. Visser. 1988. Nucleotide sequence of crystal protein gene isolated from *B. thuringiensis* subspecies *entomocidus* 60.5 coding for a toxin highly active against *Spodoptera* species. Nucleic Acids Res. **16**:6240.
 46. Hori, H., K. Suzuki, K. Ogiwara, M. Minami, M. Himejima, K. Sakanaka, Y. Kaji, S. Asano, R. Sato, M. Ohba, and H. Iwahana. 1992. Presented at the XXVth Annual Meeting of the Society for Invertebrate Pathology, Heidelberg, Germany.
 47. Ibarra, J. 1997. Unpublished observation.
 48. Kalman, S., K. L. Kiehne, J. L. Libs, and T. Yamamoto. 1993. Cloning of a novel *cryIC*-type gene from a strain of *Bacillus thuringiensis* subsp. *galleriae*. Appl. Environ. Microbiol. **59**:1131–1137.
 49. Kang, S. K., H. S. Kim, and Y. M. Yu. 1997. Unpublished observation.
 50. Kondo, S., N. Tamura, A. Kunitate, M. Hattori, A. Akashi, and I. Ohmori. 1987. Cloning and nucleotide sequencing of two insecticidal δ -endotoxin genes from *Bacillus thuringiensis* var. *kurstaki* HD-1 DNA. Agric. Biol. Chem. **51**:455–463.
 51. Koni, P. A., and D. J. Ellar. 1993. Cloning and characterization of a novel *Bacillus thuringiensis* cytolytic delta-endotoxin. J. Mol. Biol. **229**:319–327.
 52. Koo, B. T. 1995. Unpublished observation.
 53. Koo, B. T., S. H. Park, S. K. Choi, B. S. Shin, J. I. Kim, and J. H. Yu. 1995. Cloning of a novel crystal protein gene *cryIK* from *Bacillus thuringiensis* subsp. *morisoni*. FEMS Microbiol. Lett. **134**:159–164.
 54. Kostichka, K., G. W. Warren, M. Mullins, A. D. Mullins, J. A. Craig, M. G. Koziel, and J. J. Estruch. 1996. Cloning of a *cryV*-type insecticidal protein gene from *Bacillus thuringiensis*: the *cryV*-encoded protein is expressed early in stationary phase. J. Bacteriol. **178**:2141–2144.
 55. Kronstad, J. W., and H. R. Whiteley. 1986. Three classes of homologous *Bacillus thuringiensis* crystal-protein genes. Gene **43**:29–40.
 56. Lambert, B. 1993. Unpublished observation.
 57. Lambert, B., L. Buysse, C. Decock, S. Jansens, C. Piens, B. Saey, J. Seurinck, K. Van Audenhove, J. Van Rie, A. Van Vliet, and M. Peferoen. 1996. A *Bacillus thuringiensis* insecticidal protein with a high activity against members of the family Noctuidae. Appl. Environ. Microbiol. **62**:80–86.
 58. Lambert, B., H. Höfte, K. Anny, S. Jansens, P. Soetaert, and M. Peferoen. 1992. Novel *Bacillus thuringiensis* insecticidal crystal protein with a silent activity against coleopteran larvae. Appl. Environ. Microbiol. **58**:2536–2542.
 59. Lambert, B., W. Theunis, R. Agouda, K. Van Audenhove, D. C., S. Jansens, J. Seurinck, and M. Peferoen. 1992. Nucleotide sequence of gene *cryIIID* encoding a novel coleopteran-active crystal protein from strain BT1109P of *Bacillus thuringiensis* subsp. *kurstaki*. Gene **110**:131–132.
 60. Lee, C.-S., and A. I. Aronson. 1991. Cloning and analysis of δ -endotoxin genes from *Bacillus thuringiensis* subsp. *alesti*. J. Bacteriol. **173**:6635–6638.
 61. Lee, H.-K., and S. S. Gill. 1997. Molecular cloning and characterization of a novel mosquitocidal protein gene from *Bacillus thuringiensis* subsp. *fukuokaensis*. Appl. Environ. Microbiol. **63**:4664–4670.
 62. Masson, L., P. Marcotte, G. Préfontaine, and R. Brousseau. 1989. Nucleotide sequence of a gene cloned from *Bacillus thuringiensis* subspecies *entomocidus* coding for an insecticidal protein toxic for *Bombyx mori*. Nucleic Acids Res. **17**:446.
 63. Masson, L., A. Mazza, L. Gringorten, D. Baines, V. Aneliunas, and R. Brousseau. 1994. Specificity domain localization of *Bacillus thuringiensis* insecticidal toxins is highly dependent on the bioassay system. Mol. Microbiol. **14**:851–860.
 64. McLinden, J. H., J. R. Sabourin, B. D. Clark, D. R. Genster, W. E. Workman, and D. H. Dean. 1985. Cloning and expression of an insecticidal k-73 type crystal protein gene from *Bacillus thuringiensis* var. *kurstaki* into *Escherichia coli*. Appl. Environ. Microbiol. **50**:623–628.
 65. McPherson, S. A., F. J. Perliak, R. L. Fuchs, P. G. Marrone, P. B. Lavrik, and D. A. Fischhoff. 1988. Characterization of the coleopteran-specific protein gene of *Bacillus thuringiensis* var. *tenebrionis*. Bio/Technology **6**:61–66.
 66. Michaels, T. E., K. E. Narva, and L. Foncerrada. August 1993. World Intellectual Property Organization patent WO 93/15206.
 67. Narva, K. E., J. M. Payne, G. E. Schwab, L. A. Hickle, T. Galasan, and A. J. Sick. December 1991. European Patent Office no. EP 0462721.
 68. Narva, K. E., G. E. Schwab, T. Galasan, and J. M. Payne. August 1993. U.S. patent 5,236,843.
 - 68a. Nelson, D. R., L. Koymans, T. Kamataki, J. J. Stegeman, R. Feyerisen, D. J. Waxman, M. R. Waterman, O. Gotoh, M. J. Coon, R. W. Estabrook, I. C. Gunsalus, and D. W. Nebert. 1996. P450 superfamily: update on new sequences, gene mapping, accession numbers and nomenclature. Pharmacogenetics **6**:1–42.
 69. Oeda, K., K. Oshie, M. Shimizu, K. Nakamura, H. Yamamoto, I. Nakayama, and H. Ohkawa. 1987. Nucleotide sequence of the insecticidal protein gene of *Bacillus thuringiensis* strain *aizawai* IPL7 and its high-level expression in *Escherichia coli*. Gene **53**:113–119.
 70. Ogiwara, K., H. Hori, M. Minami, K. Takeuchi, R. Sato, M. Ohba, and H. Iwahana. 1995. Nucleotide sequence of the gene encoding novel delta-endotoxin from *Bacillus thuringiensis* serovar *japonensis* strain Buibui specific to scarabaeid beetles. Curr. Microbiol. **30**:227–235.
 71. Omolo, E. O., J. M. D., O. E. O., and J. A. Thomson. 1997. Cloning and expression of a *Bacillus thuringiensis* (L1-2) gene encoding a crystal protein active against *Glossina morsitans morsitans* and *Chilo partellus*. Curr. Microbiol. **34**:118–121.
 72. Orduz, S. Unpublished observation.
 73. Page, R. D. M. 1996. TREEVIEW: an application to display phylogenetic trees on personal computers. CABIOS **12**:357–358.
 74. Payne, J. M., K. E. Narva, and J. Fu. December 1996. U.S. patent 5,589,382.
 75. Payne, J. M., and J. M. Fu. February 1994. U.S. patent 5,286,486.
 76. Payne, J. M., M. K. Kennedy, J. B. Randall, H. Meier, H. J. Uick, L. Foncerrada, H. E. Schnepf, G. E. Schwab, and J. Fu. January 1997. U.S. patent 5,596,071.
 77. Payne, J. M., and K. E. Narva. July 1994. World Intellectual Property Organization patent WO 94/16079.
 78. Payne, J. M., K. E. Narva, K. A. Uyeda, C. J. Stalder, and T. E. Michaels. July 1995. U.S. patent 5,436,002.
 79. Payne, J. M., and A. J. Sick. September 1993. U.S. patent 5,246,852.
 80. Payne, J. M., and A. J. Sick. February 1993. U.S. patent 5,188,960.
 81. Payne, J. M., and A. J. Sick. April 1993. U.S. patent 5,206,166.
 82. Payne, J. M., and A. J. Sick. August 1991. U.S. patent 5,039,523.
 83. Payne, J. M., A. J. Sick, and M. Thompson. August 1992. U.S. patent 5,135,867.
 84. Payne, J. M., G. G. Soares, H. W. Talbot, and T. C. Olson. October 1991. U.S. patent 4,990,332.
 85. Peferoen, M., B. Lambert, and H. Joos. August 1990. European patent Office no. EP 0382990-A1.
 86. Rosso, M. L., and A. Delecluse. 1997. Contribution of the 65-kilodalton protein encoded by the cloned gene *cry19A* to the mosquitocidal activity of *Bacillus thuringiensis* subsp. *jegathesan*. Appl. Environ. Microbiol. **63**:4449–4455.
 87. Saitoh, H. 1996. Unpublished observation.
 88. Sanchis, V., D. Lereclus, G. Menou, J. Chaufaux, S. Guo, and M.-M. Lecadet. 1989. Nucleotide sequence and analysis of the N-terminal coding region of the *Spodoptera*-active δ -endotoxin gene of *Bacillus thuringiensis* *aizawai* 7.29. Mol. Microbiol. **3**:229–238.

89. Sasaki, J., S. Asano, N. Hashimoto, B.-W. Lay, S. Hastowo, H. Bando, and T. Iizuka. 1997. Characterization of a *cry2A* gene cloned from an isolate of *Bacillus thuringiensis* serovar *sotto*. *Curr. Microbiol.* **35**:1–8.
90. Schnepf, H. E., G. E. Schwab, J. M. Payne, K. E. Narva, and L. Foncerrada. November 1992. World Intellectual Property Organization patent WO 92/19739.
91. Schnepf, H. E., and H. R. Whiteley. 1981. Cloning and expression of the *Bacillus thuringiensis* crystal protein gene in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* **78**:2893–2897.
92. Schnepf, H. E., H. C. Wong, and H. R. Whiteley. 1985. The amino acid sequence of a crystal protein from *Bacillus thuringiensis* deduced from the DNA base sequence. *J. Biol. Chem.* **260**:6264–6272.
93. Sekar, V., D. V. Thompson, M. J. Maroney, R. G. Bookland, and M. J. Adang. 1987. Molecular cloning and characterization of the insecticidal crystal protein gene of *Bacillus thuringiensis* var. *tenebrionis*. *Proc. Natl. Acad. Sci. USA* **84**:7036–7040.
94. Selvapandiyar, A. 1996. Unpublished observation.
95. Sen, K., G. Honda, N. Koyama, M. Nishida, A. Neki, H. Sakai, M. Himeno, and T. Komano. 1988. Cloning and nucleotide sequences of the two 130 kDa insecticidal protein genes of *Bacillus thuringiensis* var. *israelensis*. *Agric. Biol. Chem.* **52**:873–878.
96. Shevelev, A. B., Y. N. Kogan, A. M. Busheva, E. J. Voronina, D. V. Tebrikov, S. I. Novikova, G. G. Chestukhina, V. Kubshinov, E. Pehu, and V. M. Stepanov. 1997. A novel delta-endotoxin gene *cryIM* from *Bacillus thuringiensis* subsp. *wuhanensis*. *FEBS Lett.* **404**:148–152.
97. Shevelev, A. B., M. A. Svarinsky, A. I. Karasin, Y. N. Kogan, G. G. Chestukhina, and V. M. Stepanov. 1993. Primary structure of *cryX*, the novel delta-endotoxin-related gene from *Bacillus thuringiensis* spp. *galleriae*. *FEBS Lett.* **336**:79–82.
98. Shibano, Y., A. Yamagata, N. Nakamura, T. Iizuka, H. Sugisaki, and M. Takanami. 1985. Nucleotide sequence coding for the insecticidal fragment of the *Bacillus thuringiensis* crystal protein. *Gene* **34**:243–251.
99. Shimizu, M., K. Oshie, K. Nakamura, Y. Takada, K. Oeda, and H. Ohkawa. 1988. Cloning and expression in *Escherichia coli* of the 135-kDa insecticidal protein gene from *Bacillus thuringiensis* subsp. *aizawai* IPL7. *Agric. Biol. Chem.* **52**:1565–1573.
100. Shin, B.-S., S.-H. Park, S.-K. Choi, B.-T. Koo, S.-T. Lee, and J.-I. Kim. 1995. Distribution of *cryV*-type insecticidal protein genes in *Bacillus thuringiensis* and cloning of *cryV*-type genes from *Bacillus thuringiensis* subsp. *kurstaki* and *Bacillus thuringiensis* subsp. *entomocidus*. *Appl. Environ. Microbiol.* **61**:2402–2407.
101. Sick, A., F. Gaertner, and A. Wong. 1990. Nucleotide sequence of a coleopteran-active toxin gene from a new isolate of *Bacillus thuringiensis* subsp. *tolworthi*. *Nucleic Acids Res.* **18**:1305.
102. Sick, A. J., G. E. Schwab, and J. M. Payne. January 1994. U.S. patent 05281530.
103. Smith, G. P., and D. J. Ellar. 1994. Mutagenesis of two surface-exposed loops of the *Bacillus thuringiensis* CryIC δ -endotoxin affects insecticidal specificity. *Biochem. J.* **302**:611–616.
104. Smulevitch, S. V., A. L. Osterman, A. B. Shevelev, S. V. Kaluger, A. I. Karasin, R. M. Kadyrov, O. P. Zagnitko, G. G. Chestukhina, and V. M. Stepanov. 1991. Nucleotide sequence of a novel delta-endotoxin gene *cryIG* of *Bacillus thuringiensis* ssp. *galleriae*. *FEBS Lett.* **293**:25–28.
105. Soetaert, P. 1996. Unpublished observation.
106. Strizhov, N. 1996. Unpublished observation.
107. Sun, M. 1997. Unpublished observation.
108. Tailor, R., J. Tippett, G. Gibb, S. Pells, D. Pike, L. Jordon, and S. Ely. 1992. Identification and characterization of a novel *Bacillus thuringiensis* delta-endotoxin entomocidal to coleopteran and lepidopteran larvae. *Mol. Microbiol.* **6**:1211–1217.
109. Thiery, I., A. Delécluse, M. C. Tamayo, and S. Orduz. 1997. Identification of a gene for Cyt1A-like hemolysin from *Bacillus thuringiensis* subsp. *medellin* and expression in a crystal-negative *B. thuringiensis* strain. *Appl. Environ. Microbiol.* **63**:468–473.
110. Thompson, J. D., D. G. Higgins, and T. J. Gibson. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **22**:4673–4680.
111. Thorne, L., F. Garduno, T. Thompson, D. Decker, M. Zounes, M. Wild, A. M. Walfield, and T. J. Pollock. 1986. Structural similarity between the Lepidoptera- and Diptera-specific insecticidal endotoxin genes of *Bacillus thuringiensis* subsp. "*kurstaki*" and "*israelensis*." *J. Bacteriol.* **166**:801–811.
112. Tungradubkul, S., C. Settasatien, and S. Panyim. 1988. The complete nucleotide sequence of a 130 kDa mosquito-larvicidal delta-endotoxin gene of *Bacillus thuringiensis* var. *israelensis*. *Nucleic Acids Res.* **16**:1637–1638.
113. Udayasuriyan, V., A. Nakamura, H. Mori, H. Masaki, and T. Uozumi. 1994. Cloning of a new *cryIA(a)* gene from *Bacillus thuringiensis* strain FU-2-7 and analysis of chimeric *cryIA(a)* proteins for toxicity. *Biosci. Biotechnol. Biochem.* **58**:830–835.
114. Van Mellaert, H., J. Botterman, J. Van Rie, and H. Joos. January 1991. European Patent EP Office no. 0408403.
115. Visser, B., E. Munsterman, A. Stoker, and W. G. Dirkse. 1990. A novel *Bacillus thuringiensis* gene encoding a *Spodoptera exigua*-specific crystal protein. *J. Bacteriol.* **172**:6783–6788.
116. Von Tersch, M. A., and J. M. Gonzalez. October 1994. U.S. patent 5,356,623.
117. Von Tersch, M. A., H. L. Robbins, C. S. Jany, and T. B. Johnson. 1991. Insecticidal toxins from *Bacillus thuringiensis* subsp. *kenyae*: gene cloning and characterization and comparison with *B. thuringiensis* subsp. *kurstaki* CryIA(c) toxins. *Appl. Environ. Microbiol.* **57**:349–358.
118. Waalwijk, C., A. M. Dullemans, M. E. S. vanWorkum, and B. Visser. 1985. Molecular cloning and the nucleotide sequence of the Mr28,000 crystal protein gene of *Bacillus thuringiensis* subsp. *israelensis*. *Nucleic Acids Res.* **13**:8207–8217.
119. Wabiko, H., K. C. Raymond, and L. A. Bulla, Jr. 1986. *Bacillus thuringiensis* entomocidal protoxin gene sequence and gene product analysis. *DNA* **5**:305–314.
120. Ward, E. S., and D. J. Ellar. 1986. *Bacillus thuringiensis* var. *israelensis* delta-endotoxin: nucleotide sequence and characterization of the transcripts in *Bacillus thuringiensis* and *Escherichia coli*. *J. Mol. Biol.* **191**:1–11.
121. Ward, E. S., and D. J. Ellar. 1987. Nucleotide sequence of a *Bacillus thuringiensis* var. *israelensis* gene encoding a 130 kDa delta-endotoxin. *Nucleic Acids Res.* **15**:7195.
122. Wasano, N., and M. Ohba. 1998. Unpublished observation.
- 122a. White, J. A., L. J. Maltais, and D. W. Nebert. 1998. An increasingly urgent need for standardized gene nomenclature. See: http://genetics.nature.com/web_specials/nomen/nomen_article.html.
123. Widner, W. R., and H. R. Whiteley. 1989. Two highly related insecticidal crystal proteins of *Bacillus thuringiensis* subsp. *kurstaki* possess different host range specificities. *J. Bacteriol.* **171**:965–974.
124. Wu, D., X. L. Cao, Y. Y. Bai, and A. I. Aronson. 1991. Sequence of an operon containing a novel δ -endotoxin gene from *Bacillus thuringiensis*. *FEMS Microbiol. Lett.* **81**:31–36.
125. Yamamoto, T., I. A. Watkinson, L. Kim, M. V. Sage, R. Stratton, N. Akande, Y. Li, D.-P. Ma, and B. A. Roe. 1988. Nucleotide sequence of the gene coding for a 130-kDa mosquitoicidal protein of *Bacillus thuringiensis israelensis*. *Gene* **66**:107–120.
126. Zhang, J., T. C. Hodgman, L. Krieger, W. Schnetter, and H. U. Schairer. 1997. Cloning and analysis of the first *cry* gene from *Bacillus popilliae*. *J. Bacteriol.* **179**:4336–4341.

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