



EUROPEAN COMMISSION
HEALTH AND CONSUMERS DIRECTORATE-GENERAL

Director General

SANCO/10256/2013

Programmes for the eradication, control and monitoring of certain animal diseases and zoonoses

Survey programme for Avian Influenza

Lithuania

Approved* for 2013 by Commission Decision 2012/761/EU

* in accordance with Council Decision 2009/470/EC

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

1. Identification of the programme

Member state : LIETUVA

Disease : avian influenza in poultry and wild birds

Request of Community co-financing from beginning of: to end of

1.1 Contact

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2. Description and implementation of the surveillance programme in poultry

2.1.1 Designation of the central authority in charge of supervising and coordinating the departments responsible for implementing the programme

(max. 32000 chars) :

State Food and Veterinary Service (hereinafter – SFVS) is the sole competent authority in Lithuania responsible for the official control of implementation of the legislation on food, feed, animal health and welfare. State Food and Veterinary Service (central service) consists of Administration and 15 Departments. Administration is comprised of the Director and 4 his deputies. The director is also Chief Veterinary Inspector of the State. Four Departments are concerned with animal health. (Emergency Response Department, Food Department, Veterinary Sanitary Department, Animal Health and Welfare Department)

Functions of Animal Health and Welfare Department, responsible for implementation of programme, are as follow:

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

The Animal Health and Welfare Department is responsible for the coordination and control of all territorial State Food and Veterinary Services involved in the implementation of this program. This department collects the data, performs statistical analysis and evaluation of the surveillance program and informs the relevant authorities in European Union about the progress of the control and surveillance program.

Also:

- Analyses epizootic situation of contagious animal diseases in Lithuania and other countries, makes assessment of risk factors and adopts decisions on control of infectious animal diseases and contingency actions.
- Organises monitoring and control of contagious animal diseases and zoonoses, and eradication of outbreaks of infectious animal diseases.
- Carries out control over identification and registration of animals, over trade of animals, animal by-products, feeds and feed additives, over import, distribution and use of veterinary medicines.
- Analyses, implements EU legislation or drafts national legislation on animal welfare, control, monitoring and eradication of infectious animal diseases.

51 territorial State Food and Veterinary Services (administrative divisions) perform official food and veterinary control.

2.1.2 System in place for the registration of holdings

(max. 32000 chars):

The commercial poultry farms are registered in accordance with Order No. B1-517 of the Director of State Food and Veterinary Service of 3 July 2012 on the approval of legal subject that are objects of veterinary supervision by the local state food and veterinary services. The local SFVS provide the updates to the list of approved legal subjects to the Animal Health and Welfare Department monthly by the local state food and veterinary services and included into national register available on Internet <http://www.vet.lt/objektai/LTindex.html>. The registered poultry farms are under veterinary supervision.

The following programs must be presented and operational for the veterinary approval of the holding:

1. The description of technological process and schemes of the holding and the territory.
2. Self – control program.
3. Control Program of drinking water.
4. Program of collection and disposal of waste water
5. Program of use and disposal of animal by products.
6. Control Program of pests.
7. Program of cleaning and disinfection.
8. Program of personal hygiene and health control
9. Program of training of personnel.

The means of transport must be cleansed and disinfected using method and means, approved by SFVS

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

before and after the transportation of live birds.

Small poultry keepers are registered in to Farm animal register which is central Data Base for all farm animals including poutries.

2.1.3 *Design (risk based or surveillance based on representative sampling)*

(max. 32000 chars) :

1. The location of the poultry holding in proximity to wet areas, ponds, swamps, lakes, rivers or sea shores where migratory wild water birds may gather.
2. The location of the poultry holding in areas with a high density of migratory wild birds, in particular of those birds that are characterised as 'target species' (TS) for HPAI H5N1 detection.
3. The location of poultry holding in proximity to resting and breeding places of migratory wild water birds, in particular where these areas are linked through migratory birds' movements to areas where HPAI H5N1 is known to occur in wild birds or poultry.
4. Poultry holdings with free range production, or poultry holdings where poultry or other captive birds are kept in the open-air in any premises in which contact with wild birds cannot be sufficiently prevented.
5. Low biosecurity level in the poultry holding, including the method of storage of feed and the use of surface water.

2.1.3.1 *Short description of predominant poultry population and types of poultry production*

(max. 32000 chars) :

The predominant poultry population in Lithuania in 2012 are as follows:

Laying hens of Gallus gallus: 26 approved holdings with 127 flocks (approx. 2 844 500 birds);

Broilers of Gallus gallus: 41 approved holdings with 272 flocks (approx. 7 250 000 birds);

Breeding poultry of Gallus gallus: 13 approved holdings with 65 flocks (approx. 362100 birds);

Ducks: 1 approved holding with 1 flock (approx. 15 000 birds);

Fattening turkeys: 19 approved holdings with 46 flocks (approx. 451 700 birds).

2.1.3.2 *Criteria and risk factors for risk based surveillance(1)*

(max. 32000 chars) :

1. A risk-based surveillance will be implemented as a "passive" surveillance system by laboratory

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

- investigation of moribund wild birds or birds found dead and it will be specifically directed towards water bird species.
2. Wild birds, in particular migratory water birds, will be specifically targeted and the samples will be taken by hunters and official vets.
 3. Areas close to the sea, lakes and waterways where birds were found dead will be targeted;
 4. Detecting subclinical infections in domestic poultry population with AI of subtypes H5 and H7 thereby complementing early detection systems.
 5. Detecting infections of AI H5 and H7 subtypes in specifically targeted poultry populations at specific risk for infection due to their husbandry system or species specific susceptibility.
 6. Continuing a "baseline" surveillance of different species of free living migratory birds as part of continuous monitoring of LPAI viruses. Anseriformes (water fowl) and Charadriiformes (shorebirds and gulls) shall be the main sampling targets to assess if they carry LPAI viruses of H5 and H7 subtypes (which would in any case also detect HPAI H5N1 and other HPAI, if present). "Higher risk species" will be targeted in particular.

- (1) Including maps showing target sampling sites identified as being particularly at risk for the introduction of avian influenza virus, taking into account criteria set out in point 4 of Annex I to Commission Decision 2010/367/EC.

2.2 Target populations (2)

(max. 32000 chars):

1. Investigation of living, sick or dead wild birds and increased mortalities, in particular in selected higher risk species and in areas epidemiologically linked to cases of HPAI H5N1 to identify asymptomatic carriers and investigation of the risk that these species in close contact with domestic poultry holdings might function as bridge species.
2. Investigation domestic poultry populations at specific risk for infection due to their husbandry system or specific susceptibility species and surveys of different species of free living migratory birds. Waterfowl and shorebirds are the main sampling targets to assess if they carry LPAI viruses of H5 and H7 subtypes.

- (2) including MS specific exceptional circumstances as described in Annex I point 3 of Commission Decision 2010/367/EU

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

2.2.1 POULTRY HOLDINGS (a) (except ducks, geese and farmed game birds (waterfowl e.g. mallards) to be sampled

Serological investigation according to Annex I to Commission Decision 2010/367/EU

Category : Laying hens

delete this category

| NUTS (2) (b) | Total number of holdings(c) | Total number of holdings to be sampled | Number of samples per holding | Total number of tests | Method of laboratory analysis | X |
|----------------------|-----------------------------|--|-------------------------------|-----------------------|-------------------------------|---|
| Lithuania | 26 | 26 | 10 | 260 | ELISA test | |
| Total | 26 | 26 | 10 | 260 | | |
| Add a new row | | | | | | |

(a) Holdings or herds or flocks or establishments as appropriate.

(b) Refers to the location of the holding of origin. In case NUTS (Nomenclature of Territorial Units for Statistics) can not be used, region as defined in the programme by the Member States is requested

(c) Total number of holdings of one category of poultry in concerned NUTS 2 region.

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

Category : chicken breeders

delete this category

| NUTS (2) (b) | Total number of holdings(c) | Total number of holdings to be sampled | Number of samples per holding | Total number of tests | Method of laboratory analysis |
|----------------------|-----------------------------|--|-------------------------------|-----------------------|-------------------------------|
| Lithuania | 13 | | 13 | 10 | 130 ELISA test |
| Total | 13 | | 13 | 10 | 130 |
| Add a new row | | | | | |

(a) Holdings or herds or flocks or establishments as appropriate.

(b) Refers to the location of the holding of origin. In case NUTS (Nomenclature of Territorial Units for Statistics) can not be used, region as defined in the programme by the Member States is requested

(c) Total number of holdings of one category of poultry in concerned NUTS 2 region.

Category : fattening turkeys

delete this category

| NUTS (2) (b) | Total number of holdings(c) | Total number of holdings to be sampled | Number of samples per holding | Total number of tests | Method of laboratory analysis |
|----------------------|-----------------------------|--|-------------------------------|-----------------------|-------------------------------|
| Lithuania | 19 | | 19 | 10 | 190 ELISA test |
| Total | 19 | | 19 | 10 | 190 |
| Add a new row | | | | | |

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

| Add a new row | | | |
|--|----|----|-----|
| <p>(a) Holdings or herds or flocks or establishments as appropriate.</p> <p>(b) Refers to the location of the holding of origin. In case NUTS (Nomenclature of Territorial Units for Statistics) can not be used, region as defined in the programme by the Member States is requested.</p> <p>(c) Total number of holdings of one category of poultry in concerned NUTS 2 region.</p> | | | |
| Add a category | | | |
| | | | |
| | | | |
| Total Poultry | 58 | 30 | 580 |

2.2.2 DUCKS, GEESE AND FARMED GAME BIRDS (WATERFOWL e.g. MALLARD) HOLDINGS (a) to be sampled.

Serological investigation according to Annex I to Commission Decision 2010/367/EU

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

Category : Ducks

delete this category

| NUTS (2) (b) | Total number of duck and geese holdings | Total number of duck and geese; holdings to be sampled | Number of samples per holding | Total number of tests | Method of laboratory analysis |
|----------------------|---|--|-------------------------------|-----------------------|-------------------------------|
| 1Lithuania | 1 | 1 | 1 | 10 | 10 ELISA test |
| Total | 1 | 1 | 1 | 10 | 10 |
| Add a new row | | | | | |

(a) Holdings or herds or flocks or establishments as appropriate.

(b) Refers to the location of the holding of origin. In case NUTS (2) code can not be used, region as defined in the programme by the Member State is requested

Add a category

| | | | | |
|--------------|---|---|----|----|
| | | | | |
| | | | | |
| Total | 1 | 1 | 10 | 10 |

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

| | | | |
|-------------------------------|----|----|-----|
| | | | |
| | | | |
| Grand Total | 59 | 59 | 40 |
| Poultry + Ducks/Gueese | | | 590 |

2.3 Sampling procedures, sampling periods and frequency of testing

(max. 32000 chars):

Sampling will be carried out in accordance with the approved surveillance programme from 1 January to 31 December of the year of implementation of that programme.

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

2.4. Laboratory testing : description of the laboratory tests used and follow up investigations

Description of the used serological tests : (max 32000 chars)

Serological testing

The State Food and Veterinary Service has had a serological monitoring programme for avian influenza in place since 1999. The programme is part of the Animal Health Programme, and monitors commercial breeding poultry (chickens, turkeys, ducks and geese) just before they come into lay, and when they move between sites. In addition blood samples from poultry are screened, as are commercial laying flocks prior to export. The serological test will be the HI test.

Laboratory performs detection of avian influenza (HPA) disease antibodies against in serum by Enzyme-linked immunosorbent assay (ELISA) and haemagglutination inhibition (HI) method, according to OIE recommendations (as described in "Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2012" Chapter 10.4).

Detection of antibodies against highly pathogenic avian influenza (HPA) A viruses in serum by ELISA method:

The IDEXX AI Ab Test is an enzyme-linked immunosorbent assay (ELISA) designed to detect antibody to avian influenza viruses (AI) in chicken serum. The IDEXX AI Ab Test detects antibody reactivity to 20 different subtypes, including 14 hemagglutinin glycoproteins and the H5N1 subtype. (http://www.idexx.com/view/xhtml/en_us/livestock-poultry/avian-influenza.jsf)

Detection of antibodies against highly pathogenic avian influenza (HPA) A viruses in serum by haemagglutination inhibition (HI) method. (http://vla.defra.gov.uk/services/ser_tests.htm)

Detection of Influenza virus A subtypes H5 and H7 using one-step RT-PCR:

Laboratory performs detection of avian influenza (HPA), according to OIE recommendations (as described in "Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2012" Chapter 10.4).

All avian influenza virus isolates of cases in wild birds shall be submitted to the CRL in accordance with Community legislation, unless derogation according to paragraph 4 of Chapter V under Differential diagnosis in the avian influenza Diagnostic Manual laid down in Commission Decision 2006/437/ EC is granted. Viruses of H5/H7 subtype shall be submitted without delay and shall be subjected to the standard characterisation tests (nucleotide

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

sequencing/IVP) according to the said avian influenza Diagnostic Manual.

All samples collected in the surveillance for avian influenza in wild birds shall be tested as soon as possible by molecular techniques if available and according to the diagnostic manual (Commission Decision 2006/437/EC). These tests shall only be carried out in laboratories able to guarantee quality assurance and using methods recognized by the CRL for avian influenza. In addition, methods used must be produced acceptable results in the most recent comparative ring test of national laboratories. Initial screening using M gene PCR is recommended, with rapid testing of positives for H5 (but within 2 weeks) and in case of a positive finding analysis of the cleavage site must be undertaken as soon as possible to determine whether or not it has a highly pathogenic avian influenza (HPAI) or a low pathogenic avian influenza (LPAI) motif. If H5 HPAI is confirmed further analysis to determine the N type must be done rapidly (even this can only provide evidence eliminating N1). (http://www.oie.int/fileadmin/Home/eng/Health_standards/tahc/2010/en_chapitre_1.10.4.htm)

Virus isolation testing:

Virus isolation testing is carried out on all suspicious cases. All viruses isolated will be sent to the CRL, where H5 and H7 subtypes will be subjected to characterisation (IVP) and nucleotide sequencing). The CRL will provide the protocol for sending isolates to the CRL, and the reporting tables for collection of survey data.

3. Description and implementation of the surveillance programme in wild birds

- 3.1.1 Designation of the central authority in charge of supervising and coordinating the departments responsible for implementing the programme and relevant collaborating partners (e.g. epidemiologists, ornithologists, nature bird observation and hunter organisations).*

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

(max. 32000 chars):

State Food and Veterinary Service (hereafter - SFVS) with all territorial units (in total 51) is a central authority in charge of supervising and coordinating of sampling procedure. The Animal Health and Welfare Department is responsible for the coordination and control of all territorial State Food and Veterinary Services involved in the implementation of this program. This department collects the data, performs statistical analysis and evaluation of the surveillance program and informs the relevant authorities in European Union about the progress of the control and surveillance program. The territorial SFVS have the contracts with private vets (authorized vets), which are involved in sampling. In each territorial SFVS are contracts with hunting clubs and with regional environment protection departments. Wildlife inspectors from the National Parks and Wildlife Service will participate in the sampling programme by reporting abnormal death of the bird. Ornithologists are also involved. In case they found dead wild birds they must to report to regional territorial SFVS and authorized vets or governmental veterinary inspectors take a samples and deliver to the National Food and Veterinary Risk Assessment Institute for AI testing.

3.1.2 Description and delimitation of the geographical and administrative areas in which the programme is to be applied

max. 32000 chars):

Samples will be taken in whole territory of Lithuania in each region (in total 44 regions) by 51 territorial SFVS. In total 612 samples will be taken from wild poultry (12 samples from each territorial SFVS).

3.1.3 Estimation of the local and/or migratory wildlife population

(max. 32000 chars):

NA

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

3.2 Design, criteria, risk factors and target population⁽³⁾

(max. 32000 chars):

1. Wet areas, ponds, swamps, lakes, rivers or sea shores where migratory wild water birds may gather.
2. Areas with a high density of migratory wild birds, in particular of those birds that are characterised as 'target species' (TS) for HPAI H5N1 detection.
3. The location of resting and breeding places of migratory wild water birds, in particular where these areas are linked through migratory birds' movements to areas where HPAI H5N1 is known to occur in wild birds or poultry.

⁽³⁾ *Areas at risk (wetlands in particular where links with high density poultry populations), previous positive findings as referred to in point 2 of Part 1 of Annex II to Commission Decision 2010/367/EC should be taken into account and if possible complemented by a map.*

3.2.1 WILD BIRD'S focussed on target species

Investigations according to the surveillance programme set out in Part 2 of Annex II to Decision 2010/367/EC

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

| NUTS (2) code/region (a) | Wild birds to be sampled (b) | Total number of birds to be sampled | Estimated total number of samples to be taken for active surveillance (c) | Estimated total number of samples to be taken for passive surveillance | |
|--------------------------|------------------------------|-------------------------------------|---|--|-----|
| Lithuania | 612 | 612 | 500 | 500 | X |
| Total | 612 | 612 | 500 | 500 | 112 |
| Add a new row | | | | | |

- (a) Refers to the place of collection of birds/samples. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) can not be used, region as defined in the programme by the Member State is requested. Please fill-in these values directly in the field.
- (b) General description of the wild birds are intended to be sampled in the framework of the active and passive surveillance.
- (c) Voluntary, to be included for information purposes, not eligible for cofinancing.

3.3 Sampling procedures and sampling periods

max 32000 chars :

Samples will be taken by the official veterinarians of the territorial State Food and Veterinary Services and by the official (authorized) vets. Sampling period will cover spring (from 1 of March till 1 of June) and autumn (from 15 of August till 15 of December) periods.

3.4 Laboratory testing : description of the laboratory tests used

max 32000 chars :

National Food and Veterinary Risk Assessment Institute (hereafter - Laboratory) performs detection of avian influenza (HPAI) disease antibodies against in

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

serum by Enzyme-linked immunosorbent assay (ELISA) and haemagglutination inhibition (HI) method, according to OIE recommendations (as described in "Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2012" Chapter 10.4).

Detection of antibodies against highly pathogenic avian influenza (HPAI) A viruses in serum by ELISA method:
The IDEXX AI Ab Test is an enzyme-linked immunosorbent assay (ELISA) designed to detect antibody to avian influenza viruses (A) in chicken serum. The IDEXX AI Ab Test detects antibody reactivity to 20 different subtypes, including 14 hemagglutinin glycoproteins and the H5N1 subtype. (http://www.idexx.com/view/xhtml/en_us/livestock-poultry/avian-influenza.jsf)

Detection of antibodies against highly pathogenic avian influenza (HPAI) A viruses in serum by haemagglutination inhibition (HI) method. (http://vla.defra.gov.uk/services/ser_tests.htm)

Detection of Influenza virus A subtypes H5 and H7 using one-step RT-PCR:
Laboratory performs detection of avian influenza (HPAI), according to OIE recommendations (as described in "Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2012" Chapter 10.4).

All avian influenza virus isolates of cases in wild birds shall be submitted to the CRL in accordance with Community legislation, unless derogation according to paragraph 4 of Chapter V under Differential diagnosis in the avian influenza Diagnostic Manual laid down in Commission Decision 2006/437/EC is granted. Viruses of H5/H7 subtype shall be submitted without delay and shall be subjected to the standard characterisation tests (nucleotide sequencing/IVP) according to the said avian influenza Diagnostic Manual.
All samples collected in the surveillance for avian influenza in wild birds shall be tested as soon as possible by molecular techniques if available and according to the diagnostic manual (Commission Decision 2006/437/EC). These tests shall only be carried out in laboratories able to guarantee quality assurance and using methods recognized by the CRL for avian influenza. In addition, methods used must be produced acceptable results in the most recent comparative ring test of national laboratories. Initial screening using M gene PCR is recommended, with rapid testing of positives for H5 (but within 2 weeks) and in case of a positive finding analysis of the cleavage site must be undertaken as soon as possible to determine whether or not it has a highly pathogenic avian influenza (HPAI) or a low pathogenic avian influenza (LPAI) motif. If H5 HPAI is confirmed further analysis to determine the N type must be done rapidly (even this can only provide evidence eliminating N1). (http://www.oie.int/fileadmin/Home/eng/Health_standards/tahc/2010/en_chapitre_1.10.4.htm)

Virus isolation testing:

Virus isolation testing is carried out on all suspicious cases. All viruses isolated will be sent to the CRL, where H5 and H7 subtypes will be subjected to

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

characterisation (IVPI and nucleotide sequencing). The CRL will provide the protocol for sending isolates to the CRL, and the reporting tables for collection of survey data.

4. Description of the epidemiological situation of the disease in poultry during the last five years

(max 32000 chars):

No cases of Avian influenza have been recorded in Lithuania during the last five years.

5. Description of the epidemiological situation of the disease in wild birds during the last five years

(max. 32000 chars):

No cases of Avian influenza in wild birds have been recorded in Lithuania during the last five years.

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

6. Measures in place as regards the notification of the disease

(max. 32000 chars):

"Law on Veterinary Activities" (Official Gazette, 1992, No. 2-15; 2010, No. 148-7563);
The Order of Director of State Food and Veterinary Service No. B1-281 of 12 April 2006 "Programme for animal contagious diseases" (Official Gazette, 2006, No. 48-1760).

7. Costs

7.1 Detailed analysis of the costs

7.1.1 Poultry

(max. 32000 chars):

Sampling shall cover a period appropriate to production periods for each poultry category. Broilers will not be included in this survey, as their short life means that they are unlikely to sero-convert before they are slaughtered. Samplers will be asked to take 10 samples from 59 poultry keeping places. All positive ELISA shall be confirmed by the National Food and Veterinary Risk Assessment Institute by a for virus isolation and molecular detection test (PCR) and a haemagglutination-inhibition test.

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

7.1.2 Wild birds

(max. 32000 chars):

112 dead birds will be investigated under the programme of passive surveillance and 500 birds will be hunted and investigated under the programme of active surveillance in the territory of the Republic of Lithuania. A total of 612 samples will be taken from birds in wildlife sanctuaries of international interest and game clubs. Migrating birds will be sampled during the early spring/early summer and autumn/early winter and will target mallard ducks and "first year" birds, where possible. The occurrence of mortality in several species at the same site shall be an additional factor to be considered. Wild birds found dead or shot shall be sampled for virus isolation, molecular detection and HI test for H5/H7 (double HI) will be applied.

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

7.2 Summary of the costs

7.2.1 Poultry surveillance

Detailed analysis of the cost of the programme - poultry

| Laboratory testing | Methods of laboratory analysis | Number of tests | Unitary test cost (per method) in € | Total cost (€) |
|--|--------------------------------|-----------------|-------------------------------------|----------------|
| ELISA test | | 590 | 2,3 | 1357 |
| agar gel immune diffusion test | | 0 | 0 | 0 |
| Haemagglutination-inhibition-test (HI) for H5 (specify number of tests for H5) | | 0 | 0 | 0 |
| Haemagglutination-inhibition-test (HI) for H7 (specify number of tests for H7) | | 0 | 0 | 0 |
| Virus isolation test | | 0 | 0 | 0 |
| PCR test | | 30 | 45 | 1350 |
| Other please specify here | | 0 | 0 | 0 |
| Add a new row | | | | |

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

| Sampling | | | | Total cost (€) |
|--|-------------------|-------------------|-------------------|----------------|
| | Number of samples | Unitary cost in € | | |
| Samples | 590 | 0.5 | 295 | X |
| Other measures | | | | |
| Other please specify here | 0 | 0 | 0 | Add a new row |
| Testing + Sampling + Other measures | 1 210 | | 3 002,00 € | |

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

7.2.2 Wild bird surveillance

Detail analysis of the cost of the programme - wild birds

| Laboratory testing | | Number of tests | Unitary test cost (per method) in € | Total cost (€) |
|--|--|-------------------|-------------------------------------|----------------|
| Methods of laboratory analysis | | | | |
| Virus isolation test | | 0 | 0 | 0 |
| PCR test | | 612 | 45 | 27540 |
| Haemagglutination-inhibition-test (HI) for H5/H7 | | 112 | 24 | 2688 |
| Other please specify here | | 0 | 0 | 0 |
| Other please specify here | | 0 | 0 | 0 |
| | | Add a new row | | |
| Sampling | | Number of samples | Unitary cost in € | Total cost (€) |
| Samples | | 612 | 5 | 3060 |

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.2

Attachments

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- 3) *The total file size of the attached files should not exceed 2 500Kb (+ 2.5 Mb). You will receive a message while attaching when you try to load too much.*
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