INSECT RESISTANCE MONITORING REPORT FOR *Sesamia nonagrioides* ASSOCIATED WITH MON 810 MAIZE CULTIVATION IN THE EU

Season 2021

CONTENTS

1. Introduction

Maize containing event MON 810 is transgenic improved maize expressing the Cry1Ab protein derived from *Bacillus thuringiensis* subsp. *kurstaki*, and conferring protection against certain lepidopteran insect pests such as *Ostrinia nubilalis* and *Sesamia nonagrioides*. Resistance development in targeted lepidopteran pests is a potential concern arising from the cultivation of MON 810 maize varieties. In order to maintain the benefits obtained from growing MON 810 maize varieties, Bayer, following directions described in the industry IRM (Insect Resistance Management) working group guidelines proposed to the competent authority (EU Commission), available since 2003 but published in 2007 (Alcalde et al., 2007) and subsequently updated as the EuropaBio harmonised IRM plan (EuropaBio, 2012; 2017; 2019), established an insect resistance monitoring program across Europe and in particular, in areas where commercial activity of MON 810 genetically improved maize is occurring or planned for the European targeted pests *O. nubilalis* and *S. nonagrioides*. The objective is to detect, in a timely manner, the potential development of resistance that could result in inadequate protection against the target species. This report focuses on the monitoring plan for *S. nonagrioides* in the 2021 growing season

The Mediterranean corn borer, *Sesamia nonagrioides* Lefèbvre (Lepidoptera: Noctuidae), is one of the most damaging pests of maize in Spain and the circum-Mediterranean countries (Castañera, 1986; Farinós et al., 2012). This species completes a variable number of generations per year depending on latitude, ranging from two in southern France to up to four in Morocco (Anglade, 1972, Eizaguirre and Fantinou, 2012). Larvae of the first generation are very destructive because they tunnel throughout the maize stem during the whole larval stage, causing great damage to maize seedlings and making their control particularly difficult. Second generation larvae infest older maize plants, damaging stems and ears, also causing significant yield losses (Velasco et al., 2004).

Routine monitoring for changes in the susceptibility of EU field populations of *S. nonagrioides* to the Cry1Ab protein has been carried out in the period 2004-2021[1]. During the period 2004-2015, the plan covered the three maize-growing areas in the EU where MON 810 hybrids have been grown and *S. nonagrioides* is present:

[1] https://ec.europa.eu/food/plant/gmo/post_authorisation/plans_reports_opinions_en

Northeast Iberia, Central Iberia and Southwest Iberia. Baselines were also gathered for other areas in Europe but no further samplings were performed in these areas because the adoption rate of Bt maize is less than 20% and monitoring resistance is not necessary according to the EuropaBio Harmonized IRM plan (EuropaBio, 2012). In Iberia, each target field population was initially monitored every two years, but for practical reasons they were divided into two groups so that each year sampling was carried out in one of the groups. The susceptibility of *S. nonagrioides* field populations to the Cry1Ab protein expressed in MON 810 maize varieties was estimated by means of dose-response bioassays. Measured endpoints of the tests were mortality (lethal concentration, LC) and moulting inhibition (moulting inhibition concentration, MIC). From 2010 onward, it was decided to switch from LC to MIC values because it was observed that variations in the susceptibility were better reflected in MIC_{50} values than in LC_{50} values, given the characteristics of the bioassay and the biology of the species. In both cases, MIC_{50} and LC_{50} values of field populations were compared with previous baseline susceptibility data established for this species or with a susceptible laboratory strain assayed with the same batch of toxin (González-Núñez et al., 2000; Farinós et al., 2004, 2018).

The harmonised IRM plan was subsequently updated to accommodate the upgrades in the regulatory framework, and to incorporate the available scientific information and new learnings gained from this and other IRM plans (EuropaBio, 2017; Farinós et al., 2018; Thieme et al., 2018; Bertho et al., 2020). The revised plan establishes that sampling for resistance monitoring will take place in areas where the Bt maize adoption is over 60% and where the target pest is present. Currently, this situation only occurs in the North-eastern Spain within the EU. Since *S. nonagrioides* and *O. nubilalis* are multivoltine species, the revised plan proposes that monitoring for these corn borers in this area should be carried out on an annual basis. This revised plan was put in practice for the first time during the season of 2016, being this season (2021) the sixth time.

In addition, the EFSA Scientific Opinion (EFSA GMO Panel, 2017) and Statement (EFSA 2018) on the annual post-market environmental monitoring (PMEM) reports on the cultivation of genetically modified maize MON 810 in 2015 and 2016, respectively, included the recommendations to (1) perform annual sampling of target pests in the North-eastern Spain, the area where deployment of Bt-maize is the highest and where resistance is likely to evolve more quickly; and (2) set a maximum detection threshold for resistance allele frequency at 3% to enable the early detection of resistance so that

alternative management measures can be implemented in time to delay the development of resistance.

In accordance with these recommendations and following the revised harmonized IRM plan (EuropaBio, 2017), from the 2016 season onwards the collection of field larvae has focused in North-eastern Spain (including the Autonomous Communities of Aragón, Cataluña and Navarra), where the adoption rate of Bt maize is over 60%. Moreover, a diagnostic concentration bioassay (Sims et al., 1997; Marçon et al., 2000) has been used to monitor for changes in susceptibility to the Cry1Ab protein in *S. nonagrioides* field populations from North-eastern Spain. The aim of using this methodology is that a high number of field-collected individuals are represented in the laboratory bioassays as F1 larvae, thereby helping to decrease the detection limit for resistance allele frequency.

The tasks carried out in the 2021 maize growing season were the following:

1. Collection of larvae of *S. nonagrioides* in three different zones from North-eastern Spain to be used in: i) diagnostic concentration bioassays, to compare the susceptibility (in terms of moult inhibition) of the field populations with that of a susceptible laboratory strain and with the hypothetical value of 99%; and ii) plant bioassays, to compare larval mortality when feeding on Bt maize *vs.* conventional maize.

2. Collection of larvae of *O. nubilalis* in three different zones from North-eastern Spain to be sent to the laboratory BTL GmbH Sagerheide (Germany), which is carrying out the European resistance monitoring programme of *O. nubilalis* for MON 810 maize.

3. Analysis of the susceptibility to Cry1Ab of laboratory strains of *S. nonagrioides and O. nubilalis* by means of dose-response bioassays, aiming at verifying the activity of the batch of protein used in the bioassays.

2. Materials and Methods

2.1. Insect collection

Last-instar larvae of *S. nonagrioides* and *O. nubilalis* were collected from three sampling zones for each species in the North-eastern Spain (NE Spain), each zone comprising at least three maize fields located as close together as possible. A minimum of 1000 larvae were targeted for collection per species, about 350 larvae

collected in each of the three sampling zones and, if possible, a minimum of 50 larvae per maize field. However, due to the habitual high mortality rates of field larvae when kept in the laboratory, an effort has been made to collect as many larvae as possible to achieve the target for maximum detection threshold for resistance allele frequency of 3%.

The process of identifying fields for insect collection starts by contacting local field technicians of seed companies that commercialize MON 810 in Spain. They are asked if they are aware of the presence of corn borers (*Sesamia* and/or *Ostrinia*) in the area to be sampled, for which they may consult a number of growers about the detection of corn borer damage in their fields. When that is the case, the growers are asked for permission to enter their fields during one to three days to collect larvae, which requires destructive sampling of the maize stalk. The amount of maize plants used depends on the presence of one or two species, the severity of the attack, etc.

Last-instar larvae of both corn borers were collected following standard operative procedures (SOP) of each species (EuropaBio, 2017). The samples were collected at the end of the maize-growing season, during September and October 2021, from refuges and fields of conventional maize adjacent to MON 810 maize, by cutting the stalk of the maize plants and taking only one larvae of each species per plant to avoid collecting siblings.

2.2. Insect rearing

Field collected larvae were brought to the laboratory, dipped in a solution containing 1% sodium hypochlorite (bleach) to avoid contamination by pathogens and placed in 21x16x4 cm plastic boxes. Both species were fed on an artificial diet established from that described by Poitout and Buès (1970) with some modifications (**Tables 1** and **2**). Immediately after asepsis, collected larvae of *O. nubilalis* were sent to BTL GmbH Sagerheide (Germany) to be analyzed there.

Larvae of *S. nonagrioides* were in diapause at the time of collection, so they were placed on a rearing chamber (Sanyo MLR-350 H, Sanyo, Osaka, Japan) at 14 ± 1ºC, 70 ± 5% relative humidity and a photoperiod of 12:12 hours (L:D). They were kept at these conditions until the larvae showed signs of diapause break. Then, larvae were placed under conditions 28 ± 1ºC, 70 ± 5% relative humidity and continuous light until pupation. The sex was determined at the pupal stage and a variable number of couples from the same zone (normally from 3 to 6), in function of the day of adult emergence,

were confined in ventilated plastic cylinders (12 cm diameter x 30 cm high) containing 5-7 maize seedlings for oviposition at standard rearing conditions (25 ± 1ºC, 70 ± 10% relative humidity and a photoperiod of 16:8 hours (L:D). After 7 days the egg masses were collected, placed into ventilated plastic boxes containing wet filter paper and incubated under the same conditions. Neonate larvae (< 24 h old) were utilized in the bioassays.

2.3. Laboratory strains

Reference susceptible strains of *S. nonagrioides* and *O. nubilalis* have been maintained under laboratory conditions to serve as control in this study. As a general rule, these populations are formed from individuals collected in non-Bt fields from Galicia (Northwest of Spain), where Bt maize has never been commercially grown, so corn borers have low or no selection pressure, making them a good option to be used as the reference strain. Formerly, Galician populations of *S. nonagrioides* had not shown differences in susceptibility to the Cry1Ab toxin with respect to those of other Spanish populations in bioassays carried out in 1998 (González-Núñez et al., 2000) and in 2011 (unpublished results).

Populations maintained for many years in the laboratory typically suffer excessive inbreeding (Roush, 1986). To ensure that this does not influence bioassay results, new individuals are needed every few years. Two stocks of *O. nubilalis* and *S. nonagrioides* originated from field populations collected in Galicia in 2019 and 2020, respectively, have been used this season. The collection was carried out with the assistance of La Misión Biológica de Galicia (MBG, CSIC) staff, and they have been incorporated as reference populations after adaptation to the artificial diet and to laboratory conditions (Hoffmann & Ross, 2018).

In the laboratory, a minimum of 300 adults are crossed every generation. Neonate larvae are taken from all the oviposition cages formed with the adults of the previous generation, unless any of them have symptoms of any disease, in which case it is removed. In addition, the larvae are periodically checked for the presence of pathogens (namely *Nosema* sp.) by inspecting a number of them in slides under the microscope and by molecular methods (PCR).

2.4. Cry1Ab protein

Two batches of Cry1Ab protein have been used since the start of the MON 810 monitoring plan in 2004. The first batch (B1) was provided by Bayer in 2003 (2.03 mg/ml in sodium bicarbonate buffer, pH 10.5; purity 95%). The second batch (B2) (1.8 mg/ml in 50 mM sodium bicarbonate buffer, pH 10.25; purity 91%) has been provided at different times: B2-1 was sent in October 2011, B2-2 in February 2014, B2-3 in April 2016, B2-4 in July 2017, B2-6 in July 2018, B2-7 in September 2019, B2-8 in September 2020 and B2-9 in July 2021. Stock solutions were prepared from the original and kept in the freezer at -80ºC until used. Aliquots were thawed only when the bioassay was ready to be performed. To prepare the test concentrations, a sodium bicarbonate buffer (50 mM) with pH 10.25 was used. The lot of Cry1Ab toxin B2-9 has been used for the bioassays of this season.

2.5. Bioassays

2.5.1. Susceptibility of the reference strains of *S. nonagrioides* and *O. nubilalis* to the Cry1Ab protein in dose-response (DR) bioassays

The bioassays were carried out in accordance with the methods described by Farinós et al. (2004), using “Bio-Assay Tray-128 Cells (BAW128)” plastic trays (Frontier Scientific Services Agriculture, DE, USA). Each tray contains 128 wells, where 0.5 ml of rearing diet is placed and flattened, corresponding to a surface of 1.77 cm^2 and a height of about 10 mm. Once solidified, 50 µl of a solution containing different concentrations of Cry1Ab were added to the surface of the diet. The controls consisted of the sodium bicarbonate buffer solution used to dilute the toxin. After drying the wells under a laminar flow hood, one neonate larva (<24 h) was placed in each well using a fine paintbrush and it was covered with a breathing adhesive cover “Bio-Assay Tray Lid-16 Cells (BACV16)” (Frontier Scientific Services Agriculture, DE, USA). The trays were incubated in rearing chambers at 25 ± 1ºC, 70 ± 5% relative humidity and total darkness. Measured endpoint of the test in both species was moulting inhibition (moulting inhibition concentration, MIC) relative to the negative control after 7 days of exposure, where moulting inhibition equals larvae that have either died or not moulted to the 2nd larval instar (L2) after 7 days.

The concentration ranges were comprised between 2 and 128 ng Cry1Ab/cm^2 for *S. nonagrioides* and between 0.5 and 64 ng Cry1Ab/cm^2 for *O. nubilalis*. To determine the susceptibility of each population, 5 to 7 different concentrations resulting in

moulting inhibition higher than 0% and below 100% were used. Three replicates were prepared for each concentration and the control for both species. Each replicate consisted of 32 larvae per concentration (64 for controls), giving 96 larvae for each concentration tested (192 for controls). For each replicate neonate larvae from different oviposition cages were used. The MIC_{50} values obtained for both *O. nubilalis* and *S. nonagrioides* were compared with those obtained with the reference populations in previous years.

2.5.2. Susceptibility of *S. nonagrioides* to the Cry1Ab protein in diagnostic concentration (DC) bioassays

A diagnostic concentration (DC) of 1091 ng Cry1Ab/cm^2, intended to cause moulting inhibition between 99 and 100% to first-instar (L1) larvae of *S. nonagrioides*, was used for DC bioassays to measure susceptibility to the Cry1Ab protein. The value of the DC was estimated by using all the available data of MIC bioassays performed with larvae collected in NE Spain over the seasons 2009, 2011, 2013 and 2015. Hence, the resulting value represented the response of more than 4300 larvae in four dose-response bioassays. This DC has been used from the 2016 campaign onwards, as no new dose-response bioassay data have been available since then.

The susceptibility to the protein Cry1Ab by the use of DC bioassays was tested on F1 progeny of the field populations collected in NE Spain in 2021 and on the reference laboratory strain of *S. nonagrioides,* which served as control. The methodology of the bioassay was the same as that explained above (Section 2.5.1.), with the exception that only DC (1091 ng Cry1Ab/cm^2) and control (sodium bicarbonate buffer solution) are tested. A minimum of 1000 neonates per field population (zone) were treated, and a minimum of 100 neonates per zone were used as controls. Moulting inhibition was recorded after 7 days.

It was ensured that as many field individuals as possible were represented in the bioassays. To this end, individuals of *S. nonagrioides* from each of the three zones were tracked and the following variables were recorded: number of field collected larvae, number of emerged adults, number of oviposition cages settled and used in bioassays and number of neonates used in the bioassays from each oviposition cage (treated and controls). The number of egg masses used for the bioassays was maximized by doing the bioassay at a daily basis, whenever neonates of an oviposition cage emerged (infertile oviposition cages are not represented in the bioassay). This

procedure forced to (i) estimate in advance the number of oviposition cages to be set, (ii) determine the approximate number of neonates to be used from each oviposition cage (treated and controls), and (iii) prepare a lot of small bioassay arenas to conduct the bioassay little by little. For all that, the DC bioassay with the F1 generation extended for about ten weeks.

2.5.3. Larval development on MON 810 tissue: Plant bioassays

Plant bioassays were performed to verify that there were no resistant individuals in the field-collected populations even if some larvae from the DC bioassay had moulted to the 2nd larval instar.

About two-hundred neonates (not used in the DC bioassays) of each oviposition cage of the F1 generation coming from the three NE Spain zones and of the laboratory strain were exposed to MON 810 fresh leaves, and about 10 larvae of each cage, which served as control, were exposed to conventional maize leaves. Larvae were kept in plastic boxes provided with new maize leaves without the central nerve and they were allowed to feed *ad libitum*. If necessary, fresh tissue was added every 2-3 days. Moulting to the 2nd larval instar was recorded during 10 days.

It was ensured that all the Bt plants used in the bioassay were transgenic by two means: 1) use of ImmunoStrip® for Bt-Cry1Ab/1Ac (Agdia Inc., Elkhart, IN); 2) testing each plant with susceptible neonates of *O. nubilalis*: 10-15 neonates per plant were fed ad libitum on maize tissue, and for a plant to be used in bioassays, mortality after one week should be 100%. This experiment was performed at the same conditions of insect culture: 25 ± 1ºC, 70 ± 10% relative humidity and a photoperiod of 16:8 hours (L:D).

2.5.4. Confirmatory experiments

Different experiments aimed at confirming that survivors of DC and plant bioassays from field-collected populations were not resistant individuals were performed when necessary after the F1 bioassays.

Firstly, all L2 larvae recovered alive after 7 days in the DC bioassay were placed in plastic boxes of 9 cm in diameter and 3 cm height, with those coming from the same oviposition cage grouped together. Then, they were fed *ad libitum* on Bt maize leaves, following the same procedure of section 2.5.3. If any of these larvae fed on MON 810 during 10 days were able to moult to the 3rd larval instar and they were alive at the end of this period, their siblings (not used in the F1 bioassays) were fed on an artificial diet

and reared until the next generation (F2) to perform new DC and plant bioassays, as explained in sections 2.5.2. and 2.5.3.

In the case of plant bioassays, if any neonate fed on MON 810 during 10 days were able to moult to the 2nd larval instar and they were alive at the end of this period, their siblings (not used in the F1 bioassays) were fed on an artificial diet and reared until the next generation (F2) to perform a new DC bioassay and plant bioassay, as explained in sections 2.5.2. and 2.5.3.

The confirmatory experiments were carried out under the same conditions of temperature, humidity and light as the insect culture and bioassays described above.

2.6. Statistical analysis

The results of moulting inhibition of laboratory populations at different concentrations of Cry1Ab (dose-response bioassays) were adjusted by probit weighted regression lines. The moulting inhibition concentrations (MICs) for 50% (MIC_{50}) and 90% (MIC_{90}) of each population were estimated together with their 95% confidence limits using PoloPlus 1.0 (LeOra Software, 2002-2022). Mortality of the control must be below 25% for *S. nonagrioides* and 20% for *O. nubilalis*, so that the replicate is included in the statistical analysis. The bioassay was considered valid if the obtained MIC_{50} was comprised between at least 2 concentrations above it and 2 concentrations below it, from all the concentrations tested.

The average percentage of moulting inhibition of neonates after treatment at the diagnostic concentration (DC) was estimated to determine if it was significantly lower than (i) the percentage of moulting inhibition observed in the susceptible reference strain after treatment at the same DC and (ii) the expected generic value of 99%. Values were compared by a one-sample t-test and a one-tailed probability distribution (IBM SPSS Statistics 26). Moulting inhibition values of each zone were corrected with Abbott's formula (Abbott, 1925) and logit transformed before analyses.

3. Results and Discussion

The stepwise approach followed to perform the bioassays is shown in **Annex I**.

3. Results and Discussion

3.1. Collection of larvae and insect rearing

This campaign, the technicians involved in the collection of field larvae for *S. nonagrioides* and *O. nubilalis* carried out about 220 hours of fieldwork, travelled over 2700 km and made a total of three round trips, one for each different field zone.

A total of 1699 last-instars larvae of *S. nonagrioides* were collected between September and October 2021 from three different Zones in NE Spain (542, 553 and 604 larvae from Zones 1, 2 and 3, respectively; **Table 3**). A map showing the sampling points for *S. nonagrioides* is displayed in **Annex IIa.** Nine, six and nine fields in Zones 1, 2 and 3, were searched, respectively, although larvae were successfully collected in four fields in each Zone **(Figure 1, Annex IIb**). The maximum distance between successfully sampled fields was about 7, 7 and 4 Km within Zones 1, 2 and 3, respectively (**Annex IIb**). Thus, the minimum of 1000 larvae targeted for collection could be fulfilled.

Larvae of *O. nubilalis* were collected between September and October 2021 from the same three Zones in the North-eastern Spain, yielding a total of 811 larvae (304, 503 and 4 larvae from Zones 1, 2 and 3, respectively; **Table 4**). A map showing the sampling points for *O. nubilalis* is displayed in **Annex IIIa.** Larvae were mainly gathered in one field in Zone 1 and four fields in Zone 2 (**Figure 1, Annex IIIb**), although the number of fields surveyed was higher (nine in Zone 1, six in Zone 2 and fourteen in Zone 3). The maximum distance between successfully sampled fields was about 7 Km within Zone 2 (**Annex IIIb**). Even though 29 maize fields have been sampled in all three Zones, the minimum of 1000 larvae targeted for collection could not be reached for this species.

A number of larvae above the minimum target was collected so that the highest possible number of field individuals would be represented in the bioassays and thus reach the maximum detection threshold of 3% of the resistance allele frequency. However, this is not always achieved, usually due to high mortality rates of field larvae reared in the laboratory. There are different reasons that could explain the high mortality of both lepidopteran species during the diapause period (about 3 months). In the case of *S. nonagrioides*, larval mortality in the laboratory has been found to be higher when reared under diapause conditions than that observed when reared under normal maintenance conditions (Fantinou and Tsitsipis, 1999). In addition, mortality during the diapause time may increase due to the transmission of diseases or to the

emergence of parasitoids from larvae carried from the field (Eizaguirre et al., 1990; Fantinou and Tsitsipis, 1999; Monetti et al., 2003). Another cause could be a low adaptation to artificial breeding conditions (Hoffmann and Ross, 2018), which may be different depending on the population, even if they are of the same species (Carpenter and Bloem, 2002). To reduce these effects as much as possible in *S. nonagrioides*, the following measures are taken: (a) a low number of larvae per box is maintained to reduce mortality by limiting disease spread (Fantinou & Tsitsipis, 1999); (b) the vermiculite of the boxes is frequently renewed during the diapause period to prevent fungal growth; (c) those larvae from boxes suspected of containing larvae with pathogens go through additional asepsis (dipping of larvae in 1% bleach solution); and (d) the diet in the rearing boxes is renewed once a week in order not to disturb diapause conditions.

3.2. Susceptibility of the reference strains to the Cry1Ab protein in dose-response bioassays

The susceptibility to Cry1Ab toxin of the laboratory population of *S. nonagrioides* was performed with 768 neonates using a dose-response bioassay, resulting in a MIC_{50} value of 25 (14-40) ng Cry1Ab/cm^2, (**Table 5**, **Figure 2a**), which is in the range of MIC_{50} values obtained with laboratory populations in previous years with the same batch of toxin (between 5 and 30 ng Cry1Ab/cm^2; **Table 6**). However, the MIC_{90} obtained, 292 (139-1336) ng Cry1Ab/cm^2, was higher than in previous years, when it ranged between 42 and 233**.** In addition, the 95% confidence limits were also wider than in previous years, probably because moult inhibition at the highest concentration used did not reach 90% in any of the 3 replicates, so there is not a good fit to the regression line at that point (**Table 6)**.

A number of 959 neonates of the *O. nubilalis* laboratory strain were used for the Cry1Ab susceptibility assessment bioassay. The MIC_{50} value obtained with the reference strain was 1.7 (1.4-2.0) ng Cry1Ab/cm^2 (**Table 5**, **Figure 2b**), in the range of values obtained with laboratory strains with the same batch of toxin (0.8-5.4 ng Cry1Ab/cm^2; **Figure 3**).

Variations in laboratory-reared insects regarding their susceptibility to pesticides or insecticidal proteins, as we have observed historically during this monitoring program, are not unusual. Different reasons have been proposed, such as diverse geographical sources of individuals, varying testing personnel, different protein preparations, etc.

(Robertson et al., 1995; Marçon et al., 1999; Da Silva et al., 2016; Farinós et al., 2018). Even so, MIC values of the control laboratory strains have been, in general, very consistent in the interval of years examined using the same batch of toxin (B2), being the maximum magnitude of variation 6- and 7-fold for *S. nonagrioides* (**Table 6**) and *O. nubilalis*, (**Figure 3**)**,** respectively.

3.3. Diagnostic concentration bioassays

From the 1699 *S. nonagrioides* last-instars larvae collected, 531 (31.3%), combining larvae and pupae, died in the process of rearing in the laboratory, mainly during the diapause period. In addition, 51 adults (3.0%) did not emerge in the date range for oviposition cages or had some malformation upon emergence, so they were not used in the bioassays (**Table 7**)**.** Thus, of the 1168 adults that emerged, 1117 did so between 10th November 2021 and 10th February 2022 and were placed in 103 oviposition cages for mating. The offspring of 1076 (63%) of these adults was used in the bioassays and treated with the diagnostic concentration (DC) of 1091 ng Cry1Ab/cm^2 (**Table 8**). Therefore, the detection limit for resistance allele frequency in 2021 is 0.030 (3.0%), calculated considering the model developed by Andow and Ives (2002).

Of the total F1 neonates originated from the field collected larvae, 3467 were used in the bioassays. The DC (1091 ng Cry1Ab/cm^2) caused a corrected moulting inhibition (MI) of 99.64%, 98.88% and 96.28% in Zones 1, 2 and 3, respectively (**Table 8**).

No statistically significant differences were observed between field-collected populations MI value (98.27 ± 1.02%) and the expected MI value of 99% (t = 0.2028, df = 2, p = 0.429), nor were they found between field-collected populations and the laboratory strain (99.20%) MI values (t = 0.5276, df = 2, p = 0.355) (**Table 9**). Likewise, no significant differences were found between these values in 2016, 2018 and 2020. On the contrary, significant differences between the MI value of the field populations and the expected MI value were obtained in 2017 and 2019, but only in 2017 the MI value of the field populations was significantly lower than that of the laboratory strain (**Table 9**). Therefore, no trend over time is observed in terms of changes in the susceptibility of populations from NE Spain to the Cry1Ab toxin **(Figure 4)**.

It is noteworthy that in four (2017, 2018, 2019 and 2020), of the six carried out campaigns focused on NE Spain, the percentage of moulting inhibition of *S. nonagrioides* obtained with the laboratory susceptible strain was below the expected

value of 99% (97.69%, 97.75%, 97.02% and 98.67%, respectively), and only in 2016 and 2021 the MI (99.20% both years) was over the expected value (**Table 9**). Fluctuations of about 6-fold for both LC_{50} and MIC_{50} were also found in the laboratory strain during the period in which monitoring was performed by means of dose-response bioassays (2004–2015), although no trends were observed over time (**Table 6**). This underlines the importance of testing the field populations against a reference population from areas where Bt maize is not grown, enabling the correct interpretation of the results.

3.4. Larval development on MON 810 tissue: plant bioassays

18,950 F1 first-instar larvae of *S. nonagrioides*, from the populations collected in three zones in NE Spain in 2021, and 5200 from the laboratory strain were fed *ad libitum* on MON 810 tissue to test if they were able to moult to the 2nd larval instar within ten days. As a control, 970 neonates of these field-collected populations and 260 neonates of the laboratory strain were reared on conventional maize. During the assay none of the larvae from the field-collected populations nor from the laboratory strain were able to moult to the 2nd larval instar and survive after 10 days feeding on Bt maize. Most larvae fed on conventional maize from the field and the laboratory populations (98.5% and 97.3%, respectively) moulted to 2nd or 3rd larval instar **(Table 10)**.

3.5 Confirmatory experiments

A total of 54 (1.56%) larvae from 16 oviposition cages reached the 2nd larval instar in the F1 DC bioassay. Consequently, the following confirmatory experiments were conducted.

The 54 surviving L2 larvae from the DC bioassays were individualized in boxes and fed on MON 810 leaves. One of these larvae, from a single oviposition cage from Zone 2, moulted to the 3rd larval instar and survived 10 days feeding on Bt maize leaves (although they died without reaching the fourth larval instar; **Table 11a**). Thus, the siblings of this larva (about 125 larvae from the original oviposition cage) were raised on artificial diet up to the next generation (F2). As a result, 168 F2 neonates were treated with the diagnostic concentration (1091 ng/cm^2). One of the treated larvae moulted to the 2nd larval instar after 7 days, but it did not survive for ten days when it was subsequently fed on MON 810 maize. In addition, 1200 F2 neonates, siblings of the above, were fed on MON 810 maize and 60 neonates, used as controls, on

conventional maize. After 10 days, none of the larvae fed on Bt tissue were able to moult, whereas 58 larvae (97%) fed on conventional maize had moulted to 2nd or 3rd larval instar (**Table 11b**).

4. Summary of results

1. Monitoring for changes in the susceptibility of EU field populations of *S. nonagrioides* and *O. nubilalis* to the Bt Cry1Ab in 2021 has been focused for the sixth time in the North-eastern Spain, where the adoption rate of Bt maize in 2021 was over 60%. A total of 1699 larvae of *S. nonagrioides* and 811 larvae of *O. nubilalis* were collected in three sampling zones. The minimum of 1000 larvae targeted for collection could be fulfilled for *S. nonagrioides*, but not for *O. nubilalis,* due to the low levels of larval infestation in most of the sampled fields. Larvae of *O. nubilalis* were sent to the laboratory BTL GmbH Sagerheide (Germany) for testing their susceptibility to the Cry1Ab protein.

2. From the 1699 larvae of *S. nonagrioides* collected, 1168 adults (69%) emerged, of whom 1117 mated. The offspring of 92% of these adults (1076) was used in the bioassays and treated with the diagnostic concentration (DC) of 1091 ng Cry1Ab/cm^2, intended to cause moulting inhibition $\geq$ 99% to first-instar larvae of *S. nonagrioides*. These values indicate that despite the application of best practices in larvae rearing only 63% of the field collected larvae were represented in the DC bioassays. The detection limit for resistance allele frequency in field populations of *S. nonagrioides* in 2021 is 0.030 (3.0%).

3. The values of the detection limit for resistance allele frequency in field populations of *S. nonagrioides* in the six last seasons, (3.3, 3.7, 4.2, 3.4, 3.6 and 3.0 in 2016, 2017, 2018, 2019, 2020 and 2021, respectively) *vs.* the number of larvae collected in the field each year (1364, 1452, 1490, 1644, 1569 and 1699), highlight the technical difficulties that can be encountered, depending on different factors, in each campaign, regardless of the number of larvae collected.

4. The treatment with the DC caused mean moulting inhibition of 98.27% (S.E. 1.02%) to F1 neonates from the field collected larvae of the NE Spain. This outcome was not significantly different from the expected value of 99% (t = 0.2028, df = 2, p = 0.355) nor from the laboratory strain moulting inhibition value (99.20%) (t = 0.5276, df = 2, p = 0.429).

5. None of the 18,950 neonates of the F1 generation of the field collected populations was able to moult to the 2nd larval instar and survive after 10 days feeding on MON 810 leaves.

6. Laboratory *S. nonagrioides* and *O. nubilalis* strains showed susceptibility levels to the batch B2-9 of the Cry1Ab toxin (MIC_{50} values of 25 and 1.7 ng Cry1Ab/cm^2, respectively) comparable with those obtained from laboratory strains in previous years.

5. Concluding remarks

Considerable effort has been made over the past six seasons to collect increasing numbers of last instar larvae of *S. nonagrioides*. However, this was the first year in which the required limit of detection of resistance allele frequency of 3% could be reached, highlighting the technical difficulties encountered in achieving this goal. In addition, it should be mentioned that the number of larvae that can be kept in the laboratory after being collected in the field is limited, for reasons of space, facilities and handling.

The moult inhibition (98.27%) of *S. nonagrioides* F1 neonates from NE Spain in 2021, treated with a diagnostic concentration (DC), was not significantly different than the hypothetical value of 99%, nor from the moult inhibition value (99.20%) caused to neonates of a laboratory strain with the same DC. The results obtained in recent years, as well as the experience accumulated in the more than 15 years of MON 810 monitoring, underscore the importance of maintaining a susceptible laboratory strain against which the field populations should be compared, enabling correct interpretation of the results.

In summary, the results obtained indicate that there are no evidences of resistance development of *S. nonagrioides* to MON 810 maize in NE Spain. Our results are in line with those revealed in ten years of surveys of farmers, in which no evidence of any unexpected adverse effect associated with the cultivation of MON 810 was found (Bertho et al., 2020).

6. References

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7. Tables and figures

Table 1. Artificial diet used for *S. nonagrioides*.

Components	Amount	Provider
Distilled H_2O	1 l	
Agar	26 g	Conda Pronadisa
Maize flour	160 g	Santiveri
Wheat germ	40 g	Santiveri
Yeast	43 g	Santiveri
Ascorbic acid	6 g	Panreac
Benzoic acid	1.25 g	Merck Millipore
Nipagin (Methyl p-hidroxibenzoato)	1 g	Sigma-Aldrich
Wesson's salts mixture	1.55 g	Sigma-Aldrich

Table 2. Artificial diet used for *O. nubilalis*.

Components	Amount	Provider
Distilled H_2O	1 l	
Agar	24 g	Conda Pronadisa
Maize flour	168 g	Santiveri
Wheat germ	42 g	Santiveri
Yeast	45 g	Santiveri
Ascorbic acid	9 g	Panreac
Benzoic acid	3 g	Merck Millipore
Nipagin (Methyl p-hydroxybenzoate)	1.5 g	Sigma-Aldrich
Sorbic acid	1.2 g	Sigma-Aldrich

7. Tables and figures

Table 3. *Sesamia nonagrioides* larvae collection details for the 2021 season in NE Spain.

Zone	Field	Province [a]	Postal Code	Date	Surface (Ha) [b, c]	Distance to the nearest MON810 field (m) [d]	No of larvae collected
1	2021-Sariñena 1	HU	22200	25-28/10/2021	32		52
	2021-Sariñena 2	HU	22200	25-28/10/2021	27		0
	2021-Sariñena 3	HU	22200	25-28/10/2021	35		172
	2021-Sariñena 4	HU	22200	25-28/10/2021	15.7		173
	2021-Sariñena 5	HU	22200	25-28/10/2021	7		0
	2021-Sariñena 6	HU	22200	25-28/10/2021	15		0
	2021-Sariñena 7	HU	22200	25-28/10/2021	17		0
	2021-Sariñena 8	HU	22200	25-28/10/2021	15.2		0
	2021-Sariñena 9	HU	22200	25-28/10/2021	48		145
	Total						**542**
2	2021-Candasnos 1	HU	22591	27-29/09/2021	4.5	80	136
	2021-Candasnos 2	HU	22591	27-29/09/2021	11.3	260	31
	2021-Candasnos 3	HU	22591	27-29/09/2021	20.8	500	122
	2021-Candasnos 4	HU	22591	27-29/09/2021	37.7	300	137
	2021-Candasnos 5	HU	22591	27-29/09/2021	16.6	100	106
	2021-Candasnos 6	HU	22592	27-29/09/2021	6.6	240	21
	Total						**553**
3	2021-Mendigorria 1	NA	31140	4-6/10/2021	31.0	300	192
	2021-Mendigorria 2	NA	31140	4-6/10/2021	3.0	10	198
	2021-Mendigorria 3	NA	31140	4-6/10/2021	4.5	0	83
	2021-Mendigorria 4	NA	31140	4-6/10/2021	3.1	0	33
	2021-Mendigorria 5	NA	31150	4-6/10/2021	12.6	0	70
	2021-Mendigorria 6	NA	31150	4-6/10/2021	5.2	0	5
	2021-Mendigorria 7	NA	31150	4-6/10/2021	8.1	0	17
	2021-Mendigorria 8	NA	31150	4-6/10/2021	3.3	0	4
	2021-Mendigorria 9	NA	31150	4-6/10/2021	3.6	0	2
	Total						**604**
	GRAND TOTAL						**1699**

[a] Provinces: HU = Huesca; NA = Navarra.
[b] Data are approximate.
[c] Area of the whole field, even though larvae were collected on the refuge area of the field.
[d] There could be other nearer fields that are not known by the technician and/or the farmer. "0" means that it is adjacent to a MON 810 field.

7. Tables and figures

Table 4. *Ostrinia nubilalis* larvae collection details for the 2021 season in NE Spain.

Zone	Field	Province [a]	Postal Code	Date	Surface (ha) [b, c]	Distance to the nearest MON810 field (m) [d]	No of larvae collected
1	2021-Sariñena 1	HU	22200	25-28/10/2021	32		0
	2021-Sariñena 2	HU	22200	25-28/10/2021	27		0
	2021-Sariñena 3	HU	22200	25-28/10/2021	35		27
	2021-Sariñena 4	HU	22200	25-28/10/2021	15.7		0
	2021-Sariñena 5	HU	22200	25-28/10/2021	7		0
	2021-Sariñena 6	HU	22200	25-28/10/2021	15		0
	2021-Sariñena 7	HU	22200	25-28/10/2021	17		0
	2021-Sariñena 8	HU	22200	25-28/10/2021	15.2		0
	2021-Sariñena 9	HU	22200	25-28/10/2021	48		277
	Total						**304**
2	2021-Candasnos 1	HU	22591	27-29/09/2021	4.5	80	110
	2021-Candasnos 2	HU	22591	27-29/09/2021	11.3	260	6
	2021-Candasnos 3	HU	22591	27-29/09/2021	20.8	500	109
	2021-Candasnos 4	HU	22591	27-29/09/2021	37.65	300	50
	2021-Candasnos 5	HU	22591	27-29/09/2021	16.6	100	212
	2021-Candasnos 6	HU	22592	27-29/09/2021	6.6	240	16
	Total						**503**
3	2021-Mendigorria 1	NA	31140	4-6/10/2021	31.0	300	0
	2021-Mendigorria 2	NA	31140	4-6/10/2021	3.0	10	1
	2021-Mendigorria 3	NA	31140	4-6/10/2021	4.5	0	1
	2021-Mendigorria 4	NA	31140	4-6/10/2021	3.1	0	0
	2021-Mendigorria 5	NA	31150	4-6/10/2021	12.6	0	2
	2021-Mendigorria 6	NA	31150	4-6/10/2021	5.2	0	0
	2021-Mendigorria 7	NA	31150	4-6/10/2021	8.1	0	0
	2021-Mendigorria 8	NA	31150	4-6/10/2021	3.3	0	0
	2021-Mendigorria 9	NA	31150	4-6/10/2021	3.6	0	0
	2021-Aibar 1	NA	31460	4-6/10/2021	39.0	100	0
	2021-Aibar 2	NA	31460	4-6/10/2021	31.0	10	0
	2021-Aibar 3	NA	31460	4-6/10/2021	20.0	10	0
	2021-Aibar 4	NA	31460	4-6/10/2021	14.0	0	0
	2021-Aibar 5	NA	31460	4-6/10/2021	21.5	0	0
	Total						**4**
	GRAND TOTAL						**811**

[a] Provinces: HU = Huesca; NA = Navarra
[b] Data are approximate.
[c] Area of the whole field.
[d] There could be other nearer fields that are not known by the technician and/or the farmer. "0" means that it is adjacent to a MON 810 field.

7. Tables and figures

Table 5. Susceptibility to Cry1Ab toxin of the reference laboratory populations of *S. nonagrioides* and *O. nubilalis*.

Species	Toxin batch	n	Slope ± SE	χ^2	d.f.	MIC_{50}[a] (CI 95%)	MIC_{90}[a] (CI 95%)
S. nonagrioides	B2-9	768	1.2 ± 0.1	36.2	16	25 (14-40)	292 (139-1336)
O. nubilalis	B2-9	959	3.7 ± 0.3	28.0	13	1.7 (1.4-2.0)	3.7 (3.0-5.0)

[a] 50% and 90% moulting inhibition concentrations (MIC_{50} and MIC_{90}) and their 95% confidence intervals (CI 95%) are expressed in ng Cry1Ab/cm^2.

Table 6. Susceptibility to Cry1Ab toxin of laboratory populations of *S. nonagrioides* between 2004 and 2021. The bioassay performed during the present campaign is shaded.

Population	Season	Batch of toxin	MIC_{50}[a] (CI 95%)	MIC_{90}[a] (CI 95%)
Laboratory	2004	B1	18 (11-25)	99 (66-208)
Laboratory	2007	B1	16 (11-22)	94 (69-147)
Laboratory	2008-9	B1	19 (10-30)	120 (76-255)
Laboratory	2010	B1	8 (5-11)	74 (51-117)
Laboratory	2011	B2-1	9 (6-13)	68 (45-127)
Laboratory	2012	B2-1	7 (5-10)	62 (41-107)
Laboratory	2013	B2-1	7 (5-10)	48 (31-88)
Laboratory	2013	B2-2	5 (3-9)	42 (26-87)
Laboratory	2014	B2-2	17 (11-25)	91 (57-209)
Laboratory	2015	B2-2	28 (21-36)	67 (50-110)
Laboratory	2016	B2-3	30 (24-38)	83 (62-132)
Laboratory	2017	B2-4	24 (15-35)	162 (100-363)
Laboratory	2018	B2-6	19 (13-26)	116 (76-224)
Laboratory	2019	B2-7	27 (16-40)	233 (133-656)
Laboratory	2020	B2-8	14 (10-19)	93 (59-180)
Laboratory	2021	B2-9	25 (14-40)	292 (139-1336)

[a] 50% and 90% moulting inhibition concentration (MIC_{50} and MIC_{90}) and their 95% confidence intervals (CI 95%) are expressed in ng Cry1Ab/cm^2.

7. Tables and figures

Table 7. Individuals of *S. nonagrioides* lost in the process of rearing or discarded for susceptibility bioassays. Percentages are shown with respect to the number of field larvae collected in each zone.

Fields	Field larvae collected	Dead larvae and pupae	Adults not used for mating[a]
Zone 1	542	167 (30.8%)	9 (1.7%)
Zone 2	553	173 (31.3%)	21 (3.8%)
Zone 3	604	191 (31.6%)	21 (3.5%)
All zones	1699	531 (31.3%)	51 (3.0%)

[a] Adults that did not emerge between 10th November 2021 and 10th February 2022, and adults having some malformation upon emergence.

7. Tables and figures

Table 8. Tracking of *S. nonagrioides* from the NE Spain populations and from the laboratory population used in the diagnostic concentration (DC) bioassays, and outcomes of the bioassays.

	Tracking of the larvae used in the DC bioassays							Results of DC bioassays				
Population	**Fields**	**Last-instar larvae collected**	**Adults emerged[a]**	**Adults mated[b]**	**Oviposition cages**	**Oviposition cages used in bioassays[c]**	**Adults used in bioassays[d]**	**Nº larvae treated in bioassays**	**MI (%)[e]**	**Nº larvae control**	**MI in control (%)[e]**	**Corrected MI (%)[f]**
NE Spain	Zone 1	542	375 (69%)	366 (68%)	35	33	354 (65%) (94%)	1178	99.66	198	5.05	99.64
	Zone 2	553	380 (69%)	359 (65%)	33	30	340 (61%) (89%)	1141	99.04	125	13.60	98.88
	Zone 3	604	413 (68%)	392 (65%)	35	34	382 (63%) (92%)	1148	96.60	150	8.67	96.28
	All zones[g]	1699	1168 (69%)	1117 (66%)	103	97	1076 (63%) (92%)	3467	98.44	473	8.46	98.30
Laboratory	-	-	-	479	26	26	479	1152	99.31	112	13.39	99.20

[a] The percentage with respect to the number of larvae collected is in brackets
[b] Adults that mated, after excluding those that did not emerge between 10th November 2021 and 10th February 2022, and those that presented malformations at the time of their emergence. For field populations, the percentage with respect to the number of larvae collected is in brackets.
[c] Oviposition cages were discarded when or when the fecundity and/or fertility was too low.
[d] Adults used in the bioassays, after excluding those that laid infertile eggs. For field populations, percentages with respect to the number of collected larvae and with respect to the number of emerged adults are in brackets (in this order).
[e] MI, moulting inhibition: larvae that have not reached the 2nd larval instar.
[f] Calculated using Abbot's formula (Abbot, 1925).
[g] Results obtained pooling the data of the three zones.

Table 9. Moulting inhibition values of F1 neonates of the NE Spain population compared with those of the laboratory population and with the expected value of 99%.

Year	Moulting inhibition at DC (%)			*p*-values [b]	
	NE Spain[a]	Lab strain[a]	Expected	Lab strain	Expected
2016	97.96 ± 0.71	99.20	99	0.066	0.107
2017	94.14 ± 1.40	97.69	99	**0.038***	**0.011***
2018	98.65 ± 0.40	97.75	99	0.081	0.253
2019	97.97 ± 0.36	97.02	99	0.067	**0.029***
2020	98.31 ± 0.39	98.67	99	0.291	0.113
2021	98.27 ± 1.02	99.20	99	0.355	0.429

[a] Moulting inhibition values reported 7 days after treatment with a diagnostic concentration (DC) of 1091 ng Cry1Ab/cm^2.

[b] *p*-values of one-sample t-test analyses performed to compare the percentage of moulting inhibition of the field population (NE Spain) with respect to the observed moulting inhibition obtained with the susceptible laboratory strain and with the expected theoretical value of 99%. Moulting inhibition values were previously logit transformed.

7. Tables and figures

Table 10. Larval growth of neonates of the F1 generation of *S. nonagrioides* after 10 days feeding on Bt (MON 810) or non-Bt (conventional) maize tissue.

Population	Fields	Nº of F0 oviposition cages used[a]	Maize leaves	Nº of F1 neonates exposed[b]	Nº of moulted larvae (≥ L2)	% moulting
NE Spain	Zone 1	33	MON 810	6450	0	0.00
			Conventional	330	325	98.48
	Zone 2	30	MON 810	5850	0	0.00
			Conventional	300	293	97.67
	Zone 3	34	MON 810	6650	0	0.00
			Conventional	340	337	99.12
	All zones	97	MON 810	18950	0	0.00
			Conventional	970	955	98.45
Laboratory	-	26	MON 810	5200	0	0.00
			Conventional	260	253	97.31

[a] F0 is the generation collected in the field.
[b] F1 neonates were < 24 h.

7. Tables and figures

Table 11. Confirmatory bioassays

11a. Larvae that were able to moult to the 2nd larval instar (L2) in the DC bioassay and then moulted again to the 3rd larval instar (L3) when fed MON 810 maize leaves.

Population	Fields	Nº larvae treated in DC bioassays	L2 (%)[a]	L3 (%)[b]	L4 (%)[b]
NE Spain	Zone 1	1178	4 (0.34)	0 (0.00)	0 (0.00)
	Zone 2	1141	11 (0.96)	1 (0.09)	0 (0.00)
	Zone 3	1148	39 (3.4)	0 (0.00)	0 (0.00)
	All zones	3467	54 (1.56)	1 (0.03)	0 (0.00)
Laboratory	-	1152	8 (0.69)	0 (0.00)	0 (0.00)

[a] Number of larvae that moulted to L2 in the DC bioassay, and then were fed on MON 810 maize. Percentages with respect to the number of treated larvae.
[b] Number of larvae that moulted to L3 and L4 after feeding on MON 810. Percentages with respect to the number of treated larvae.

11b. Larval growth of neonates of the F2 generation of *S. nonagrioides* after 10 days feeding on Bt (MON 810) or non-Bt (conventional) maize tissue.

Population	Field	Nº of F1 oviposition cages used[a]	Maize leaves	Nº of F2 neonates exposed[b]	Nº of moulted larvae (≥ L2)	% moulting
NE Spain	Zone 2	6	MON 810	1200	0	0.00
			Conventional	60	58	96.67

[a] F2 neonates were < 24 h.

7. Tables and figures

Figure 1. Successful field collections of *S. nonagrioides* and *O. nubilalis* in three different zones (Z1, Z2 and Z3) searched in the NE Spain in 2020. A collection at a field within a zone was considered successful if at least 50 larvae were gathered.

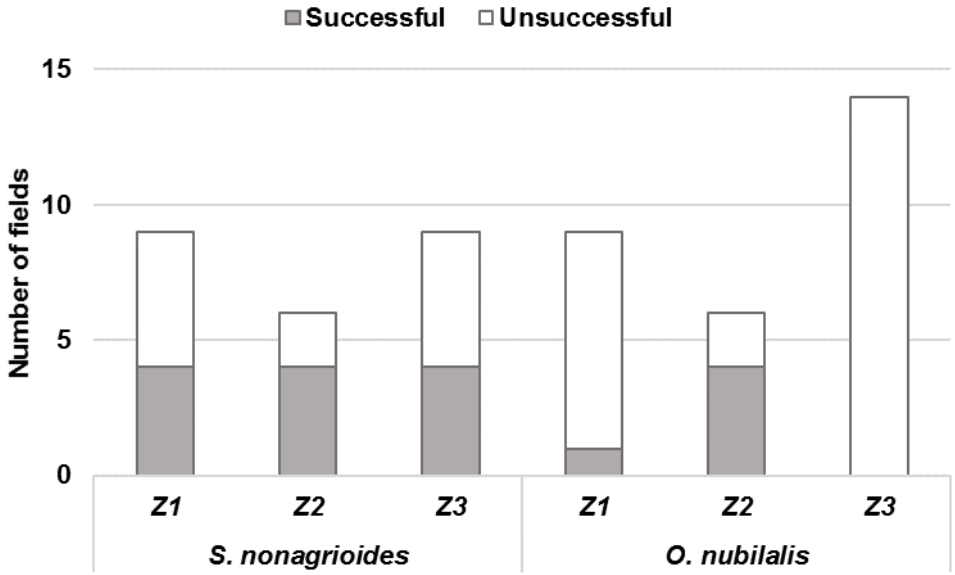


Figure 2. Fitted curves of susceptibility to the toxin Cry1Ab of the laboratory populations of *S. nonagrioides* and *O. nubilalis* (PoloPlus 1.0, LeOra Software 2002-2022). Response is moulting inhibition after seven days feeding on treated diet. a: *S. nonagrioides*. b: *O. nubilalis*.

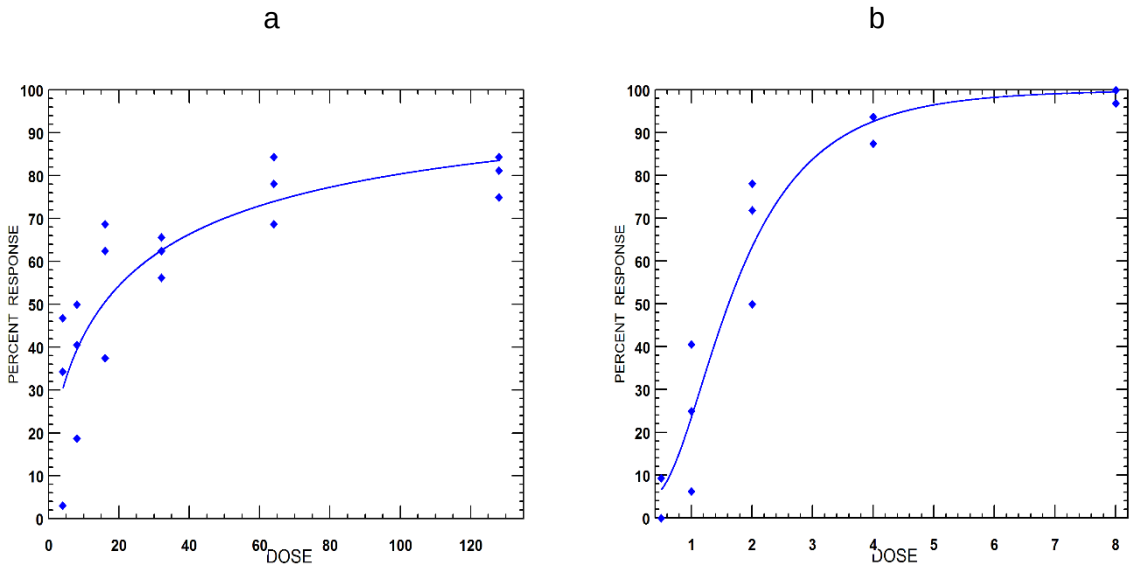


Figure 3. Susceptibility to Cry1Ab toxin measured by MIC_{50} values of a laboratory population of *O. nubilalis.* Colours indicate the B1 (blue) and B2 (pink) toxin batches.

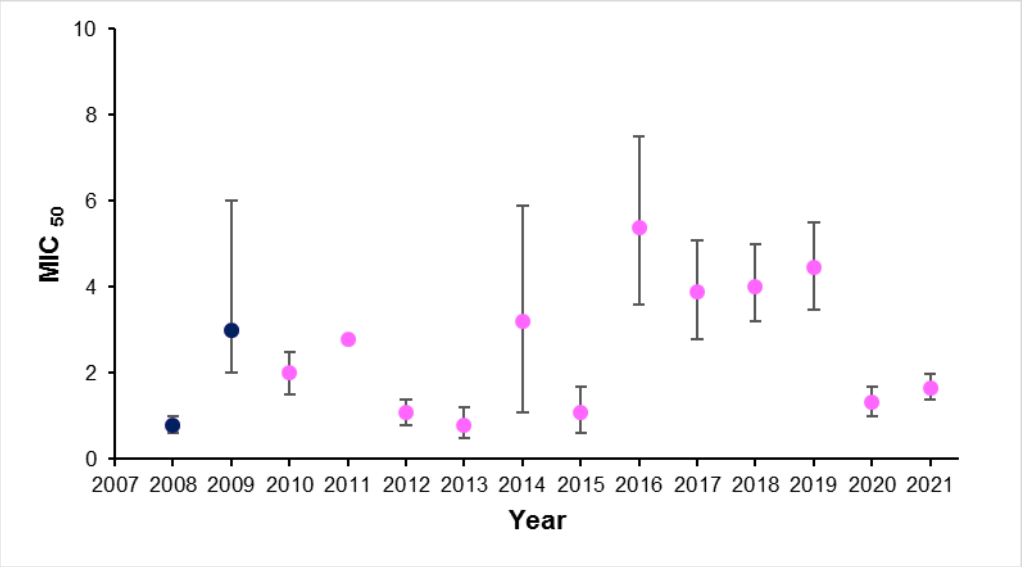
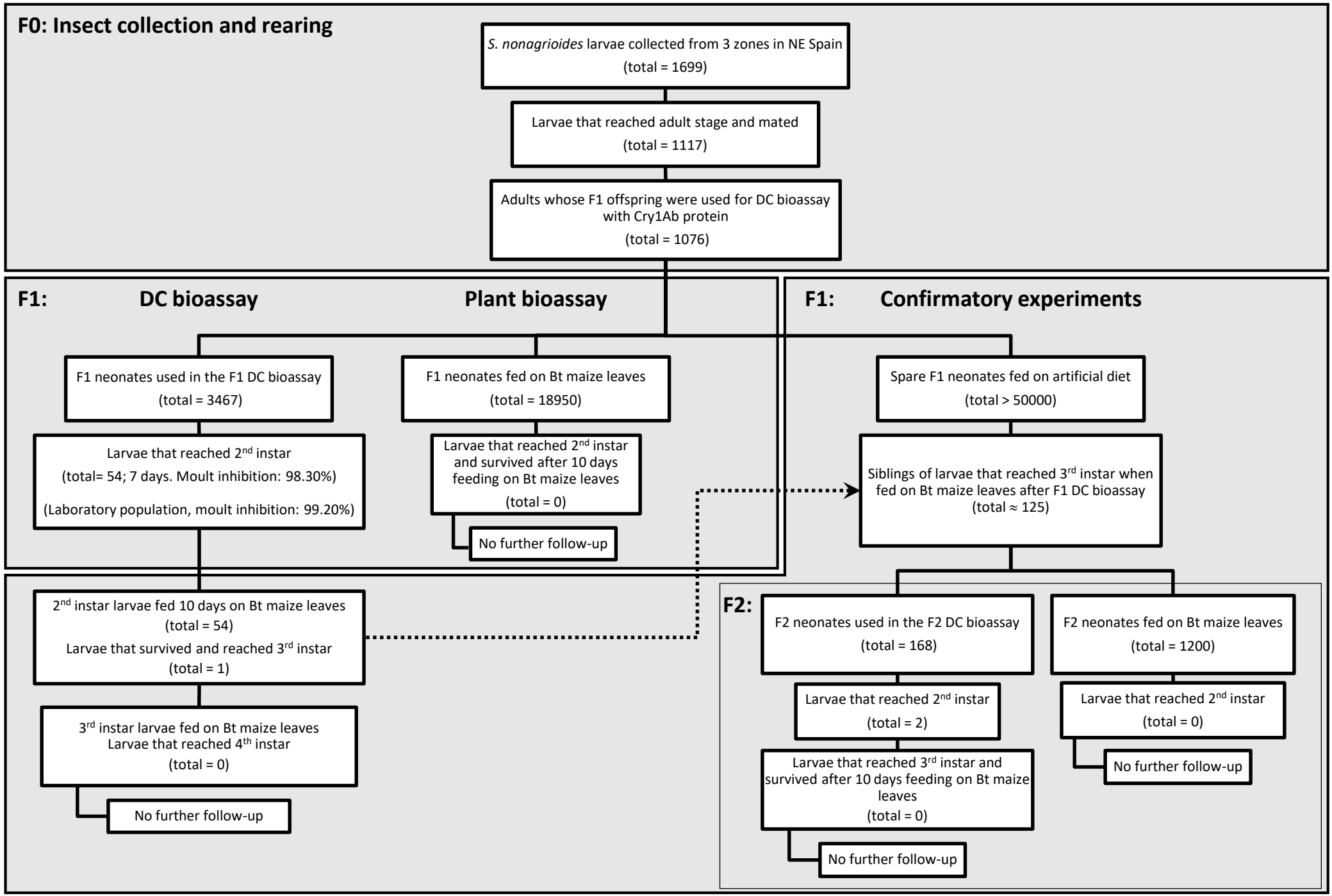


Figure 4. Moulting inhibition of neonates of *S. nonagrioides* from three zones of NE Spain (mean ± SE) and from the laboratory population, treated with a diagnostic concentration (DC) of 1091 ng/cm^2 bioassays. The dotted black line represents the expected 99% moult inhibition (MI) value.

Annex I. Stepwise approach followed to do the bioassays with *Sesamia nonagrioides*

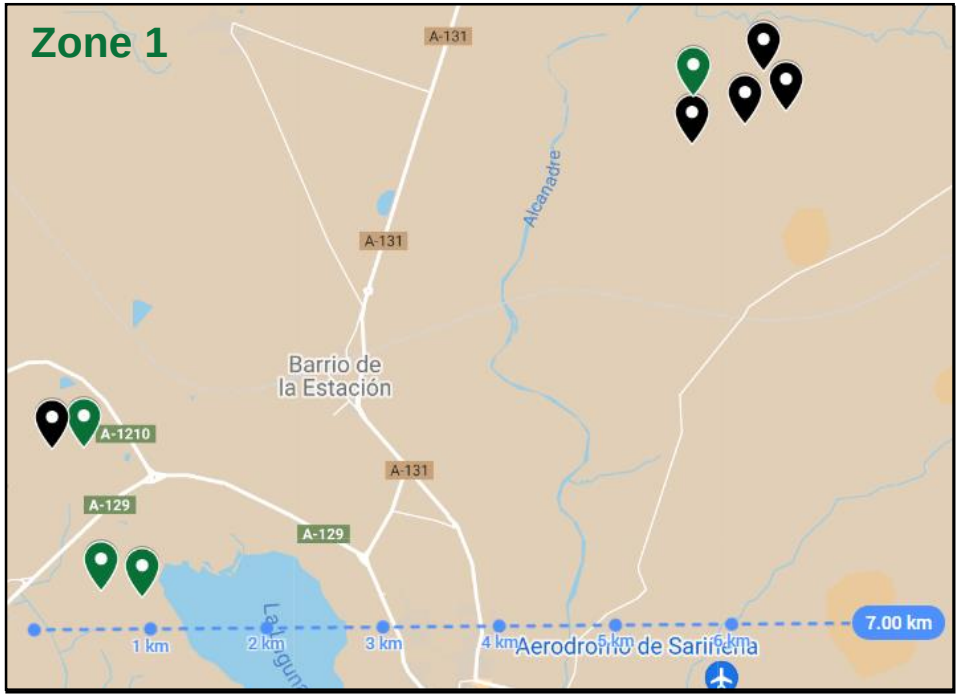


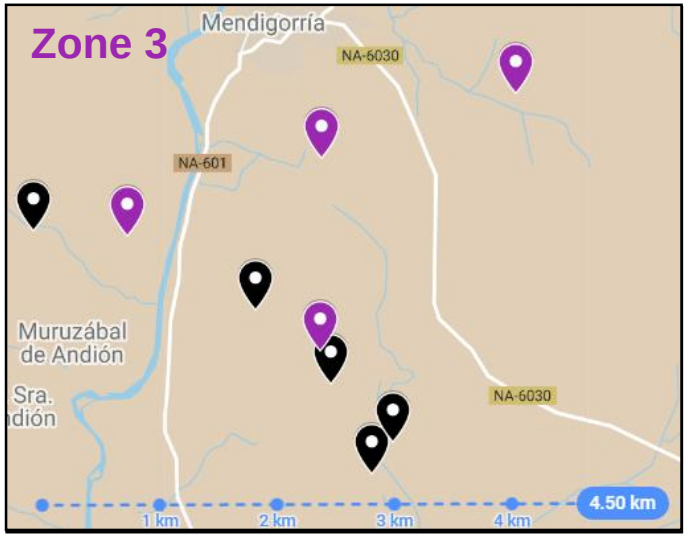
ANNEX IIa. Collection of *S. nonagrioides* larvae in NE Spain in 2021

Successful sampling site: at least 50 larvae were gathered.

Unsuccessful sampling site: less than 50 or no larvae were gathered.

ANNEX IIb. Collection of *S. nonagrioides* larvae in NE Spain in 2021





Successful sampling sites-Zone 1

Successful sampling sites-Zone 2

Successful sampling sites-Zone 3

Unsuccessful sampling sites

ANNEX IIIa. Collection of *O. nubilalis* larvae in NE Spain in 2021

Successful sampling site: at least 50 larvae were gathered.

Unsuccessful sampling site: less than 50 or no larvae were gathered.

ANNEX IIIb. Collection of *O. nubilalis* larvae in NE Spain in 2021

Successful sampling sites-Zone 1

Successful sampling sites-Zone 2

Successful sampling sites-Zone 3

Unsuccessful sampling sites